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(54) Title: ION CHANNEL

(57) Abstract

The present invention relates to a novel 1,957 amino acid tetrodotoxin-insensitive voltage-gated sodium channel specifically located in mammalian sensory neurons. Nucleic acid sequences coding for the novel sodium channel, vectors, host cells and methods of identifying modulators of the novel sodium channel for use in treatment of pain are also provided.

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ION CHANNEL

Voltage-gated sodium channels are transmembrane proteins which cause sodium permeability to increase. Depolarization of the plasma membrane causes sodium channels to open allowing sodium ions to enter along the electrochemical gradient creating an action potential.

Voltage-gated sodium channels are expressed by all electrically excitable cells, where they play an essential role in action potential propagation. They comprise a major subunit of about 2000 amino acids which is divided into four domains (D1-D4), each of which contains 6 membrane-spanning regions (S1-S6). The alpha-subunit is usually associated with 2 smaller subunits (beta-1 and beta-2) that influence the gating kinetics of the channel. These channels show remarkable ion selectivity, with little permeability to other monovalent or divalent cations. Patch-clamp studies have shown that depolarisation leads to activation with a typical conductance of about 20pS, reflecting ion movement at the rate of 10⁷ ions/second/channel. The channel inactivates within milliseconds (Caterall, W.A., Physiol. Rev. 72, S4-S47 (1992); Omri et al, J. Membrane Biol 115, 13-29; Hille, B, Ionic Channels in Excitable Membranes, Sinauer, Sunderland, MA (1991)).

Sodium channels have been pharmacologically characterised using toxins which bind to distinct sites on sodium channels. The heterocyclic guanidine-based channel blockers tetrodotoxin (TTX) and saxitoxin (STX) bind to a site in the S5-S6 loop, whilst μ -conotoxin binds to an adjacent overlapping region. A number of toxins from sea anemones or scorpions binding at other sites alter the voltage-dependence of activation or inactivation.

Voltage-gated sodium channels that are blocked by nanomolar concentrations of tetrodotoxin are known as tetrodotoxin sensitive sodium channels (Hille (1991) "Ionic Channels in Excitable Membranes", Sinauer Sunderland, MA (1991)) whilst sodium channels that are blocked by concentrations greater than 1 micromolar are known as tetrodotoxin-insensitive (TTXi) sodium channels (Pearce and Duchen Neuroscience 63, 1041-1056 (1994)).

Dorsal root ganglion (DRG) neurons express at least three types of sodium channels which differ in kinetics and sensitivity to TTX. Neurons with small-diameter cell bodies and unmyelinated axons (C-fibers) include most of the nociceptor (damage-sensing)

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population and express a fast TTX-sensitive current and a slower TTX-insensitive current. Of the five cloned sodium channel α -subunit transcripts known to be present in dorsal root ganglia, none exhibits the properties of the TTX-insensitive channel.

Sodium channel blockers are used clinically to provide pain relief. Three classes of sodium channel blockers in common clinical use are: local anesthetics such as lidocaine, some anticonvulsants such as phenytoin and carbamazepine, and some antiarrhythmics such as mexiletine. Each of these is known to suppress ectopic peripheral nervous system discharge in experimental preparations and to provide relief in a broad range of clinical neuropathic conditions.

Applicants have now found a novel voltage-gated sodium channel (hereinafter referred to as a sodium channel specifically located in sensory neurons or also referred to as SNS sodium channel) that is present in sensory neurons (or neurones) but not present in glia, muscle, or the neurons of the sympathetic, parasympathetic, enteric or central nervous systems. Preferably the sodium channel of the invention is found in the neurons of the dorsal root ganglia (DRG) or cranial ganglia. More preferably the sodium channel of the invention is found in the neurons of the dorsal root ganglia. Preferably the sodium channel is specifically located in rat sensory neurons or human sensory neurons.

The sodium channel of the present invention is believed to play a role in nociceptive transmission because some noxious input to the central nervous system is known to be insensitive to TTX. Persistent activation of peripheral nociceptors has been found to result in changes in excitability in the dorsal horn associated with the establishment of chronic pain. Increased sodium channel activity has also been shown to underlie neuroma-induced spontaneous action potential generation. Conversely, chronic pain may be successfully treated by surgical or pharmacological procedures which block peripheral nerve activation. Blockage of nociceptor input may therefore produce useful therapeutic effects, even though central nervous system plasticity plays a pivotal role in the establishment of chronic pain. Sensory neuron-specific voltage-gated sodium channels, particularly sub-types associated with a nociceptive modality such as the sodium channel of the invention, thus provide targets for therapeutic intervention in a range of pain states. The electrophysiological and pharmacological properties of the expressed SNS sodium channel are similar to those described for the small diameter sensory neuron tetrodotoxin-

resistant sodium channels. As some noxious input into the spinal cord is resistant to

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tetrodotoxin, block of expression or function of such a C-fiber-restricted sodium channel may have a selective analgesic effect.

In another aspect the present invention provides an isolated protein comprising a sodium channel specifically located in rat sensory neurons as encoded by the insert deposited in NCIMB deposit number 40744, which was deposited at The National Collections of Industrial and Marine Bacteria, 23 St Machar Drive, Aberdeen AB2 1RY, Scotland, United Kingdom on 27 June 1995 in accordance with the Budapest Treaty.

The invention also provides nucleotide sequences coding for the SNS sodium channel. In a preferred embodiment, the nucleotide sequence encodes a sodium channel specifically located in rat sensory neurons which is as set out in Figure 1a or a complementary strand thereof.

The approximately 6.5 kilobase (kb) transcript expressed selectively in rat dorsal root ganglia that codes for the novel sodium channel of the invention shows sequence similarities with known voltage-gated sodium channels. The cDNA codes for a 1,957 amino acid protein. In particular, the novel sodium channel of the invention shows 65% identity at the amino acid level with the rat cardiac tetrodotoxin-insensitive (TTXi) sodium channel. The aromatic residue that is involved in high-affinity binding of TTX to the channel atrium of TTX-sensitive sodium channels is altered to a hydrophilic serine in the predicted protein of the SNS sodium channel, whereas the residues implicated in sodium-selective permeability are conserved. The novel sodium channel specifically located in sensory neurons shows relative insensitivity to TTX (IC50>1 micromolar) and thus exhibits properties different from other cloned sodium channel transcripts known to be present in dorsal root ganglia.

The invention also provides expression and cloning vectors comprising a nucleotide sequence as hereinabove defined. In order to effect transformation, DNA sequences containing the desired coding sequence and control sequences in operable linkage (so that hosts transformed with these sequences are capable of producing the encoded proteins) may be included in a vector, however, the relevant DNA may then also – be integrated into the host chromosome.

The invention also provides a screening assay for modulators of the sodium channel which is specifically located in sensory neurons wherein the assay comprises

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adding a potential modulator to a cell expressing the SNS sodium channel and detecting any change in activity of the sodium channel.

The present invention also provides a modulator which has activity in the screening assay hereinabove defined. Modulators of the sodium channel as hereinabove defined are useful in modulating the sensation of pain. Blockers of the sodium channel will block or prevent the trasmission of impulses along sensory neurons and thereby be useful in the treatment of acute, chronic or neuropathic pain.

The present invention thus relates to novel voltage-gated sodium channel proteins specific to sensory neurons, to nucleotide sequences capable of encoding these sodium channel proteins, to vectors comprising a nucleotide sequence coding for a sodium channel of the invention, to host cells containing these vectors, to cells transformed with a nucleic acid sequence coding for the sodium channel, to screening assays using the sodium channel proteins and/or host cells, to complementary stands of the DNA sequence which is capable of encoding the sodium channel proteins and to antibodies specific for the sodium channel proteins. These and other aspects of the present invention are set forth in the following detailed description.

Brief Description of the Drawings:

Figure 1a shows the nucleic acid and amino acid sequences of the sodium channel specific to the rat DRG (SNS-B) (SEQ ID NO: 1 and SEQ ID NO: 2).

Figure 1b shows the structure of the SNS-B voltage-gated sodium channel in pGEM-3Z.

Figure 1c shows a schematised drawing of a known voltage-gated sodium channel.

Figure 2 shows sequences of examples of PCR primers for isolation of human clone probes. RLLRVFKLAKSWPTL - SEQ ID NO: 21; 5' gettgetgegggtetteaage 3' SEQ ID NO: 22; LRALPLRALSRFEG - SEQ ID NO: 23; 5' ategagacagagecegeageg 3' SEQ ID NO: 24; 5' acgggtgeegeaaggaeggegteteegtgtggaaeggegagaag 3' SEQ ID NO: 25; and 5' ggetateetteeteteeageteteaeceaggtatggagecaggt 3' - SEQ ID NO: 26.

Figure 3 shows a film of ³⁵S radio-labelled SNS-B voltage-gated sodium channel protein in a coupled transcription/translation system.

Figure 4a and Figure 4b show SNS-GST fusion protein constructs for antibody generation. TCCCGTACGCTGCAGCTCTTT - SEQ ID NO: 27; CCCGGGGAAGGCTAC - SEQ ID NO: 28: GTCGACACCAGAAAT - SEQ ID NO: 29; GGATCCTCTAGAGTCGACCTGCAGAAGGAA - SEQ ID NO: 30

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In accordance with one aspect of the invention there is provided an isolated and/or purified nucleic acid sequence (or polynucleotide or nucleotide sequence) which comprises a nucleic acid sequence which encodes the mammalian sodium channel specifically located in sensory neurons or a complementary strand thereof. Preferably, the nucleic acid sequence encodes the sodium channel specifically located in mammalian dorsal root ganglia. More preferably, the nucleic acid sequence encodes the rat or human sodium channel specifically located in dorsal root ganglia. The rat nucleic acid sequence preferably comprises the sequence of the coding portion of the nucleic acid sequence shown in Figure 1a (SEQ ID NO:1) or the coding portion of the cDNA deposited in NCIMB deposit number 40744 which was deposited at the National Collections of Industrial and Marine Bacteria, 23 St. Machar Drive, Aberdeen AB21RY, Scotland, United Kingdom on June 27, 1995 in accordance with the Budapest Treaty.

A nucleic acid sequence encoding a sodium channel of the present invention may be obtained from a cDNA libraray derived from mammalian sensory neurons, preferably dorsal root ganglia, trigeminal ganglia or other cranial ganglia, more preferably rat or human dorsal root ganglia. The nucleotide sequence described herein was isolated from a cDNA library derived from rat dorsal root ganglia cells. The nucleic acid sequence coding for the SNS sodium channel has an open reading frame of 5,871 nucleotides encoding a 1,957 amino acid protein. A nucleic acid sequence encoding a sodium channel of the present invention may also be obtained from a mammalian genomic library, preferably a human or rat genomic library. The nucleic acid sequence may be isolated by the subtraction hybridization method described in the examples, by screening with a probe derived from the rat sodium channel sequence, or by other methodologies known in the art such as polymerase chain reaction (PCR) with appropriate primers derived from the rat sodium channel sequence and/or relatively conserved regions of known voltage-gated sodium channels.

The nucleic acid sequences of the present invention may be in the form of RNA or in the form of DNA, which DNA includes cDNA, genomic DNA, and synthetic

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DNA. The DNA may be double-stranded or single-stranded, and if single stranded may be the coding strand or non-coding (anti-sense) strand. The coding sequence which encodes the rat SNS sodium channel or variant thereof may be identical to the coding sequences set forth herein or that of the deposited clone, or may be a different coding sequence which coding sequence, as a result of the redundancy or degeneracy of the genetic code, encodes the same protein as the sequences set forth herein or the deposited cDNA.

The nucleic acid sequence which encodes the SNS sodium channel may include: only the coding sequence for the full length protein or any variant thereof; the coding sequence for the full length protein or any variant thereof and additional coding sequence such as a leader or secretory sequence or a proprotein sequence; the coding sequence for the full length protein or any variant thereof (and optionally additional coding sequence) and non-coding sequences, such as introns or non-coding sequences 5' and/or 3' of the coding sequence for the full length protein.

The present invention further relates to variants of the hereinabove described nucleic acid sequences which encode fragments, analogs, derivatives or splice variants of the SNS sodium channel. The variant of the SNS sodium channel may be a naturally occurring allelic variant of the SNS sodium channel. As known in the art, an allelic variant is an alternate form of a protein sequence which may have a substitution, deletion or addition of one or more nucleotides, which does not substantially alter the function of the encoded protein. The present invention relates to splice variants of the SNS sodium channel that occur physiologically and which may play a role in changing the activation threshold of the sodium channel.

Variants of the sequence coding for the rat SNS sodium channel have been identified and are listed below:

- 1) a 2573 base pair nucleic acid sequence shown in SEQ ID NO:3. This sequence codes for a 521 amino acid protein that corresponds to amino acids 1437-1957 of Figure 1a (SEQ ID NO:1) and has the same sequence as bases 4512 through 6524 of Figure 1a in the coding portion and 3' untranslated region.
- a 7052 base pair nucleic acid sequence shown in SEQ ID NO: 5. SEQ
 ID NO: 6 codes for a 2.132 amino acid protein that contains a 176 amino acid repeat
 (amino acids 586-760 of SEQ ID NO:6) inserted after amino acid 585 in Figure 1a or SEQ
 ID NO:2.

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A preferred sequence for the rat SNS sodium channel is shown in Figure 1a (SEQ ID NO: 1). However, sequencing variations have been noted. Sequencing has provided

a 6,321 base pair nucleic acid sequence coding for a 1957 amino acid protein that has the same base sequence as bases 1-6321 of Figure 1a or SEQ ID NO:1 with the following changes: bases 1092 G to A, base 1096 C to T, base 2986 G to T, base 3525 C to G and base 3556 G to C.

a 6,527 base pair nucleic acid sequence coding for a 1,957 amino acid protein as shown in SEQ ID NO:7 that has the same base sequence as bases 1-6524 of Figure 1a (SEQ ID NO:1) with an additional 3 bases AAA, at the 3' end, and the following changes: base 299 C to G, base 1092 G to A, base 1096 C to T, base 1964 G to C, base 1965 C to G, base 2472 A to T, base 2986 G to T, base 3019 A to G, base 3158 C to T, base 3525 C to G, base 3556 G to C and base 5893 T to G. The sequence of SEQ ID NO: 7 is also a preferred sequence coding for the rat SNS sodium channel.

a 6524 base pair nucleic acid sequence that has the same sequence as Figure 1a (SEQ ID NO: 1) except for the following base changes: base 1092 G to A (resulting in a change at amino acid 297 of SEQ ID NO: 2 from Val to Ile), base 1096 C to T (resulting in a change at amino acid 298 from Ser to Phe), base 1498 C to A (resulting in a change at amino acid 432 from Ala to Glu), and base 2986 G to T (resulting in a change at amino acid 928 form Ser to Ile).

Sequence variability has been identified in different isolates. One such sequence has been identified that has the sequence of the third sequencing variation shown immediately above except for eight base differences, five of which resulted in an altered amino acid sequence F16-S16, L393-P393, T470-I470, R278-H278, and I1,876-M1,876.

The present invention also relates to nucleic acid probes constructed from the nucleic acid sequences of the invention or portion thereof. Such probes could be utilized to screen a dorsal root ganglia cDNA library to isolate a nucleic acid sequence encoding the sodium channel of the present invention. The nucleic acid probes can include portions of the nucleic acid sequence of the SNS sodium channel or variant thereof useful for hybridizing with mRNA or DNA for use in assays to detect expression of the SNS

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sodium channel or localize its presence on a chromosome, such as the *in situ* hybridization assay described herein.

A conservative analogue is a protein sequence which retains substantially the same biological properties of the sodium channel but differs in sequences by one or more conservative amino acid substitutions. For the purposes of this document a conservative amino acid substitution is a substitution whose probability of occuring in nature is greater than ten times the probability of that substitution occuring by chance (as defined by the computational methods described by Dayhoff et al, Atlas of Proteins Sequence and Structure, 1971, page 95-96 and figure 9-10).

A splice variant is a protein product of the same gene, generated by alternative splicing of mRNA, that contains additions or deletions within the coding region (Lewin B. (1995) Genes V Oxford University Press, Oxford, England)

The nucleic acid sequences of the present invention may also have the coding sequence fused in frame to a marker sequence which allows for purification of the protein of the present invention such as a hexa-histidine tag or a hemagglutinin (HA) tag.

The present invention further relates to nucleic acid sequences which hybridize to the hereinabove-described sequences if there is at least 50% and preferably 70% identity between the sequences. The present invention particularly relates to nucleic acid sequences which hybridize under stringent conditions to the hereinabove-described nucleic acid sequences. As herein used, the term "stringent conditions" means hybridization will occur only if there is at least 95% and preferably at least 97% identity between the sequences preferably the nucleic acid sequences which hybridize to the hereinabove described nucleic acid sequences encode proteins which retain substantially the same biological function or activity as the SNS sodium channel, however, nucleic acid sequences that have different properties are also within the scope of the present invention. Such sequences, while hybridizing with the above described nucleic acid sequences may encode a protein having diffferent properties, such as sensitivity to tetrodotoxin which property is found in the altered SNS sodium channel protein described herein.

In accordance with another aspect of the invention there is provided purified mammalian sensory neuron sodium channel protein, wherein the sodium channel is insensitive to tetrodotoxin. Preferably the sodium channel of the invention is found in the neurons of the dorsal root ganglia or cranial ganglia, more preferably the neurons of the

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dorsal root ganglia. The sodium channel protein may be derived from any mammalian species, preferably the rat or human sodium channel protein. The rat SNS sodium channel protein preferably has the deduced amino acid sequence shown in Figure 1a (SEQ ID NO:2) or SEQ ID NO: 8, or the amino acid sequence encoded by the deposited cDNA.

Fragments, analogues, derivatives, and splice variants of the sodium channel specifically located in sensory neurons are also within the scope of the present invention.

The terms "fragment," "derivative" and "analogue" when referring to the DRG sodium channel of the invention refers to a protein which retains substantially the same biological function or activity as such protein. Thus, an analogue includes a proprotein which can be activated by cleavage of the proprotein portion to produce an active mature protein. In addition, the present invention also includes derivatives wherein the biological function or activity of the protein is significantly altered, including derivatives that are sensitive to tetrodotoxin.

The protein of the present invention may be a recombinant protein, a natural protein or a synthetic protein, preferably a recombinant protein.

The fragment, derivative or analog of the SNS sodium channel protein includes, but is not limited to, (i) one in which one or more of the amino acid residues are substituted with a conserved or non-conserved amino acid residue (preferably a conserved amino acid residue) and such substituted amino acid residue may or may not be one encoded by the genetic code, or (ii) one in which one or more of the amino acid residues includes a substituted group, or (iii) one in which the mature polypeptide is fused with another compound, such as a compound to increase the half-life of the protein (for example, polyethylene glycol), or (iv) one in which the additional amino acids are fused to the mature protein, such as a leader or secretory sequence or a sequence which is employed for purification of the mature protein or a proprotein sequence, or (v) one in which one or more amino acids has/have been deleted so that the protein is shorter than the full length protein. Variants of the rat SNS sodium channel are discussed hereinabove and shown in SEQ ID NO:4 and SEQ ID NO:6.

The proteins and nucleic acid sequences of the present invention are

preferably provided in an isolated form, and pre rably are purified to at least 50% purity,
more preferably about 75% purity, most preferably about 90% purity.

The terms "isolated" and/or "purified" mean that the material is removed from is original environment (e.g., the natural environment if it is naturally occurring). For example, a naturally-occurring nucleic acid sequence or protein present in a living animal is not isolated or purified, but the same nucleic acid sequence or DNA or protein, separated from some or all of the coexisting materials in the natural system, is isolated or purified. Such nucleic acid sequence could be part of a vector and/or such nucleic acid sequence or protein could be part of a composition, and still be isolated or purified in that such vector or composition is not part of its natural environment.

The present invention also provides vectors comprising a nucleic acid sequence of the present invention, and host cells transformed or transfected with a nucleic of the invention.

The nucleic acid sequences of the present invention may be employed for producing the SNS sodium channel protein or variant thereof by recombinant techniques. Thus, for example, the nucleic acid sequence may be included in any one of a variety of expression vehicles or cloning vehicles, in particular vectors or plasmids for expressing a protein. Such vectors include chromosomal, nonchromosomal and synthetic DNA sequences. Examples of suitable vectors include derivatives of SV40; bacterial plasmids; phage DNA; yeast plasmids; vectors derived from combinations of plasmids and phage DNA, viral DNA such as vaccinia, adenovirus, fowl pox virus, pseudorabies and baculovirus. However, any other plasmid or vector may be used as long as it is replicable and viable in the host.

More particularly, the present invention also provides recombinant constructs comprising one or more of the nucleic acid sequences as broadly described above. The constructs comprise an expression vector, such as a plasmid or viral vector, into which a sequence of the invention has been inserted, in a forward or reverse orientation. In a preferred aspect of this embodiment, the construct further comprises one or more regulatory sequences, including, for example, a promoter, operably linked to the sequence. Large numbers of suitable vectors and promoters are known to those of skill in the art, and are commercially available. The following vectors are provided by way of example. Bacterial: pQE70, pQE60, pQE-9 (Qiagen) pBs, phagescript, psiX174, pBluescript SK, pBsKS, pNH8a, pNH16a, pNH18a, pNH461 (Stratagene); pTrc99A, pKK223-3, pKK233-3, pDR540, pRIT5 (Pharmacia). Eukaryotic: pWLneo, pSV2cat.

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pOG44, pXT1, pSG (Stratagene), pSVK3, pBPV, pMSG, pSVL (Pharmacia) pcDNA 3.1 (Invitrogen, San Diego, CA), pEE14 (WO 87/04462) and pREP8 (Invitrogen). Preferred vectors include pcDNA 3.1, pEE14 and pREP8. However, any other plasmid or vector may be used as long as it is replicable and viable in the host.

As hereinabove indicated, the appropriate DNA sequence may be inserted into the vector by a variety of procedures. In general, the DNA sequence is inserted into appropriate restriction endonuclease sites by procedures known in the art. Such procedures and others are deemed to be within the scope of those skilled in the art.

The DNA sequence in the expression vector is operatively linked to an appropriate expression control sequence(s) (promoter) to direct mRNA synthesis. As representative examples of such promoters, there may be mentioned: LTR or SV40 promoter and other promoters known to control expression of genes in prokaryotic or eukaryotic cells or their viruses. The expression vector may contain a ribosome binding site for translation initiation and transcription terminator. The vector may also include appropriate sequences for amplifying expression.

Promoter regions can be selected from any desired gene using CAT (chloramphenicol transferase) vectors or other vectors with selectable markers. Two appropriate vectors are pKK232-8 and pCM7. Particular named bacterial promoters include LacI, LacZ, T3, T7, gpt, lambda P_R, P_L and trp. Eukaryotic promoters include CMV immediate early, HSV thymidine kinase, early and late SV40. LTRs from retrovirus, and mouse metallothionein-I. Selection of the appropriate vector and promoter is well within the level of ordinary skill in the art.

Depending on the expression system employed in addition, the expression vectors preferably contain a gene to provide a phenotypic trait for selection of transformed host cells such as dihydrofolate reductase or neomycin resistance for eukaryotic cell culture, or such as tetracycline or ampicillin resistance in *E. coli*.

Transcription of DNA encoding the protein of the present invention by higher eukaryotes can be increased by inserting an enhancer sequence into the vector. Enhancers are cis-acting elements of DNA, usually about from 10 to 300 bp, that act on a promoter to increase its transcription. Examples include the SV40 enhancer on the late side of the replication origin (bp 100 to 270), a cytomegalovirus early promoter enhancer, a polyoma enhancer on the late side of the replication origin, and adenovirus enhancers.

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Useful expression vectors for bacterial use may be constructed by inserting a structural DNA sequence encoding a desired protein together with suitable translation initiation and termination signals in operable reading phase with a functional promoter. The vector will comprise one or more phenotypic selectable markers and an origin of replication to ensure maintenance of the vector and to, if desirable, provide amplification within the host. Suitable prokaryotic hosts for transformation include *E. coli, Bacillus subtilis, Salmonella typhimurium* and various species within the genera Pseydomonas, Streptomyces, and Staphylococcus, although others may also be employed as a matter of choice.

As a representative but nonlimiting example, useful expression vectors for bacterial use can comprise a selectable marker and bacterial origin of replication derived from commercially available plasmids comprising genetic elements of the well known cloning vector pBR322 (ATCC 37017). Such commercial vectors include, for example, PKK223-3 (Pharmacia Fine Chemicals, Uppsala, Sweden) and GEM1 (Promega Biotec, Madison, Wis., U.S.A.). These pBR322 "backbone" sections are combined with an appropriate promoter and the structural sequence to be expressed.

The sodium channel can be expressed in insect cells with the baculovirus expression system which uses baculovirus such as Autographa Californica nuclear polyhydrosis virus (AcNPV) to produce large amounts of protein in insect cells such as the Sf9 or 21 clonal lines derived from *Spodoptera frugiperda* cells. See for example O'Reilly et al., (1992) Baculovirus Expression Vectors: A Laboratory Manual, Oxford University Press.

Mammalian expression vectors will comprise an origin of replication, a suitable promoter and enhancer, and also any necessary ribosome binding sites, polyadenylation site, splice donor and acceptor sites, transcriptional termination sequences, and 5' flanking nontranscribed sequences. DNA sequences derived from the SV40 viral genome, for example, SV40 origin, early promoter, enhancer, splice, and polyadenylation sites may be used to provide the required nontranscribed genetic elements.

Mammalian expression vectors will comprise an origin of replication, a suitable promoter and enhancer, and also any necessary ribosome binding sites, polyadenylation site, splice donor and acceptor sites, transcriptional termination sequences, and 5' flanking nontranscribed sequences. DNA sequences derived from the SV40 viral

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genome, for example, SV40 origin, early promoter, enhancer, splice, and polyadenylation sites may be used to provide the required nontranscribed genetic elements.

In a further embodiment, the present invention provides host cells capable of expressing a nucleic acid sequence of the invention. The host cell can be, for example, a higher eukaryotic cell, such as a mammalian cell, a lower eukaryotic cell, such as a yeast cell, a prokaryotic cell, such as a bacterial cell. Introduction of the construct into the host cell may be effected by calcium phosphate transfection, DEAE-Dextran mediated transfection, electroporation (Davis, L., Dibner, M., Battey, I., Basic Methods in Molecular Biology, 1986) or any other method known in the art.

Host cells are genetically engineered (transduced, transformed or transfected) with the vectors of this invention which may be, for example, a cloning vector or an expression vector. The vector may be, for example, in the form of a plasmid, a viral particle, a phage, etc. The engineered host cells can be cultured in conventional nutrient media modified as appropriate for activating promoters, selecting transformants or amplifying the SNS sodium channel genes. The culture conditions, such as temperature, pH and the like, are those previously used with the host cell selected for expression, and will be apparent to the ordinarily skilled artisan.

The vector containing the appropriate DNA sequence as hereinabove described, as well as an appropriate promoter or control sequence, may be employed to transform an appropriate host to permit the host to express the protein. As representative examples of appropriate hosts, there may be mentioned: bacterial cells, such as *E. coli*, and *Salmonella typhimurium*: Streptomyces; fungal cells, such as yeast; insect cells such as Drosophila and *Spodoptera fugiperda* Sf9; animal cells such as CHO, COS or Bowes melanoma Ltk - and Y1 adrenal carcinoma; plant cells, etc. The selection of an appropriate host is deemed to be within the scope of those skilled in the art based on the teachings herein. Preferred host cells include mammalian cell lines such as CHO-K1, COS-7; Y1 adrenal; carcinoma cells. More preferably, the host cells are CHO-K1 cells. Preferred host cells for transient expression of the SNS sodium channel include *Xenopus laevis* oocytes.

The sodium channel may be transiently expressed in *Xeropus laevis* oocytes.

Cell-free translation systems can also be employed to produce such proteins using RNAs derived from the DNA constructs of the present invention. Appropriate cloning and

expression vectors for use with prokaryotic and eukaryotic hosts are described in Sambrook et al., Molecular Cloning: A Laboratory Manual, Second Edition, Cold Spring Harbor, N.Y., (1989).

Various mammalian cell culture systems can also be employed to express recombinant protein. Examples of mammalian expression systems include the COS-7 lines of monkey kidney fibroblasts, described by Gluzman, Cell. 23:175 (1981), and other cell lines capable of expressing a compatible vector, for example, the C127, 3T3, CHO, CHO-K1, HeLa, HEK 293, NIH 3T3 and BHK cell lines.

The constructs in host cells can be used in a conventional manner to produce the gene product encoded by the recombinant sequence. Alternatively, the proteins of the invention can be synthetically produced by conventional peptide synthesizers.

Cells are typically harvested by centrifugation, disrupted by physical or chemical means, and the resulting crude extract retained for further purification.

Microbial cells employed in expression of proteins can be disrupted by any convenient method, including freeze-thaw cycling, sonication, mechanical disruption, or use of cell lysing agents, such methods are well-known to those skilled in the art.

The SNS sodium channel protein is recovered and purified from recombinant cell cultures by methods known in the art, including ammonium sulfate or ethanol precipitation, acid extraction, anion or cation exchange chromatography, phosphocellulose chromatography, hydrophobic interaction chromatography, hydroxyapatite chromatography and lectin chromatography. Protein refolding steps may be used, as necessary, in completing configuration of the mature protein. Finally, high performance liquid chromatography (HPLC) can be employed for final purification steps.

The SNS sodium channel protein of the present invention may be naturally purified products expressed from a high expressing cell line, or a product of chemical synthetic procedures, or produced by recombinant techniques from a prokaryotic or eukaryotic host (for example, by bacterial, yeast, higher plant, insect and mammalian cells in culture).

The present invention also provides antibodies specific for the SNS sodium channel hereinabove defined. The term antibody as used herein includes all immunoglobulins and fragments thereof which contain recognition sites for antigenic

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determinants of proteins of the present invention. The antibodies of the present invention may be polyclonal or preferably monoclonal, may be intact antibody molecules or fragments containing the active binding region of the antibody, e.g. Fab or F(ab)₂ and can be produced using techniques well established in the art [see e.g. R.A DeWeger et al; Immunological Rev., 62 p29-45 (1982)].

The proteins, their fragments or other derivatives, or analogs thereof, or cells expressing them can be used as an immunogen to produce antibodies thereto. These antibodies can be, for example, polyclonal or monoclonal antibodies. The present also includes chimeric, single chain and humanized antibodies, as well as Fab fragments, or the product of an Fab expression library. Various procedures known in the art may be used for the production of such antibodies and fragments.

Antibodies generated against the SNS sodium channel can be obtained by direct injection of the polypeptide into an animal or by administering the protein to an animal, preferably a nonhuman. The antibody so obtained will then bind the protein itself. In this manner, even a sequence encoding only a fragment of the protein can be used to generate antibodies binding the whole native protein. Such antibodies can then be used to locate the protein in tissue expressing that polypeptide. For preparation of monoclonal antibodies, any technique which provides antibodies produced by continuous cell line cultures can be used. Examples include the hybridoma technique (Kohler and Milstein, 1975, Nature 256:495-497), the trioma technique, the human B-cell hybridoma technique (Kozbor et al., 1983, Immunology Today 4:72), and the EBV-hybridoma technique to produce human monoclonal antibodies (Cole, 35 al., 1985, in Monoclonal Antibodies and Cancer Therapy, Alan R. Liss., pp. 77-96).

Techniques described for the production of single chain antibodies (U.S. Pat. No. 4,946,778) can be adapted to produce single chain antibodies to immunogenic polypeptide products of this invention.

The antibodies of the present invention may also be of interest in purifying a protein of the present invention and accordingly there is provided a method of purifying a protein of the present invention as hereinabove defined or any portion thereof or a metabolite or degration product thereof which method comprises the use of an antibody of the present invention.

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The purification method of the present invention may be effected by any convenient technique known in the art for example by providing the antibody on a support and contacting the antibody with a solution containing the protein whereby the antibody binds to the protein of the present invention. The protein may be released from binding with the antibody by known methods for example by changing the ionic strength of the solution in contact with the complex of the protein/antibody.

The present invention also provides methods of identifying modulators of the sodium channel which is specifically located in sensory neurons comprising contacting a test compound with the sodium channel and detecting the activity of the sodium channel. Preferably, the methods of identifying modulators or screening assays employ transformed host cells that express the sodium channel. Typically, such assays will detect changes in the activity of the sodium channel due to the test compound, thus identifying modulators of the sodium channel. Modulators of the sodium channel are useful in modulating the sensation of pain. Blockers of the sodium channel will prevent the transmission of impulses along sensory neurons and thereby be useful in the treatment of acute, chronic or neuropathic pain.

The sodium channel can be used in a patch clamp or other type of assay, such as the assays disclosed herein in the examples, to identify small molecules, antibodies, peptides, proteins, or other types of compounds that inhibit, block, or otherwise interact with the sodium channel. Such modulators identified by the screening assays can then be used for treatment of pain in mammals.

For example, host cells expressing the SNS sodium channel can be employed in ion flux assays such as ²²Na+ ion flux and ¹⁴C guanidinium ion assays, as described in the examples and in the art, as well as the SFBI fluorescent sodium indicator assays as described in Levi et al., (1994) J. Cardiovascular Electrophysiology <u>5</u>:241-257. Host cells expressing the SNS sodium channel can also be employed in binding assays such as the 3H-batrachotoxin binding assay described in Sheldon et al., (1986) Molecular Pharmaeology <u>30</u>:617-623; the 3H-saxitoxin assay as described in Rogart et al (1983) Proc. Natl. Acad. Sci. USA <u>80</u>:1106-1110: and the scorpion toxin assay described in West et al., (1992) Neuron <u>8</u>:59-70. Additionally, the host cells expressing the SNS sodium channel can be used in electrophysiological assays using patch clamp or two electrode techniques. In general, a test compound is added to the assay and its effect on sodium flux is

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determined or the test compound's ability to competitively bind to the sodium channel is assessed. Test compounds having the desired effect on the SNS sodium channel are then selected. Modulators so selected can then be used for treating pain as described above.

Complementary strands of the nucleotide sequences as hereinabove defined can be used in gene therapy, such as disclosed in U.S. Patent 5,399,346. For example, the cDNA sequence or fragments thereof could be used in gene therapy strategies to down regulate the sodium channel. Antisense technology can be used to control gene expression through triple-helix formation or antisense DNA or RNA, both of which methods are based on binding of a nucleic acid sequence to DNA or RNA. For example, the 5' coding portion of the nucleic acid sequence that encodes the sodium channel is used to design an antisense RNA oligonucleotide of from about 10 to about 40 base pairs in length. A DNA oligonucleotide is designed to be complimentary to a region of the gene involved in transcription (triple helix - see Lee et al., Nucl. Acids Res. 6:3073 (1979); Cooney et al, Science 241:456 (1988); and Deruau et al., Science 251:1360 (1991)), thereby preventing transcription and the product of the sodium channel. The antisense RNA oligonucleotide hybridizes to the mRNA in vivo and blocks translation of the mRNA into the sodium channel. Antisense oligonucleotides or an antisense construct driven by a strong constituitive promoter expressed in the target sensory neurons would be delivered either peripherally or to the spinal cord.

The regulatory regions controlling expression of the sodium channel gene could be used in gene therapy to control expression of a therapeutic construct in cells expressing the sodium channel.

Such regions would be isolated by using the cDNA as a probe to identify genomic clones carrying the gene and also flanking sequence e.g. cosmids. Fragments of the cosmids containing intron or flanking sequence would be used in a reporter gene assay in e.g. DRG cultures or transgenic animals and genomic fragments carrying e.g. promoter, enhancer or LCR activity identified.

The invention will now be further described with reference to the following examples:

Example 1 - Derivation of the sequence of a rat dorsal root ganglia (DRG) sodium channel cDNA by subtraction hybridisation methodology

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1.1 cDNA synthesis from DRG-derived poly-A+ RNA

Dorsal root ganglia (DRG) from all spinal levels of neonatal Sprague-Dawley male and female rats were frozen in liquid nitrogen. RNA is extracted using guanidine isothiocyanate and phenol/chloroform extraction (Chomczynski and Sacchi 1987 Anal Biochem 162,156-159).

Total RNA isolation - the nerve tissue is homogenised using a Polytron homogeniser in 1ml extraction buffer (23.6g guanidinium isothiocyanate, 5ml of 250 mM sodium citrate (pH 7.0) made up to 50ml with distilled water. To this is added 2.5ml 10% sarcosyl and 0.36ml \(\beta\)-mercaptoethanol). 0.1ml of 2M sodium acetate (pH 4.0) is added followed by 1 ml phenol. After mixing, 0.2ml chloroform is added and this is shaken vigorously and placed on ice for 5 minutes. This is then centrifuged at 12,000 revolutions per minute (rpm) for 30 minutes at 4°C. The aqueous phase is transferred to a fresh tube, 1ml of isopropanol is added and this is left at -20°C for an hour followed by centrifuging at 12000 rpm for 30 minutes at 4°C. The pellet is dissolved in 0.1ml extraction buffer and is again extracted with isopropanol. The resulting pellet is washed with 70% ethanol and is resuspended in diethyl pyrocarbonate (DEPC)-treated water. 0.3M sodium acetate (pH5.2) and 2 volumes of ethanol are added and the mixture is placed at -20°C for 1 hour. The RNA is precipitated, washed again with 70% ethanol and resuspended in DEPC-treated water. The optical density is measured at 260 nanometres (nm) to calculate the yield of total RNA. Poly A+ RNA is isolated from the total RNA by oligo-dT cellulose chromatography (Aviv and Leder 1972 Proc Natl Acad Sci 69.1408-1411). The following procedures are carried out at 4°C as far as is possible. Oligo-dT cellulose (Sigma) is prepared by treatment with 0.1M sodium hydroxide for 5 minutes. The oligo-dT resin is poured into a column and is neutralised by washing with neutralising buffer (0.5 M potassium chloride, 0.01M Tris (Trizma base - Sigma -Tris(hydroxymethyl)aminomethane) (pH 7.5). The RNA solution is adjusted to 0.5M potassium chloride, 0.01M Tris (pH7.5) and is applied to the top of the column. The first column eluate is re-applied to the column to ensure sticking of the mRNA to the oligo-dT in the column. The column is then washed with 70ml of neutralising buffer and the polyA+ RNA is eluted with 6ml 0.01M Tris (pH7.5) and 1ml fractions are collected. The poly A+ RNA is usually in fractions 2 to 5 and this is checked by measuring the optical density at

260nm. These fractions are pooled and ethanol precipitated overnight at -70°C, washed in 70% ethanol and then redissolved in deionised water at a concentration of 1mg/ml.

First strand cDNA was generated using 0.5mg DRG poly A+ mRNA, oligo-dT/Not-I primer adapters and SuperScript reverse transcriptase (Gibco-BRL) using methodology as described in example 2. One half of the cDNA was labelled by including 2 MBq ³²P dCTP (Amersham) in the reverse transcriptase reaction. Labelled cDNA is separated from unincorporated nucleotides on Nick columns (Sephadex G50 - Pharmacia).

1.2 Enrichment of DRG-specific cDNA using subtraction hybridisation.

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Poly A+ RNA from various tissues (10µg) is incubated with 10µg photoactivatable biotin (Clontech) in a total volume of 15µl and irradiated at 4°C for 30 minutes with a 250 watt sunlamp. The photobiotin is removed by extraction with butanol, and the cDNA co-precipitated with the biotinylated RNA without carrier RNA (Sive and St. John 1988 Nuc Ac Res 16,10937).

Hybridisation is carried out at 58°C for 40 hours in 20% formamide, 50mM 3-(N-morpholino)propanesulphonic acid (MOPS) (pH 7.6), 0.2% sodium dodecyl sulphate (SDS), 0.5M sodium chloride, 5mM ethylenediaminetetraacetate (EDTA - Sigma). The total reaction volume is 5µl and the reaction is carried out under mineral oil, after an initial denaturation step of 2 minutes at 95°C. 100µl 50mM MOPS (pH 7.4), 0.5M sodium chloride, 5mM EDTA containing 20 units of streptavidin (BRL) is then added to the reaction mixture at room temperature, and the aqueous phase retained after two phenol /chloroform extraction steps. After sequential hybridisation of the cDNA from Example 1.1 with biotinylated mRNA from liver and kidney, followed by cortex and cerebellum, a 80-fold concentration of DRG-specific transcripts is achieved.

One third of the 1-2 ng of residual cDNA is then G-tailed with terminal deoxynucleotide transferase at 37°C for 30 minutes. The polymerase chain reaction is used to amplify the cDNA using an oligo-dT-Not-I primer adapter and oligo-dC primers starting with the sequence AATTCCGA(C)₁₀. Amplification is carried out using 2 cycles of 95°C for 1 min, 45°C for 1 min, 72°C for 5 min, follo 2 d by 2 cycles of 95°C for 1 minute .

58°C for 1 minute and 72°C for 5 minutes. The resulting products are then separated on a

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2% Nu-sieve agarose gel, and material running at a size of greater than 0.5 kilobase pairs (kb) is eluted and further amplified with 6 cycles of 95°C for 1 minute, 58°C for 1 minute and 72°C for 5 minutes. This material is further separated on a 2% Nu-sieve agarose gel, and the material running from 6kb on the gel is eluted and further amplified using the same PCR conditions for 27 cycles. The amplified DNA derived from this high molecular weight region is then further fractionated on a 2 % Nu-Sieve gel, and cDNA from 0.5 to 1.5kb, and from 1.5 to 5kb pooled.

1.3. Library Construction

10μg of the bacteriophage vector lambda-zap II (Stratagene) is restriction digested with NotI and EcoRI in high salt buffer overnight at 37°C followed by dephosphorylation using 1 unit of calf intestinal phosphatase (Promega) for 30 minutes at 37°C in 10mM Tris.HCl (pH9.5), 1mM spermidine, 0.1mM EDTA. DRG cDNA is digested with Klenow enzyme in the presence of dGTP and dCTP to construct an EcoRI site from the oligo-dC primer (see above) at the 5' end of the cDNA, and cut with NotI for directional cloning. The cDNA is ligated into the cloning vector bacteriophage lambda-zap II for 16 hours at 12°C. Recombinant phage DNA is then packaged into infective phage using Gigapack gold (Stratagene) and protocols specified by the suppliers. 0.1% of the packaged DNA is used to infect E.coli BB4 cells which are plated out to calculate the number of independent clones generated.

1.4 Differential Screening

The library is plated at a low density (10³ clones/ 12 x 12 cm² dish) and screened using three sets of ³²P-labelled cDNA probes and multiple filter lifts. Replica filters are made by laying them onto the plated library plates, briefly drying them and then laying onto fresh agar plates to increase the quantity of phage and the subsequent hybridisation signals of lifts taken from them. The probes are derived from: a) cortex and cerebellum poly (A)+ RNA, b) DRG poly (A)+ RNA, and c) subtracted cDNA from DRG. The two mRNA probes are labelled with ³²P dCTP using a reaction mixture containing 2-5µg RNA, 50µl 5 x RT buffer, 25 µl 0.1M dithiothreitol (DTT), 12.5µl

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10mM dATP, dGTP, dCTP, 30pM oligo-dT, 75 μl ³²P-dCTP (30MBq; Amersham), 25μl 100μM dCTP, 2μl RNasin (2units/μl) and 2μl SuperScript reverse transcriptase (GibcoBRL) in a final volume of 250µl. The reaction is incubated at 39°C for 60 minutes, and the RNA subsequently destroyed by adding 250µl water, 55µl 1M NaOH, and incubating at 70°C for 20 minutes. The reaction mixture is neutralised with acidified Tris base (pH 2.0) and precipitated with carrier tRNA (Boehringer) with isopropanol. The subtracted and amplified double-stranded DRG cDNA is random-prime labelled with ³²P dATP (Gibco multiprime kit). Replica filters are then prehybridised for 4 hours at 68°C in hybridisation buffer. Hybridisation was carried out for 20 hours at 68°C in 4x SSC (20xSSC consists of 175.3g of sodium chloride and 88.2g of sodium citrate in 800ml of distilled water. The pH is adjusted to 7.0 with 10N sodium hydroxide and this is made to 1 litre with distilled water), 5x Denhardts solution containing 150 µg/ml salmon sperm DNA, 20μg/ml poly-U, 20μg/ml poly-C, 0.5% SDS (Sigma), 5mM EDTA. The filters are briefly washed in 2 x SSC at room temperature, then twice with 2 x SSC with 0.5% SDS at 68°C for 15 minutes, followed by a 20 minute wash in 0.5% SDS, 0.2 x SSC at 68°C. The filters are autoradiographed for up to 1 week on Kodak X-omat film. Plaques that hybridise with DRG probes but not cortex and cerebellum probes are picked, phage DNA prepared and the cloned inserts released for subcloning into pBluescript (Stratagene).

The positive plaques are picked by lining up the autoradiogram with the plate using orientation marks and taking a plug from the plate corresponding to the positive hybridisation signal. The phage is eluted from the plug in 0.5ml phage dilution buffer (10mM Tris chloride (pH7.5) 10mM magnesium sulphate) and the phage re-infected into E.coli BB4 and replated at a density of 200 to 1000 plaques/150mm plate as a secondary purification step to ensure purity of the clones. The positive secondaries are then picked as described previously. In order to sub-clone the insert DNA from the positive recombinant phage, they need to be amplified. This is accomplished by plate lysis where the phage totally lyse the E.coli BB4. 0.2ml of phage suspension is mixed with 0.1ml of an overnight culture of E.coli. This is added to 2.5ml of top agar (16g bacto-tryptone 10g bacto-yeast extract. 5g sodium chloride. 7g bacto-agar in 900mls distilled water) and plated onto 9cm² agar plates. These are incubated overnight at 37°C. 5ml of phage dilution buffer is then added to the plates and is incubated overnight at 4°C or for 4 hours with gentle scraping at

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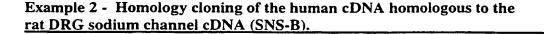
room temperature. The phage-containing buffer is then recovered, 0.1ml chloroform is added and this phage stock is titrated as above and stored at 4°C. Phage DNA is prepared by first infecting 10¹⁰ E.coli B44 with 10⁹ plaque forming units (pfus) of phage in 3ml of phage dilution buffer and shaking at 37°C for 20 minutes. The infected bacteria are added to 400ml of L broth (1.6% bactotryptone, 0.5% (w/v) Bacto yeast extract, 0.5% (w/v) magnesium sulphate) with vigorous shaking at 37°C for 9 hours. When lysis has occurred, 10ml of chloroform is added and shaking is continued for a further 30 minutes. The culture is then cooled to room temperature and pancreatic RNAase and DNAase are added to lug/ml for 40 minutes. Sodium chloride is then added to 1M and is dissolved by swirling on ice. After centrifuging at 8000rpm for 10 minutes the supernatant is recovered. Polyethylene glycol (PEG 6000) is added to 10% w/v and is dissolved by stirring whilst on ice for 2 hours. After centrifuging for 8000rpm for 10 minutes at 4°C the pellet is resuspended in 8ml of phage dilution buffer. This is extracted with an equal volume of phenol/chloroform followed by purification on a caesium chloride gradient (0.675g/ml caesium chloride - 24 hours at 38000 rpm at 4°C). The opaque phage band is removed from the centrifugation tube and dialysed against 10mM sodium chloride, 50mM Tris (pH8.0), 10mM magnesium chloride for 2 hours. EDTA is then added to 20mM, proteinase K to 50µg/ml and SDS to 0.5% and is incubated at 65°C for 1 hour. After dialysis overnight against TE pure phage DNA results. The cloned insert is digested from the purified phage DNA using restriction enzymes as previously described. Each phage insert is then ligated into a plasmid vector e.g. pBluescript - Clontech using a ligation reaction as previously described.

Clone characterisation.

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The plasmids are cross hybridised with each other. Unique clones are further analysed by Northern blotting and sequencing. The clone/s showing transcript sizes and sequence comparable with sodium channels are then used as hybridisation probes to screen a neonatal rat DRG oligo dT-primed full length cDNA library to derive full length cDNA clones using methodology as described above and in example 2. Biological activity of the rat DRG sodium channel is confirmed as in examples 4 and 7 below.



2.1. Isolation of human ganglia total RNA

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The starting material for the derivation of the human cDNA homologue of the rat DRG sodium channel cDNA is isolated human dorsal root ganglia or trigeminal ganglia or other cranial ganglia from post-mortem human material or foetuses. Total ribonucleic acid (RNA) is isolated from the human neural tissue by extraction in guanidinium isothiocyanate (Chomczynski and Sacchi 1987 Anal Biochem 162,156-159) as described in example 1.

2.2 Determination of the transcript size of the human homologue of the rat DRG sodium_channel cDNA (SNS-B).

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Human dorsal root ganglia total RNA is electrophoretically separated in a 1% (w/v) agarose gel containing a suitable denaturing agent e.g. formaldehyde (Lehrach et al 1977 Biochemistry 16,4743; Goldberg 1980 Proc Natl Acad Sci 77,5794; Seed 1982 in Genetic engineering: principles and methods (ed JK Setlow and A Hollaender) vol 4 p91 Plenum Publishing New York) or glyoxal/DMSO (McMaster GK and Carmichael GG 1977 Proc Natl Acad Sci 74,4835), followed by transfer of the RNA to a suitable membrane (e.g. nitrocellulose). The immobilised RNA is then hybridised to radioactive (or other suitable detection label) probes consisting of portions of the rat sodium channel cDNA sequence (see below). After washing of the membrane to remove non-hybridised probe, the hybridised probe is visualised using a suitable detection system (e.g. autoradiography for ³²P labelled probes) thus revealing the size of the human homologous mRNA molecule. Specifically, 20-30 µg total RNA from neonatal rat tissues are separated on 1.2% agarose -formaldehyde gels, and capillary blotted onto Hybond-N (Amersham) (Ninkina et al. 1993 Nuc Ac Res 21.3175-3182). The amounts of RNA on the blot are roughly equivalent, as judged by ethidium bromide staining of ribosomal RNA or by hybridisation with the ubiquitously expressed L-27 ribosomal protein transcripts (Le Beau et al. 1991 Nuc Ac Res 19,1337). Each Northern blot contains human DRG, cortex, cerebellum, liver kidney, spleen and heart RNA. Probes (50ng) are labelled with ³²P-dATP (Amersham) by random priming. Filters are prehybridised in 50% formaldehyde 5 x SSC containing 0.5% SDS, 5 x Denhardts solution (50x Denhardts contains 5g of Ficoll (Type 400, Pharmacia), 5g of polyvinylpyrrolidone, 5g of bovine serum albumin (Fraction V, Sigma) and water to 500ml), 100 μg/ml boiled salmon sperm DNA, 10 μg/ml poly-U and 10 μg/ml poly-C at 45°C for 6 hours. After 36 hours hybridisation in the same conditions, the filters are briefly washed in 2 x SSC at room temperature, then twice with 2 x SSC with 0.5% SDS at 68°C for 15 minutes, followed by a 20 minute wash in 0.5% SDS, 0.2 x SSC at 68°C. The filters are autoradiographed for up to 1 week on Kodak X-omat film. The transcript size is calculated from the signal from the gel in comparison with gel molecular weight standard markers.

2.3 Production of a human DRG cDNA library

In order to produce a representative cDNA library from the human dorsal root ganglia messenger RNA (poly A+ mRNA) is first isolated from the total RNA pool using oligo-dT cellulose chromatography (Aviv and Leder 1972 Proc Natl Acad Sci 69,1408-1411) using methodology described in example 1. Synthesis of the first strand of cDNA from the polyA+ RNA uses the enzyme RNA-dependent DNA polymerase (reverse transcriptase) to catalyse the reaction. The most commonly used method of second strand cDNA synthesis uses the product of first strand synthesis, a cDNA:mRNA hybrid, as a template for priming the second strand synthesis. (Gubler and Hoffman 1983 Gene 25,263)).

2.3.1. First strand cDNA synthesis

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20μg of human DRG polyA+ RNA is pre-treated to destroy secondary structure which may inhibit first strand cDNA synthesis. 20μg of polyA+ RNA, 1μ1 1M Tris (pH7.5) are made up to a volume of 100μl with distilled water. This is incubated at 90°C for 2 minutes followed by cooling on ice. 4.8 μl of 100 mM methyl mercury is then added for 10 minutes at room temperature. 10μ, of 0.7M β-mercaptoethanol and 100 units of human placental RNAase inhibitor are then added for 5 minutes at room temperature.

The first strand synthesis reaction consists of 8µl 20mM dATP, 5µl 20mM dCTP, 8µl 20mM dGTP 8µl 20mM dTTP, 10µl 1mg/ml oligo-dT (12-18), 20µl 1M Tris (pH 8.3) (at 45°C), 8µl 3M potassium chloride, 3.3µl 0.5M magnesium chloride, 3µl a³²P dCTP, 100 units Superscript II reverse transcriptase (GibcoBRL) made up to 200µl with distilled water. This reaction mixture is incubated at 45°C for 45 minutes after which another 50 units of Superscript reverse transcriptase is added and incubated for a further 30 minutes at 45°C. EDTA is then added to 10mM to terminate the reaction and a phenol/chloroform extraction is carried out. The DNA is then precipitated using ammonium acetate (freezing in dry ice/ethanol before centrifuging), washed with 70% ethanol and resuspended in 50ml distilled water. The size of the single stranded DNA is assessed by electrophoretically separating it out on an agarose gel (1% w/v) and autoradiographing the result against markers.

2.3.2 Second strand synthesis

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The second strand synthesis reaction mixture consists of 0.5µg human DRG single stranded DNA, 2µl 1M Tris (pH7.5), 1µl 0.5M magnesium chloride, 3.33µl 3M potassium chloride, 2µl 0.5M ammonium sulphate, 1.5µl 10mM βnicotinamide adenine dinucleotide (NAD), 4µl of each of the 1mM dNTPs, 5µl 1mg/ml bovine serum albumin (BSA), 1 unit RNAase-H, 25 units Klenow polymerase all made up to 100µl with distilled water. This is incubated at 12°C for 1 hour and then at 20°C for 1 hour. The reaction is stopped by addition of EDTA to 20mM followed by a phenol/chloroform extraction. The DNA is ethanol precipitated (-70°C overnight) and is then washed with 70% ethanol followed by resuspension in 20µl distilled water. Size is checked by gel electrophoresis and autoradiography.

2.3.3 Double stranded cDNA end repair

In order to add linkers to the end of the cDNA molecules for subsequent cloning, the ends must first be repaired. The human DRG cDNA is treated with 500 units/ml of \$1 nuclease in 0.25M sodium chloride. 1mM zinc sulphate. 50mM sodium

acetate (pH4.5). Incubation is at 30°C for 40 minutes followed by neutralisation with Tris (pH 8.0) to 0.2M. The DNA is again ethanol precipitated, washed in 70% ethanol and resuspended in 20ul distilled water. The size is again checked to ensure that S1 nuclease digestion has not radically reduced the average DNA fragment size. The repair reaction consists of 19µl cDNA, 3µl 10xT4 polymerase buffer (0.33M Tris acetate (pH7.9), 0.66M potassium acetate, 0.1M magnesium acetate, 1mg/ml BSA and 5mM DTT), 2µl of each dNTP at 2mM, 2µl T4 polymerase and 4µl distilled water. This is incubated at 37°C for 30 minutes followed by addition of 1µl Klenow polymerase for 1 hour at room temperature. The DNA is then ethanol precipitated. washed in 70% ethanol and resuspended in 5µl distilled water. In order to protect naturally occurring restriction sites within the cDNA from being cleaved, the cDNA is treated with a methylase before the addition of linkers. The reaction mixture consists of 5µl human DRG double stranded DNA, 1µl Sadenosylmethionine, 2µl 1mg/ml BSA, 2µl 5x methylase buffer (0.5M Tris (pH8.0), 5mM EDTA), 0.2µl EcoRI methylase (NEB). This is incubated at 37°C for 20 minutes followed by phenol extraction, ethanol precipitation washing with 70% ethanol and resuspension in 20µl distilled water.

2.3.4. Addition of linkers to cDNA

EcoRI linkers are ligated to the cDNA molecules to facilitate cloning into lambda vectors. The ligation reaction mixture consists of 1μl 10x ligation buffer (0.5M Tris chloride (pH7.5), 0.1M magnesium chloride and 0.05M DTT), 1μl 10mM ATP, 100ng cDNA, 5μg EcoRI linkers, 1 unit T4 DNA ligase, distilled water to 10μl. The reaction is incubated at 37°C for 1 hour, followed by addition of 6 more units of T4 ligase and a further incubation overnight at 15°C. The ligated samples are ethanol precipitated, washed in 70% ethanol and resuspended in 10μl distilled water. The cDNA is then digested with EcoRI to cleave any linker concatangers formed in the ligation process. This restriction digestion reaction contains 10μl cDNA, 2μl high salt buffer (10mM magnesium chloride, 50mM Tris chloride (pH7.5), 1mM DTT, 100mM sodium chloride), 2μl EcoRI (10 units/μl - NEB) and distilled water to 20μl. The digestion is carried out for 3 hours. The ligation

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and digestion steps are monitored using gel elecrophoresis to monitor the size of the products.

5 2.3.5 Size fractionation of cDNA

In order to assure that the library is not swamped with short cDNA molecules and to remove linker molecules a column purification is carried out. A 1ml Sepharose 4B column is made in a 1 ml plastic pipette plugged with a small piece of glass wool. This is equilibrated with 0.1M sodium chloride in TE. The cDNA is loaded onto the column and 1 drop fractions are collected. 2µl aliquots of each fraction are analysed by gel electrophoresis and autoradiography to determine the sizes of the cDNA in each fraction. Fractions containing cDNA of about 800 base pairs and above are pooled and purified by ethanol precipitation and resuspending in 10µl distilled water.

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2.3.6 Cloning of cDNA into bacteriophage vector

Bacteriophage vectors designed for the cloning and propagation of cDNA are provided ready-digested with EcoRI and with phosphatased ends from commercial sources (e.g. lambda gt10 from Stratagene). The prepared subtracted cDNA is ligated into lambda gt10 using a ligation rection consisting of ligase buffer and T4 DNA ligase (New England Biolabs) as described elsewhere in this document.

2.4 Labelling of cDNA fragments (probes) for library screening

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The 3' untranslated region of the rat DRG sodium channel cDNA clone (SNS-B) is subcloned using appropriate restriction enzymes into a plasmid vector e.g. pBluescript - Stratagene. The cDNA insert which is to form the labelled probe is released from the vector via digestion with appropriate restriction enzymes and the insert is separated from the vector via electrophoresis in a 1% (w/v) agarose gel. After removal of the separated insert from the agarose gel and purification it is labelled by standard

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techniques such as random priming and polymerisation (Feinberg and Vogelstein 1983 Anal Biochem 132,6) or nick translation (Rigby et al 1977 J Mol Biol 113,237) with ³²P or DIG-labelled nucleotides. Alternatively, if the probe cDNA insert is cloned into a vector containing strong bacteriophage promoters to which DNA-dependant RNA polymerases bind (SP6, T3 or T7 polymerases), synthetic cRNA is produced by in vitro transcription which incorporates ³²P or digoxygenin nucleotides. Other regions of the rat DRG sodium channel cDNA can also be used as probes in a similar fashion for cDNA library screening or Northern blot analysis. Specifically, a probe is made using a kit such as the Pharmacia oligo labelling kit. This will radioactively label the rat DRG sodium channel cDNA fragment. 50ng of denatured DNA (place in boiling waterbath for 5 minutes), 3µl of ³²PdCTP (Amersham) and 10µl reagent mix is made up to 49µl with distilled water. 1µl of Klenow fragment is added and the mixture is incubated at 37°C for one hour. To remove unincorporated nucleotides, the reaction mixture is applied to a Nick column (Sephadex G50 - Pharmacia) followed by 400µl of TE (10mM Tris chloride (pH7.4) 1mM EDTA (pH8.0)). Another 400µl of TE is added and the eluate is collected. This contains the labelled DNA to be used as a hybridisation probe.

2.5 cDNA library screening

In order to detect recombinants containing human homologues of the rat DRG sodium channel the human DRG cDNA library is screened using moderate stringency hybridisation washes (50-60°C, 5 x SSC, 30 minutes), using radiolabelled or other labelled DNA or cRNA probes derived from the 3' untranslated region as described above. Libraries are screened using standard methodologies involving the production of nitrocellulose or nylon membrane replicas of DNA from recombinant plaques formed on agar plates (Benton et al 1977 Science 196:180). These are then hybridised to single stranded nucleic acid probes (see above). Moderate stringency washes are carried out (see wash conditions for Northern analysis in section 2.2). Plaques which are positive on duplicate filters (i.e. not artefacts or background) are then purified by one or more rounds of replating after dilution to separate the colonies and further hybridisation screening. Resulting positive plaques are purified. DNA is extracted and the insert sizes of these

clones is examined. The clones are cross-hybridised to each other using standard techniques (Sambrook et al 1989 Molecular Cloning Second Edition Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York) and distinct positive clones identified. Detailed protocols for cDNA library screening are given in example 1.

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2.6 Derivation of a full-length clone of the human homologue of the rat DRG sodium channel cDNA

Overlapping positive clones from above are identified by 10 cross-hybridisation. They are then restriction mapped to identify their common portions and restriction fragments representing the separate portions from the overlapping clones are ligated together using standard cloning techniques (Sambrook et al 1989 Molecular Cloning Second Edition Cold Spring Harbor Laboratory Press). For example, the most 5' fragment will contain any 5' untranslated sequence, the start codon ATG and 5' coding 15 sequence. The most 3' clone will contain the most 3' coding sequence, a stop codon and any 3' untranslated sequence, a poly A consensus sequence and possibly a poly A run. Thus a recombinant molecule is generated which contains the full cDNA sequence of the human homologue of the rat DRG sodium channel cDNA. If overlapping clones do not produce sufficient fragments to assemble a full length cDNA clone, the full length oligo dT-primed 20 human DRG library is re-screened to isolate a full length clone. Alternatively, a full length clone is derived directly from the library screening.

2.7 Characterisation of the human homologue full-length clone

The cDNA sequence from the full-length clone is used as a probe in Northern blot analysis to detect the messenger RNA size in human tissue for comparison with the rat messenger RNA size (see sections 1.1 and 2.2 for methodology).

Confirmation of biological activity of the cloned cDNA is carried out via in vitro translation of the human sodium channel mRNA and its expression in Xenopus oocytes in an analogous manner to that for the rat DRG=specific TTXi resistant sodium channel as described in examples 4 and 7.

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cDNA sequences which are shown to have activity as defined above are completely sequenced using dideoxy-mediated chain termination sequencing protocols (Sanger et al 1977 Proc Natl Acad Sci 74,5463).

5 Example 3 - Polymerase chain reaction (PCR) approaches to clone the human DRG sodium channels using DNA sequence derived from the rat DRG sodium channel cDNA clone

Total RNA and poly A+ RNA is isolated from human dorsal root ganglia or trigeminal ganglia or other cranial ganglia from post-mortem human material or foetuses as described in example 2 above.

Random primers are hybridised to the RNA followed by polymerisation with MMLV reverse transcriptase to generate single stranded cDNA from the extracted human RNA.

Using degenerate PCR primers derived from relatively conserved regions of the known voltage-gated sodium channels (Figure 2), amplify the cDNA using the polymerase chain reaction (Saiki et al 1985 Science 230,1350). It is appreciated by those skilled in the art that there are many variables which can be manipulated in a PCR reaction to derive the homologous sequences required. These include but are not limited to varying cycle and step temperatures, cycle and step times, number of cycles, thermostable polymerase, Mg2+ concentration. It is also appreciated that greater specificity can be gained by a second round of amplification utilising one or more nested primers derived from further conserved sequence from the sodium channels.

Specifically, the above can be accomplished in the following manner. The
first strand cDNA reaction consists of lµg of total RNA made up to 13µl with
DEPC-treated water and lµl of 0.5µg/µl oligo(dT). This is heated to 70°C for 10 minutes
and then incubated on ice for 1minute. The following is then added: 2µl of 10x synthesis
buffer (200mM Tris chloride, 500mM potassium chloride, 25mM magnesium chloride,
lµg/ml BSA), 2µl of 0.1M DTT, 1µl of 200U/µl Superscript Reverse Transcriptase (Gibco
BRL). This is incubated at room temperature for 10 minutes then at 42°C for 50 minutes.
The reaction is then terminated by incubating for 15 minutes at 70°C. 1µl of E.coli RNase
H (2U/µl) is added to the tube which is then incubated for 20 minutes at 37°C.

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The PCR reaction is set up in a 0.5ml thin-walled Eppendorf tube. The following reagents are added: 10µl 10x PCR buffer, 1µl cDNA,16µl dNTP's (25µl of 100µM dATP,dCTP, dCTP and dGTP into 900µl sterile distilled water), 7µl of 25mM magnesium chloride, 1µl of Taq DNA polymerase (Amplitaq Perkin-Elmer)plus sterile distilled water to 94µl.

To each reaction tube a wax PCR bead is added (Perkin-Elmer) and the tube placed in a 70°C hot block for 1 minute. The tubes are allowed to cool until the wax sets and 3μl of each primer (33pM/μl) are added above the wax. The tubes are placed in a thermal cycler (Perkin-Elmer) and the following 3-step program used after an initial 94°C for 5 minutes; 92°C for 2 minutes, 55°C for 2 minutes, 72°C for 2 minutes for 35 cycles. A final polymerisation step is added at 72°C for 10 minutes. The reaction products are then run on a 1% agarose gel to assess the size of the products. In addition, control reactions are performed alongside the samples. These should be: 1) all components without cDNA (negative control) and 2) all reaction components with primers for constitutively expressed product e.g. α-actin or HPRT.

The products of the PCR reactions are examined on 0.8%-1.2% (w/v) agarose gels. Bands on the gel (visualised by staining with ethidium bromide and viewing under UV light) representing amplification products of the approximate predicted size were then cut from the gel and the DNA purified. Further bands of interest are also identified by Southern blot analysis of the amplification products and probing of the resulting filters with labelled primers from further conserved regions e.g. those used for secondary amplification.

The resulting DNA is ligated into suitable vectors such as, but not limited to, pCR II (Invitrogen) or pGemT. Clones are then sequenced to identify those containing sequence with similarity to the rat DRG sodium channel sequence (SNS-B).

Clone analysis

Candidate clones from above are used to screen a human cDNA DRG
library constructed using methods described in example 2. If a full length clone is not identified, positive overlapping clones which code for the full length human cDNA

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homologue are identified and a full length clone is then assembled as described in example

1. Biological activity is then confirmed as described in examples 4 and 7.

Example 4 - In vitro translation of rat and human DRG sodium channel in Xenopus laevis oocvtes

In order to demonstrate the biological activity of the protein coded for by the rat DRG sodium channel cDNA sequence (SNS-B) and its human homologue the complete double-stranded cDNA coding sequences are ligated into in vitro transcription vectors (including but not limited to the pGEM series, Promega) using one or more of the available restriction enzyme sites such that the cDNAs are inserted in the correct orientation. The constructs are then used to transform bacteria and constructs with the correct sequence in the correct orientation are identified via diagnostic restriction enzyme analysis and dideoxy-mediated chain termination DNA sequencing (Sanger et al 1977 Proc Natl Acad Sci 74,5463).

These constructs are then linearised at a restriction site downstream of the coding sequence and the linearised and purified plasmids are then utilised as a template for in vitro transcription. Sufficient quantities of synthetic mRNA are produced via in vitro transcription of the cloned DNA using a DNA-dependent RNA polymerase from a bacteriophage that recognises a bacteriophage promoter found in the cloning vector. Examples of such polymerases include (but are not limited to) T3, T7 and SP6 RNA polymerase.

A variation on the above method is the synthesis of mRNA containing a 5' terminal cap structure (7-methylguanosine) to increase its stability and enhance its translation efficiency (Nielson and Shapiro 1986 Nuc Ac Res 14,5936). This is accomplished by the addition of 7-methylguanosine to the reaction mixture used for synthetic mRNA synthesis. The cap structure is incorporated into the 5' end of the transcripts as polymerisation occurs. Kits are available to facilitate this process e.g. mCAP RNA Capping Kit - Stratagene).

The synthetic RNA produced from the in vitro transcription is isolated and purified. It is then translated via microinjection into Xenopus laevis oocytes. 50nls of 1mg/ml synthetic RNA is micro-injected into stage 5 or stage 6 oocytes according to methods established in the literature (Gurdon et al (1983) Methods in Enzymol 101.370).

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After incubation to allow translation of the mRNAs the oocytes are analysed for expression of the DRG sodium channels via electrophysiological or other methods as described in example 7.

A further method for expression of functional sodium channels involves the nuclear injection of a Xenopus oocyte protein expression vector such as pOEV (Pfaff et al., Anal. Biochem. 188, 192-195 (1990)) which allows cloned DNA to be transcribed and translated directly in the oocyte. Since proteins translated in oocytes are post-translationally modified according to conserved eukaryotic signals, these cells offer a convenient system for performing structural and functional analyses of cloned genes. pOEV can be used for direct analysis of proteins encoded by cloned cDNAs without preparing mRNA in vitro, simplifying existing protocols for translating proteins in oocytes with a very high translational yield. Transcription of the vector in oocytes is driven by the promoter for the TFIIIA gene, which can generate 1-2 ng (per oocyte within 2 days) of stable mRNA template for translation. The vector also contains SP6 and T7 promoters for in vitro transcription to make mRNA and hybridization probes. DNA clones encoding SNS channel transcripts are injected into oocyte nuclei and protein accumulated in the cell over a 2- to 10-day period. The presence of functional protein is then assessed using twin electrode voltage clamp as described in example 7.

20 Example 5 - Expression of rat and human DRG sodium channel in mammalian cells

In order to be able to establish a mammalian cell expression system capable of producing the sodium channel in a stable bioactive manner, constructs have to be first generated consisting of the cDNA of the channel in the correct vectors suitable for the cell system in which it is desired to express the protein. There are available a range of vectors containing strong promoters which drive expression in mammalian cells.

i/ Transient expression

In order to determine rapidly the bioactivity of a given cDNA it can be introduced directly into cells and resulting protein activity assayed 48-72 hours later.

Although this does not result in a cell line which is stably expressing the protein of interest

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it does give a quick answer as to the biological activity of the molecule. Specifically, the cDNA representing the human or rat DRG sodium channel is ligated into appropriate vectors (including but not limited to pRc/RSV, pRc/CMV, pcDNA1 (Invitrogen)) using appropriate restriction enzymes such that the resulting construct contains the cDNA in the correct orientation and such that the heterologous promoter can drive expression of the transcription unit. The resulting expression constructs are introduced into appropriate cell lines including but not limited to COS-7 cells (an African Green Monkey Kidney cell line), HEK 293 cells (a human embryonic kidney cell line) and NIH3T3 cells (a murine fibroblastic cell line). The DNA is introduced via standard methods (Sambrook et al 1989 Molecular Cloning Second Edition, Cold Spring Harbour Laboratory Press) including but not limited to calcium phosphate transfection, electroporation or lipofectamine (Gibco) transfection. After the required incubation time at 37°C in a humidified incubator the cells are tested for the presence of an active rat DRG sodium channel using methods described in example 7.

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ii/Stable expression

The production of a stable expression system has several advantages over transient expression. A clonal cell line can be generated that a has a stable phenotype and in which the expression levels of the foreign protein can be characterised and, with some expression systems, controlled. Also, a range of vectors are available which incorporate genes coding for antibiotic resistance, thus allowing the selection of cells transfected with the constructs introduced. Cell lines of this type can be grown in tissue culture and can be frozen down for long-term storage. There are several systems available for accomplishing this e.g. CHO, CV-1, NIH-3T3.

Specifically COS-7 cells can be transfected by lipofection using Lipofectamine (GibcoBRL) in the following manner. For each sample $2x10^6$ cells are seeded in a 90mm tissue culture plate the day prior to transfection. These are incubated overnight at 37° C in a CO₂ incubator to give 50-80% confluency the following day. The day of the transfection the following solutions are prepared in sterile 12×75 mm tubes: Solution A: For each transfection, dilute $10-50\mu g$ of DNA into 990 μ l of serum-free media (Opti-MEM I Reduced Serum Medium GibcoBRL). Solution B: For each transfection.

dilute 50μl of Lipofectamine Reagent into 950μl serum-free medium. The two solutions are combined, mixed gently and incubated at room temp for 45 minutes. During this time the cells are rinsed once with serum-free medium. For each transfection 9ml of serum-free medium is added to the DNA-lipofectamine tubes. This solution is mixed gently and overlayed on the rinsed cells. The plates are incubated for 5 hours at 37°C in a CO₂ incubator. After the incubation the medium is replaced with fresh complete media and the cells returned to the incubator. Cells are assayed for activity 72 hours post transfection as detailed in examples 4 and 7. To ascertain the efficiency of transfection, β-galactosidase in pcDNA3 is transfected alongside the DRG sodium channel cDNA. This control plate is stained for β-galactosidase activity using a chromogenic substrate and the proportion of cells staining calculated. For transient transfection of DRG the cDNA must first be cloned into a eucaryotic expression vector such as pcDNA3 (Invitrogen).

Example 6 - Expression of rat DRG sodium channel in insect cells

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The baculovirus expression system uses baculovirus such as Autographa californica nuclear polyhedrosis virus (AcNPV) to produce large amounts of target protein in insect cells such as the Sf9 or 21 clonal cell lines derived from Spodoptera frugiperda cells. Expression of the highly abundant polyhedrin gene is non-essential in tissue culture and its strong promoter (polh) can be used for the synthesis of foreign gene products (Smith et al 1983 Mol Cell Biol 3,2156-2165). The polyhedrin promoter is maximally expressed very late in infection (20 hours post infection).

A transfer vector, where the rat DRG sodium channel cDNA is cloned downstream of the polh promoter, or another late promoter such as p10, is transfected into insect cells in conjunction with modified AcNPV viral DNA such as but not limited to BaculoGold DNA (PharMingen). The modified DNA contains a lethal mutation and is incapable of producing infectious viral particles after transfection. Co-transfection with a complementing transfer vector such as (but not limited to) pAcYM1 (Matsuura et al 1987 J Gen Virol 68,1233-1250) or pVL1392/3 (InVitrogen) allows the production of viable recombinant virus. Although more than 99% of the resultant virus particles should be derived from plasmid-rescued virus it is desirable to further purify the virus particles by plaque assay. To ensure that the recombinant stock is clonal, a single plaque is picked from

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the plaque assay and amplified to produce a recombinant viral stock. Once the recombinant phenotype is verified the viral stock can be used to infect insect cells and express functional rat DRG sodium channel. There are a number of variations in the methodology of baculovirus expression which may give increased expression (O'Reilly et al 1992 Baculovirus Expression Vectors: A Laboratory Manual. Oxford University Press). The expression of the rat or human DRG sodium channel is achieved by cloning of the cDNA

Example 7 - Electrophysiological characterisation of cloned human and rat DRG sodium channel expression

into pVL1392 and introducing this into Sf21 insect cells.

Xenopus laevis oocytes are used to express the channel after injection of the mRNA or cDNA in an expression vector. Expression would be transient and thus functional studies would be made at appropriate times after the injections. Comparison with mock-injected oocytes would demonstrate lack of the novel channel as an endogenously expressed characteristic. Standard two electrode voltage clamp (TEVC) techniques as described, for example, in Fraser, Moon & Djamgoz (1993) Electrophysiology of *Xenopus* oocytes: an expression system in molecular neurobiology. In: Electrophysiology: A practical approach. Wallis, D.I., ed. Oxford University Press. Chapter 4 pp. 65-86, would be used to examine the characteristics of responses of ionic 20 currents to changes in the applied membrane potential. Appropriately modified saline media would be used to manipulate the type of ionic currents detectable. The kinetics of activation and inactivation of the sodium current, its ionic selectivity, the effects of changes in ionic concentration of the extracellular medium on its reversal potential, and the sensitivity (or resistance) to TTX would be defining characteristics.

Similar electrophysiological studies would be undertaken to assess the success of functional expression in a permanently or transiently expressing mammalian cell line, but patch clamp methods would be more suitable than TEVC. Whole cell. cell-attached patch, inside-out patch or outside-out patch configurations as described for example by Hamill et al. (1981) Pflugers Arch. 391:85-100 and Fenwick et al. (1982) J. Physiol. 331 599-635 might be used to assess the channel characteristics.

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For example, isolated transfected cells (see above) will be voltage-clamped using the whole-cell variant of the patch clamp technique for recording the expressed sodium channel current.

Recordings will be obtained at room temperature (22-24°C). Both external and internal recording solutions will be used to isolate Na+ currents as previously described (Lalik et al., Am. J. Physiol. 264:C803-C809, 1992; West et al., Neuron 8:59-70, 1992). External solution (mM): sodium chloride, 65; choline chloride, 50; TEA-Cl, 20, KCl, 1.5; calcium chloride, 1; magnesium chloride, 5; glucose 5; HEPES, 5; at a pH 7.4 and and osmolality of 320. Internal solution (mM):CsF, 90; CsCl, 60; sodium chloride, 10; MgCl, 2;EGTA, 10; HEPES, 10 at pH 7.2 and an osmolarity of 315.

The kinetics and voltage parameters of the expressed sodium channel current will be examined and compared with data existing in the literature. These include current-voltage relationships and peak current amplitude. Cells will be voltage-clamped at -70 mV and depolarizing pulses to 50 mV (at 10 mV increments) will be used to generate currents.

The pharmacology of the expressed sodium channel current will be examined with the Na channel blocker, tetrodotoxin (TTX). To date sodium channels have been classified as TTX-sensitive and TTX-resistant: block by low (1-30 nM) and high (> 1 µM) concentrations of TTX, respectively (Elliot & Elliot, J. Physiol. (Lond.) 463:39-56, 1993; Yang et al., J. Neurosci. 12:268-277, 1992; W1992).

The channel is unaffected by concentrations lower than 1 micromolar tetrodotoxin, and is only partially blocked by concentrations as high as 10 micromolar tetrodotoxin.

25 Example 8 - Production of purified channel

Using a commercial coupled transcription-translation system, 35-S methionine-labelled protein products of the SNS clone can be generated (see Figure 3). The size of the resulting protein when assessed by SDS-polyacrylamide gel electrophoresis confirms the predicted size of the protein dedu d by DNA sequencing. The system used

is the Promega TNT system (Promega Technical Bulletin 126 1993). The experiment is carried out precisely according to the protocol provided (see **Figure 3**).

Example 9 - Use of rat or human sodium channel in screening assays

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Cell lines expressing the cloned sodium channels could be used to determine the effects of drugs on the ability of the channels to pass sodium ions across the cell membranes, e.g to block the channels or to enhance their opening. Since the channel activation is voltage dependent, depolarising conditions will be required for observation of baseline activity that would be modified by drug actions. Depolarisation could be achieved by for example raising extracellular potassium ion concentration to 20 or 40 mM, or by repeated electrical pulses. Detection of the activation of sodium conducting channels could be achieved by flux of radiolabelled sodium ions, guanidine or by reporter gene activation leading to for example a colour change or to fluorescence of a light emitting protein. Subsequent confirmation of the effectiveness of the drug action on sodium channel activity would require electrophysiological studies similar to those described above.

Example 10 - In vitro influx assays

- 1. 22Na+ influx assay: A modified assay has been adapted from methods reported by Tamkum and Catterall, Mol Pharm. 19:78, (1981). Oocytes or cells expressing the sodium channel gene are suspended in a buffer containing 0.13 M sodium chloride, 5 mM KCl, 0.8 mM MgSO₄, 50 mM HEPES-Tris (pH 7.4), and 5.5 mM glucose. Aliquots of the
- cell suspension are added a buffer containing 22NaCl (1.3 μCi/ml, New England Nuclear, Boston, MA), 0.128 M choline chloride, 2.66 mM sodium chloride, 5.4 mM KCl, 0.8 mM MgSO₄, 50 mM HEPES-Tris (pH 7.4), 5 mM ouabain, 1mg/ml bovine serum albumin, and 5.5 mM glucose and then incubated at 37 oC for 20 sec in either the presence or absence of 100 μM veratridine (Sigma Chemical Co., St Louis, MO). The influx assay is stopped by the addition of 3 ml of ice-cold wash buffer containing 0.163 M sodium chloride, 0.8 mM MgSO₄, 1.8 mM CaCl₂, 50 mM HEPES-Tris (pH 7.4) and 1mg/ml bovine serum albumin,

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collected on a glass fiber filter (Whatman GF/C), and washed twices with 3 ml of wash buffer. Radioactive incorporation is determined by with a gammacounter. The specific tetrodotoxin-resistant influx is measured by the difference in 22Na+ uptake in the absence or the presence of 10 μ M transmethrin or 1 μ M (+) trans allethrin. The tetrodotoxin-sensitive influx is measured by the difference in 22Na+ uptake in the absence

or the presence of 1 µM tetrodotoxin (Sigma Chemical Co., St Louis, MO).

Guanidine influx: Another assay is modified from the method described by Reith, Eur. J. Pharmacol. 188:33 (1990). In this assay sodium ions are substituted with guanidinium ions. Oocytes or cells are washed twice with a buffer containing 4.74 mM KCl, 1.25 mM CaCl₂, 1.2 mM KH2PO4, 1.18 mM MgSO₄, 22 mM HEPES (pH 7.2), 22 mM choline chloride and 11 mM glucose. The oocytes or cells are suspended in the same buffer containing 250 µM guanidine for 5 min at 19-25 oC. An aliquot of 14C-labelled guanidine hydrochloride (30-50 mCi/mmol supplied by New England Nuclear, Boston, MA) is added in the absence or presence of 10 µM veratridine, and the mixture is incubated for 3 min. The uptake reaction is stopped by filtration through Whatman GF/F filters and followed by 2.5 ml washes with ice-cold 0.9% saline. Radioactive incorporation is determined by scintillation counting.

Example 11

In order to measure the expression of sodium channels in in vitro systems, as well as to analyse distribution and relative level of expression in vivo, and to attempt to block function, polyclonal and monoclonal antibodies will be generated to peptide and protein fragments derived from SNS protein sequence shown in Figure 1.

a) Immunogens

Glutathione-sulphotransferase (GST) - fusion proteins will be constructed (Smith and Johnson Gene 67:31-40 (1988)) using PGEX vectors obtained from Pharmacia. Fusion proteins including both intracellular and extracellular loops with little homology with known sodium channels other than SNS-B will be produced. One such method involves subcloning of fragments into pGex-5X3 or pGEX 4t-2 to produce in-frame fusion

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proteins encoding extracellular, intracellular or C-terminal domains as shown in detailed maps in Figure 4. The pGEX fusion vectors are transformed into E. coli XL-1 blue cells or other appropriate cells grown in the presence of ampicillin. After the cultures have reached an optical density of OD600 > 0.5, fusion protein synthesis is induced by the addition of 100 micromolar IPTG, and the cultures further incubated for 1-4 hours. The cells are harvested by centrifugation and washed in ice cold phosphate buffered saline. The resuting pellet (dissolved in 300 microlitres PBS from each 50 ml culture) is then sonicated on ice using a 2mm diameter probe, and the lysed cells microfuged to remove debris. 50 microlitres of glutathione-agarose beads are then added to each pellet, and after gentle mixing for 2 minutes at room temperature, the beads are washed by successive spins in PBS. The washed beads are then boiled in Laemmli gel sample buffer, and applied to 10% polyacrylamide SDS gels. Material migrating at the predicted molecular weight is identified on the gel by brief staining with coommassie blue, and comparison with molecular weight markers. This material is then electroeluted from the gel and used as an immunogen as described below.

b) Antibody production

Female Balb/c mice are immunised intraperiteonally with 1-100 micrograms of GST fusion protein emulisfied in Freunds complete adjuvant. After 4 weeks, the animals will be further immunised with fusion proteins (1-100 micrograms) emulsified in Freunds incomplete adjuvant. Four weeks later, the animals will be immunised intraperitoneally with a further 1-100 micrograms of GST fusion protein emulsified with Freunds incomplete adjuvant. Seven days later, the animals will be tail bled, and their serum assessed for the production of antibodies to the immunogen by the following screen; (protocols for the production of rabbit polyclonal serum are the same, except that all injections are subcutaneous, and 10 times as much immunogen is used. Polyclonal rabbit serum are isolated from ear-vein bleeds.)

Serial ten-fold dilutions of the sera (1:100 to 1: 1000,000) in phosphate

buffered saline (PBS) containing 0.5% NP-40 and 1% normal goat serum will be applied to

4% paraformaldehyde-fixed 10 micron sections of neonatal rat spinal cord previously

treated with 10% goat serum in PBS. After overnight incubation, the sections are washed in

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PBS, and further incubated in the dark with 1;200 FITC-conjugated F(ab)2 fragment of goat anti-mouse antibodies for 2 hours in PBS containing 1% normal goat serum. The sections are further washed in PBS, mounted in Citifluor, and examined by fluorescence microscopy. Those sera that show specific staining of laminar II in the spinal cord will be retained, and the mice generating such antibodies subsequently used for the production of monoclonal antibodies. Three weeks later, mice producing useful antibodies are immunised with GST-fusion proteins without adjuvant. After 3 days, the animals are killed, their spleens removed, and the lymphocytes fused with the thymidine kinase-negative myeloma line NSO or equivalent, using polyethylene glycol. The fused cells from each experiment are grown up in 3 x 24 well plates in the presence of DMEM medium containing 10% fotal calf serum and hypoxanthine, aminopterin and thymidine (HAT) medium to kill the myeloma cells (Kohler and Milstein, Eur. J. Immunol 6, 511-519 (1976)). The tissue culture supernatants from wells containing hybridomas are further screened by immunofluorescence as described above, and cells from positive wells cloned by limiting dilution. Antibody from the positive testing cloned hybridomas is then used to Western blot extracts of rat dorsal root ganglia, to determine if the antibody recognises a band of size approximately 200,000, confirming the specificity of the monoclonal antibody for the SNS sodium channel. Those antibodies directed against extracellular domains that test positive by both of these criteria will then be assessed for function blocking activity in electrophysiological tests of sodium channel function (see example 7), and in screens relying on ion flux or dye-based assays in cells lines expressing sodium channel (see examples 9 and 10).

Example 12 - Cell-type distribution of expression

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In situ hybridization demonstrates the presence of SNS in a subset of sensory neurons. An SNS fragment between positions 1740 and 1960 was sub-cloned into pGem4z, and DIG-UTP labeled sense or antisense cRNA generated. Sample preparation, hybridization, and visualization of in situ hybridization with alkaline phosphatase conjugated anti-DIG antibodies was carried out exactly as described in Schaeren-Wimers N. and Gerfin-Moser A. Histochemistry 100, 431-440 (1993).

Example 13 - Electrophysiological Properties of the Rat DRG Sodium Channel Expressed in *Xenopus oocytes*

pBluescript SK plasmid containing DNA encoding the SNS sodium channel was digested to position -21 upstream of the initiator methionine using a commercially available kit (Erase a base system, Promega, Madison, Wisconsin, USA). The linearized and digested plasmid was cut with Kpn1 and subcloned into an oocyte expression vector pSp64GL (Sma-Kpnl) sites. pSP64GL is derived from pSP64.T pSP64.T was cut with Sma1-EcoR1, blunt-ended with Klenow enzyme, and recircularized. Part of the pGem 72 (+) polylinker (Smal-Kpn1-EcoR1-Xhol) was ligated into the blunt-ended Bg1 II site of pSP64.T. This vector with an altered polylinker for DNA inserts (Smal-Kpn1-EcoR1-Xhol) and linearization (Sal1-Xba 1-BamH1) was named pSP64GL. The resulting plasmid was linearized with Xbal, and cRNA transcribed with SP6 polymerase using 1 mM 7-methylGppG.

cRNA (70 ng) was injected into *Xenopus* oocytes 7-14 days before recording; immature, stage IV oocytes were chosen cause of their smaller diameter and therefore capacitance. Oocytes were impaled with 3M KCl electrodes (≤1MΩ) and perfused at 3-4 ml per minute with modified Ringer solution containing 115 mM NaCl, 2.5 mM KCl, 10 mM HEPES, 1.8 mM MgCl₂, and 1 mM CaCl₂, pH 7.2, at temperature of 19.5 - 20.5 °C. Digital leak substraction of two electrode voltage-clamp current records was carried out using as leak currents produced by hyperpolarizing pulses of the same amplitude as the test depolarizing commands. Oocytes in which leak commands elicited time-dependent currents were discarded. Averages of 10 records were used for both test and leak.

Inward currents were evoked by depolarizing, in 10 mV steps, from -60 mV to a command potential of -20 to +40 mV in 10 mV steps and from -80 mV to a command potential of -30 to +2- mV in oocytes injected with sodium channel cRNA. Current traces are blanked for the first 1.5 ms from the onset of the voltage step to delete the capacity transients for clarity. The peak current is reached at the same command voltage for the two holding potentials, but is slightly smaller from -60 mV because of steady-state inactivation.

The effects of 50% or 100% replacement of external Na+ by N-methyl-D-glucosamine on the sodium channel current wer elicited by stepping the depolarizing currents given to the oocyte from -60 to +1 mV. Data were fitted with the equation $h_x = \frac{1}{2} \int_0^{\infty} \frac{1}{2} dt dt$

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 $1/(1 + \exp((V-V_{50})/k))$, where V is the prepulse potential, V_{50} the potential of 50% inactivation and the k the slope factor (best squares fit). The effect of TTX (10 μ M and 100 μ M) on the peak Na⁺ current (test pulse from -60 to +20 mV) was also determined. The effect was quickly reversible upon washout.

After a minimum incubation of 7 days from cRNA injection, step depolarizations to potentials positive to -30mV elicited inward currents which peaked between +10 and +20 mV with an average maximum amplitude of 164 ± 72 nA (from -60 mV holding potential, n = 13) and a reversal potential of $+35.5 \pm 2.2$ mV (n = 10). The inward current was reversed by total replacement of Na+ in the external medium with an impermeant cation (N-methyl-D-glucosamine). The current's reversal potential was shifted in 50% Na+ by 13.7 ± 3.2 mV in the hyperpolarizing direction (n = 3; predicted value for a Na+ -selective channel, 17.5 mV). The inactivation produced by a 1s prepulse was half-maximal at -30.0 ± 1.3 mV (slope factor 14.0 ± 1.7 mV, n = 5.

TTX had no effect at nanomolar concentrations, and produced only a 19.1 \pm 8.3% reduction at 10 μ M, n = 3). The estimated half-maximal inhibitory concentration (IC₅₀) was 59.6 \pm 10.1 μ M TTX.

The local anesthetic lignocaine was also weakly inhibitory, producing a maximum block of $41.7 \pm 5.4\%$ at 1 mM on the peak current elicited by depolarizing pulses from -60 mV to +10 mV (1 every min; n = 3), whereas under the same conditions 100 μ M phenytoin had no effect.

A similarity with the TTX-insensitive Na+ current of DRG neurons was the effectiveness and rank order of Pb²⁺ versus Cd²⁺ in reducing peak Na⁺ currents (-63.9 \pm 18.1% for Pb²⁺ versus -24.4 \pm 7.9% for Cd²⁺ at 50 μ M and 100 μ M, respectively; n = 3, P = 0.0189). The electrophysiological and pharmacological characteristics of the oocyte expressed DRG sodium channel are thus similar to the properties of the sensory neuron TTX-insensitive channel, given the constraints of expression in an oocyte system. In oocytes expressing the DRG sodium channel, the peak of the I/V plot occurred at a more depolarized potential than that of the DRG TTX-insensitive current, despite a similar reversal potential. This difference may reflect the absence of the accessory β 1 subunit found in DRG, which is known to shift activation to more negative potentials when

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expressed with the subunit of other Na⁺ channels. In addition, splice variants that exhibit an activation threshold more negative to SNS sodium channel may shift activation to the more negative potentials observed in sensory neurons.

Example 14 - Distribution of DRG Sodium Channel in Neonatal and Adult Rat Tissuesand Cell Lines

Northern blot and reverse transcriptase-polymerase chain reaction (RT-PCR) were used to examine neonatal and adult rat tissues for expression of the DRG sodium channel messenger RNA.

Random primed ³²P-labeled DNA Pst -Acc1 fragment probes (50 ng, specific activity 2 x 10⁹ c.p.m. per µg DNA) from interdomain region 1 (nucleotide position 1,478-1,892) of the SNS sodium channel nucleic acid sequence were used to probe total RNA extracted from tissues. The following tissues and cell lines were tested: central nervous system and non-neuronal tissues from neonatal rats; peripheral nervous tissue including neonatal Schwann cells and sympathetic neurons, as well as C6 glioma, human embryonal carcinoma line N-tera-2 and N-tera-2 neuro, rat sensory neuron-derived lines ND7 and ND8, and human neuroblastomas SMS-KCN and PC12 cells grown in the presence of NGF; adult rat tissue including pituitary, superior cervical ganglia, coeliac ganglia, trigeminal mesencephalic nucleus, vas deferens, bladder, ileum and DRG of adult animals treated with capsaicin (50 mg/kg) at birth and neonatal DRG control. Total RNA (10 µg) or 25 µg of RNA from tissues apart from superior cervical ganglion sample (10 µg) and capsaicin-treated adult rat DRG (5µg) were northern blotted.

Total RNA was separated on 1.2% agarose-formaldehyde gels, and capillary blotted onto Hibond-N filters (Amersham). The amounts of RNA on the blot were roughly equivalent, as judged by ethidium bromide staining of ribosomal RNA and by hybridization with the ubiquitously expressed L-27 ribosomal protein transcripts. Filters were prehybridized in 50% formamide, 5 x SSC containing 0.5% sodium dodecyl sulfate, 5 x Denhardts solution, 100 μg/ml boiled sonicated salmon sperm DNA (average size 300 bp),10 μg/ml poly-U and 10 μg/ml poly-C at 45°C for 6h. After 36 hours hybridization in the same conditions using 10⁷ c.p.m. per ml hybridization probe, the filters were briefly washed in 2 x SSC at room temperature, then twice with 2 x SSC with 0.5% SDS at 68°C

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for 15 min, followed by a 20 min wash in 0.5% SDS, 0.2 x SSC at 68°C. The filters were autoradiographed overnight or for 4 days on autoradiography film (Kodak X-omat).

For RT-PCR experiments, 10 µg total RNA from neonatal rat tissues (spleen, liver, kidney, lung, intestine, muscle, heart, superior cervical ganglia, spinal cord, brain stem, hippocampus, cerebellum, cortex and dorsal root ganglia), or 2 µg total RNA from control or capsaicin-treated rat DRG or DRG neurons in culture were treated with DNase I and extracted with acidic phenol to remove genomic DNA.

cDNA was synthesized with Superscript reverse transcriptase using oligo dT(12-18) primers and purified on Qiagen 5 tips. Polymerase chain reaction (PCR) was used to amplify cDNA (35 cycles, 94°C, 1 min; 55°C, 1 min; and 72°C, 1 min), and products separated on agarose gels before staining with ethidium bromide. L-27 primers (Ninkina et al. (1983) Nucleic Acids Res. 21, 3175-3182) were added to the PCR reaction 5 cycles after the start of the reaction with the DRG sodium channel specific primers which comprised

5'-CAGCTTCGCTCAGAAGTATCT-3' (SEQ ID NO: 9) and 5'-TTCTCGCCGTTCCACACGGAGA-3' (SEQ ID NO: 10).

Transcription of mRNA coding for the DRG sodium channel could not be detected in any non-neuronal tissues or in the central nervous system using northern blots or reverse transcription of mRNA and the polymerase chain reaction. Sympathetic neurons from the superior cervical ganglion and Schwann cell-containing sciatic nerve preparations, as well as several neuronal cell lines were also negative. However, total RNA extracts from neonatal and adult rat DRG gave a strong signal of size about 7kb on northern blots. These data suggest that the DRG sodium channel is not expressed only in early development.

RT-PCR of oligo dT-primed cDNA from various tissues using DRG sodium channel primers and L-27 ribosomal protein primer showed the presence of DRG sodium channel transcripts in DRG tissue only.

RT-PCR was also performed on DRG-sodium channel and L-27 transcripts from DRG neurons cultured and treated with capsaicin (overnight 10 µM) or dissected from neonatal animals treated with capsaicin (50 mg/kg on 2 consecutive days, followed by DRG isolation 5 days later. The signal from the L-27 probe was the same in capsaicintreated cell cultures or animals as compared with controls that were not treated with

capsaicin. There was a significant diminution in the DRG sodium channel signal from capsaicin-treated cultures or animals as compared with controls. Control PCR reactions without reverse transcriptase treatment were also done to control for contaminating genomic DNA.

When neonatal rats were treated with capsaicin and total adult DRG RNA subsequently examined by northern blotting, the signal was substantially reduced, suggesting that the DRG sodium channel transcript is expressed selectively by capsaicin-sensitive (predominantly nociceptive) neurons. These data were confirmed by RT-PCF experiments on both cultures of DRG neurons, and in whole animal studies.

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Example 15 - Distribution of DRG sodium channel in rat tissue by <u>in situ</u> hybridization

In situ hybridization was used to examine the expression of the DRG sodium channel transcripts at the single-cell level in both adult trigeninal ganglia and neonatal and adult rat DRG.

A SNS sodium channel PCR fragment of interdomain region I between positions 1,736 and 1,797 of the SNS sodium channel nucleic acid sequence was subcloned into pGem3Z (Promega, Madison, Wisconsin, USA) and digoxygenin (DIG)-UTP (Boehringer-Mannheim, Germany) labeled sense or antisense cRNA generated using SP6 or T7 polymerase, respectively. Sample preparation, hybridization and visualization of *in situ* hybridization with alkaline phosphatase conjugated anti-DIG antibodies was carried out as described in Schaeren-Wimers, et al., A. (1993) Histochemistry 100: 431-440, with the following modifications. Frozen tissue sections (10 µM-thick) of neonatal rat lumbar DRG, and adult trigeminal ganglion neurons were fixed for 10 min in phosphate buffered saline (PBS) containing 4% paraformaldehyde. Sections were acetylated in 0.1M triethanolamine, 0.25% acetic anhydride for 10 min. Prehybridization was carried out in 50% formamide, 4 x SSC, 100 µg/ml boiled and sonicated ssDNA, 50 µg/ml yeast tRNA, 2 x Denhardts solution at room temperature for 1 h. Hybridization was carried out overnight in the same buffer at 65°C. Probe concentration was 50 ng/ml. Sections were washed in 2 x SSC for 30 min at 72°C for 1 hr and twice in 0.1 SSC for 30 min at 72°C

before visualization at room temperature with anti-digoxygenin alkaline phosphatase conjugated antibodies. The same sections were then stained with mouse monoclonal antibody RT97 which is specific for neurofilaments found in large diameter neurons.

Subsets of sensory neurons from both tissues showed intense signals with a DRG sodium channel-specific probe. Combined immunohistochemistry with the large-diameter neuron-specific monoclonal antibody RT97 and the DRG sodium channel specific probe showed that most of the large diameter neurons did not express the DRG sodium channel transcript. Small diameter neurons were stained with the DRG sodium channel specific probe but not the large diameter neurons.

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Example 16 - Site Directed Mutagenesis of SNS Sodium Channel - TTX Sensitivity

The SNS sodium channel is 65% homologous to the tetrodotoxin-insensitive cardiac sodium channel. A number of residues that line the channel atrium have been implicated in tetrodotoxin binding. The amino acid sequence of the SNS sodium channel exhibits sequence identity to other tetrodotoxin-sensitive sodium channels in 7 out of 9 such residues. One difference is a conservative substitution at D(905)E. A single residue (C-357) has been shown to play a critical role in tetrodotoxin binding to the sodium channel. In the SNS sodium channel, a hydrophilic serine is found at this position, whereasa other sodium channels that are sensitive to TTX have phenylalanine in this position.

Site-directed mutagenesis using standard techniques and primers having the sequence TGACGCAGGACTCCTGGGAGCGCC (SEQ ID NO: 31) was used to substitute phenylalanine for serine at position 357 in the SNS sodium channel. The mutated SNS sodium channel, when expressed in *Xenopus* oocytes produces voltage-gated currents similar in amplitude and time course to the native channel. However, sensitivity to TTX is restored to give an IC₅₀ of 2.5 nM (+-0.4, n = 5), similar to other voltage-gated sodium channels that have aromatic residues at the equivalent position. The table below shows IC₅₀ for SNS sodium channel, and the rat brain iia, muscle type 1, and cardiac tetrodotoxin-insensitive sodium channels.

TTX Sensitivity

Sodium Channel	ss1 domain	ss2 domain	IC ₅₀
<u> </u>			

Rat brain iia	FRLM	TQDFWENLY	18 nM
muscle type 1	FRLM	TQDYWENLY	40 nM
cardiac TTXi	FRLM	TQDCWERLY	950 nM
SNS	FRLM	TQDSWERLY	60 micromolar
SNS mutant	FRLM .	TQDFWERLY	2.5 nM

FRLM - SEQ ID NO: 11; TQDFWENLY - SEQ ID NO: 12;

TQDYWENLY - SEQ ID NO: 13; TQDCWERLY - SEQ ID NO:14;

TQDSWERLY - SEQ ID NO: 15; TQDFWERLY - SEQ ID NO:16

Example 18

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Polyclonal antibodies were raised in rabbits against the following peptides derived from the SNS sodium channel protein amino acid sequence:

Peptide 1 TQDSWER (SEQ ID NO:17)

Peptide 2 GSTDDNRSPQSDPYN (SEQ ID NO: 18)

Peptide 3 SPKENHGDFI (SEQ ID NO: 19)

Peptide 4 PNHNGSRGN (SEQ ID NO: 20)

The peptides were conjugated to Keyhole limpet heocyanin (KLH) and injected repeatedly into rabbits. Sera from the rabbits was treated by Western blotting. Several sera showed positive results indicating the presence of antibodies specific for the peptide in the sera.

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SEQUENCE LISTING

5	(1) GENE	RAL INFORMATION:	
_	(i)	APPLICANT: (A) NAME: University College London (B) STREET: Gower Street (C) CITY: London	
LO		(E) COUNTRY: England (F) POSTAL CODE (ZIP): WC1E 6BT	
	(ii)	TITLE OF INVENTION: Ion Channel	
L5	(iii)	NUMBER OF SEQUENCES: 31	
20	(iv)	COMPUTER READABLE FORM: (A) MEDIUM TYPE: Floppy disk (B) COMPUTER: IBM PC compatible (C) OPERATING SYSTEM: PC-DOS/MS-DOS (D) SOFTWARE: PatentIn Release #1.0, Version #1.25	
_	(2) INFO	RMATION FOR SEQ ID NO:1:	
25	(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 6524 base pairs (B) TYPE: number of the control of the c	
30		(C) STRANDEDNESS: single (D) TOPOLOGY: linear	
	(ii)	MOLECULE TYPE: cDNA	
35	(ix)	FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 2046077	
10	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:1:	
	TAGCTTGC	TT CTGCTAATGC TACCCCAGGC CTTTAGACAG AGAACAGATG GCAGATGGAG	60
15	TTTCTTAT	TG CCATGCGCAA ACGCTGAGCC CACCTCATGA TCCCGGACCC CATGGTTTTC	120
	AGTAGACA	AC CTGGGCTAAG AAGAGATCTC CGACCTTATA GAGCAGCAAA GAGTGTAAAT	180
	TCTTCCCC	AA GAAGAATGAG AAG ATG GAG CTC CCC TTT GCG TCC GTG GGA Met Glu Leu Pro Phe Ala Ser Val Gly	230
50		1 5	
		AAT TTC AGA CGG TTC ACT CCA GAG TCA CTG GCA GAG ATC GAG Asn Phe Arg Arg Phe Thr Pro Glu Ser Leu Ala Glu Ile Glu 15 20 25	278
55		ATT GCT GCT CAC CGC GCA GCC AAG AAG GCC AGA ACC AAG CAC Ile Ala Ala His Arg Ala Ala Lys Lys Ala Arg Thr Lys His 30 35 40	326
50		CAG GAG GAC AAG GGC GAG AAG CCC AGG CCT CAG CTG GAC TTG Gln Glu Asp Lys Gly Glu Lys Pro Arg Pro Gln Leu Asp Leu 45 50 55	374

										-51-								
	AAA Lys	GAC Asp	TGT Cys 60	Asn	CAG Gln	CTG Leu	CCC Pro	AAG Lys 65	TTC Phe	TAT Tyr	GGT Gly	GAG Glu	CTC Leu 70	CCA Pro	GCA Ala	GAA Glu	422	2
5	CTG Leu	GTC Val 75	GGG Gly	GAG Glu	CCC Pro	CTG Leu	GAG Glu 80	GAC Asp	CTA Leu	GAC Asp	CCT Pro	TTC Phe 85	TAC Tyr	AGC Ser	ACA Thr	CAC His	470)
10	CGG Arg 90	ACA Thr	TTC Phe	ATG Met	GTG Val	TTG Leu 95	AAT Asn	AAA Lys	AGC Ser	AGG Arg	ACC Thr 100	ATT Ile	TCC Ser	AGA Arg	TTC Phe	AGT Ser 105	518	3
15	GCC Ala	ACT Thr	TGG Trp	GCC Ala	CTG Leu 110	TGG Trp	CTC Leu	TTC Phe	AGT Ser	CCC Pro 115	TTC Phe	AAC Asn	CTG Leu	ATC Ile	AGA Arg 120	AGA Arg	566	5
20	ACA Thr	GCC Ala	ATC Ile	AAA Lys 125	GTG Val	TCT Ser	GTC Val	CAT His	TCC Ser 130	TGG Trp	TTC Phe	TCC Ser	ATA Ile	TTC Phe 135	ATC Ile	ACC Thr	614	Ī
	ATC Ile	ACT Thr	ATT Ile 140	TTG Leu	GTC Val	AAC Asn	TGC Cys	GTG Val 145	TGC Cys	ATG Met	ACC Thr	CGA Arg	ACT Thr 150	GAT Asp	CTT Leu	CCA Pro	662	<u>:</u>
25	GAG Glu	AAA Lys 155	GTC Val	GAG Glu	TAC Tyr	GTC Val	TTC Phe 160	ACT Thr	GTC Val	ATT Ile	TAC Tyr	ACC Thr 165	TTC Phe	GAG Glu	GCT Ala	CTG Leu	- 710)
30	ATT Ile 170	AAG Lys	ATA Ile	CTG Leu	GCA Ala	AGA Arg 175	GGG Gly	TTT Phe	TGT Cys	CTA Leu	AAT Asn 180	GAG Glu	TTC Phe	ACT Thr	TAT Tyr	CTT Leu 185	758	į
35	Arg	Asp	Pro	Trp	Asn 190	TGG Trp	Leu	Asp	Phe	Ser 195	Val	Ile	Thr	Leu	Ala 200	Tyr	806	j
40	GTG Val	GGT Gly	GCA Ala	GCG Ala 205	ATA Ile	GAC Asp	CTC Leu	CGA Arg	GGA Gly 210	ATC Ile	TCA Ser	GGC Gly	CTG Leu	CGG Arg 215	ACA Thr	TTC Phe	854	,
	CGA Arg	GTT Val	CTC Leu 220	AGA Arg	GCC Ala	CTG Leu	AAA Lys	ACT Thr 225	GTT Val	TCT Ser	GTG Val	ATC Ile	CCA Pro 230	GGA Gly	CTG Leu	AAG Lys	902	
45	GTC Val	ATC Ile 235	GTG Val	GGA Gly	GCC Ala	CTG Leu	ATC Ile 240	CAC His	TCA Ser	GTG Val	AGG Arg	AAG Lys 245	CTG Leu	GCC Ala	GAC Asp	GTG Val	950	
50	ACT Thr 250	ATC Ile	CTC Leu	ACA Thr	GTC Val	TTC Phe 255	TGC Cys	CTG Leu	AGC Ser	GTC Val	TTC Phe 260	GCC Ala	TTG Leu	GTG Val	GGC Gly	CTG Leu 265	998	
55						AAC Asn											1046	
60	Asp	Pro	His	Lys 285	Ala	GAC Asp	Asn	Leu	Ser 290	Ser	Glu	Met	Ala	Glu 295	Tyr	Val	1094	
-						ACT Thr											1142	÷

		GCT Ala 315															1190
5		CCG Pro															1238
10		TCA Ser															1286
15		CAG Gln															1334
20		GTT Val															1382
		GTC Val 395															1430
25		GAA Glu															1478
30		GAA Glu															1526
35		TCC Ser															1574
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		TCA Ser 475															1670
45	Gly 490	CTG Leu	Ser	Ser	Gly	Arg 495	Arg	Arg	Ala	Ser	His 500	Gly	Ser	Val	Phe	His 505	1718
50	Phe	CGA Arg	Ala	Pro	Ser 510	Gl'n	Asp	Ile	Ser	Phe 515	Pro	Asp	Gly	Ile	Thr 520	Pro	1766
55		GAT Asp															1814
60		CTG Leu															1862
	CTG Leu	CCT Pro 555	CAG Gln	TCC Ser	CCC Pro	AAC Asn	CCT Pro 560	GGC Gly	CGT Arg	AG. Arg	CAT His	GGA Gly 565	GAA Glu	GAG Glu	GGA Gly	CAG Gln	1910

	CTC Leu 570	GGA Gly	GTG Val	CCC	ACT Thr	GGT Gly 575	GAG Glu	CTT	ACC Thr	GCT Ala	GGA Gly 580	GCG Ala	CCT Pro	GAA Glu	GGC Gly	CCG Pro 585		1958
5	Ala	Leu	His	Thr	ACA Thr 590	Gly	Gln	Lys	Ser	Phe 595	Leu	Ser	Ala	Gly	Tyr 600	Leu		2006
10	Asn	Glu	Pro	Phe 605	CGA Arg	Ala	Gln	Arg	Ala 610	Met	Ser	Val	Val	Ser 615	Ile	Met		2054
15	Thr	Ser	Val 620	Ile	GAG Glu	Glu	Leu	Glu 625	Glu	Ser	Lys	Leu	Lys 630	Суѕ	Pro	Pro		2102
20	TGC Cys	TTG Leu 635	ATC Ile	AGC Ser	TTC Phe	GCT Ala	CAG Gln 640	AAG Lys	TAT Tyr	CTG Leu	ATC Ile	TGG Trp 645	GAG Glu	TGC Cys	TGC Cys	CCC Pro		2150
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35	CAA Gln	GCC Ala	GGC Gly 700	AAC Asn	ATT Ile	GTC Val	TTC Phe	ACC Thr 705	GTG Val	TTT Phe	TTC Phe	ACA Thr	ATG Met 710	GAG Glu	ATG Met	GCC Ala		2342
40	TTC Phe	AAG Lys 715	ATC Ile	ATT Ile	GCC Ala	TTC Phe	GAC Asp 720	CCC Pro	TAC Tyr	TAT Tyr	TAC Tyr	TTC Phe 725	CAG Gln	AAG Lys	AAG Lys	TGG Trp	-	2390
	AAT Asn 730	ATC Ile	TTC Phe	GAC Asp	TGT Cys	GTC Val 735	ATC Ile	GTC Val	ACC Thr	GTG Val	AGC Ser 740	CTT Leu	CTG Leu	GAG Glu	CTG Leu	AGT Ser 745		2438
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50	CGG Arg	GTC Val	TTC Phe	AAG Lys 765	CTG Leu	GCC Ala	AAG Lys	TCC Ser	TGG Trp 770	CCC Pro	ACC Thr	CTG Leu	AAC Asn	ACC Thr 775	CTC Leu	ATC Ile		2534
55	AAG Lys	ATC Ile	ATC Ile 780	GGG Gly	AAC Asn	TCA Ser	GTG Val	GGG Gly 785	GCC Ala	CTG Leu	GGC Gly	AAC Asn	CTG Leu 790	ACC Thr	TTT Phe	ATC Ile		2582
60	CTG Leu	GCC Ala 795	ATC Ile	ATC Ile	GTC Val	TTC Phe	ATC Ile 800	TTC Phe	GCC Ala	CTG Leu	GTC Val	GGA Gly 805	AAG Lys	CAG Gln	CTT Leu	CTC Leu		2630
	TCA Ser 810	GAG Glu	GAC Asp	TAC Tyr	GGG Gly	TGC Cys 815	CGC Arg	AAG Lys	GAC Asp	GGC Gly	GTC Val 820	TCC Ser	GTG Val	TGG Trp	AAC Asn	GGC Gly 825		2678

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	GA0 Glu	3 AA 1 Ly	G CT(s Le	C CG	TGG Tri 830	o His	ATG Met	TG7	GA(TTO Pho 83	e Pho	C CA e Hi	T TC s Se	C TT r Ph	C CI e Le 84	G GTC u Val 0		2726
5	GT(Va]	TTC Pho	C CGA e Arg	A ATO E Ile 845	e Lei	TGC Cys	GGG Gly	GAG Glu	TGC Trp 850	o Ile	C GA0 ⊇ Glu	G AA	C ATO	G TG	p Va	C TGC 1 Cys		2774
10	ATC Met	GAC Glu	G GTC 1 Val 860	. Sei	CAC Glr	AAA Lys	TCC Ser	ATC Ile 865	Cys	CTC Leu	ATC	CTC	TTC Phe 870	e Le	G AC	T GTG r Val		2822
15	ATG Met	Va] 875	. Leu	GGC Gly	AAC Asn	CTA Leu	GTG Val 880	GTG Val	CTC Leu	AAC Asn	CTI Leu	TTC Phe 885	e Ile	GC:	TTI a Lei	A CTG u Leu		2870
20	CTG Leu 890	ASI.	TCC Ser	TTC Phe	AGC Ser	GCG Ala 895	GAC Asp	AAC Asn	CTC Leu	ACG Thr	GCT Ala 900	Pro	GAG Glu	GA1 Asp	GAC Asi	GGG Gly 905		2918
	GAG Glu	GTG Val	AAC Asn	AAC Asn	TTG Leu 910	CAG Gln	TTA Leu	GCA Ala	CTG Leu	GCC Ala 915	Arg	ATC	CAG Gln	GTA Val	CTI Leu 920	GGC Gly		2966
25	CAT His	CGG	GCC Ala	AGC Ser 925	AGG Arg	GCC Ala	AGC Ser	GCC Ala	AGT Ser 930	TAC Tyr	ATC Ile	AGC Ser	AGC Ser	CAC His 935	Cys	CGA Arg		3014
30	TTC Phe	CAC His	TGG Trp 940	CCC Pro	AAG Lys	GTG Val	GAG Glu	ACC Thr 945	CAG Gln	CTG Leu	GGC Gly	ATG Met	AAG Lys 950	CCC Pro	CCA Pro	CTC Leu		3062
35	ACC Thr	AGC Ser 955	TCA Ser	GAG Glu	GCC Ala	AAG Lys	AAC Asn 960	CAC His	ATT Ile	GCC Ala	ACT Thr	GAT Asp 965	GCT Ala	GTC Val	AGT Ser	GCT Ala		3110
40	GCA Ala 970	GTG Val	GGG Gly	AAC Asn	CTG Leu	ACA Thr 975	AAG Lys	CCA Pro	GCT Ala	CTC Leu	AGT Ser 980	AGC Ser	CCC Pro	AAG Lys	GAG Glu	AAC Asn 985		3158
	CAC His	GGG Gly	GAC Asp	TTC Phe	ATC Ile 990	ACT Thr	GAT Asp	CCC Pro	AAC Asn	GTG Val 995	TGG Trp	GTC Val	TCT Ser	GTG Val	CCC Pro	Ile		3206
45	GCT Ala	GAG Glu	GGG Gly	GAA Glu 1005	Ser	GAC Asp	CTC (Leu ,	Asp (GAG Glu 1010	Leu	GAG Glu	GAA Glu	GAT Asp	ATG Met 1015	Glu	CAG Gln		3254
50	GCT Ala	TCG Ser	CAG Gln 1020	ser	TCC Ser	TGG (Trp (GIN (GAA (Glu (1025	GAG Glu	GAC Asp	CCC Pro	AAG Lys	GGA Gly 1030	Gln	CAG Gln	GAG Glu	:	3302
55	CAG :	TTG Leu 1035	Pro	CAA Gln	GTC Val	Gln 1	AAG 1 Lys (rgt (Cys (GAA . Glu .	AAC Asn	His	CAG Gln 1045	Ala	GCC Ala	AGA Arg	AGC Ser	;	3350
60	CCA (Pro 1	GCC Ala	TCC . Ser !	ATG Met	met .	TCC : Ser :	CT C	GAG (Glu A	GAC (Asp 1	Leu .	GCT (Al a 1060	CCA Pro	TAC Tyr	CTG Leu	GGT Gly	GAG Glu 1065	3	3398
	AGC ' Ser '	TGG Trp	AAG / Lys /	arg .	AAG (Lys 1	GAT A	AGC C Ser F	ro C	Sln (GTC (/al L075	CCT (Pro <i>i</i>	GCC (Ala (GAG (Glu (Gly	GTG Val 1080	Asp	3	3446

	GAC AC Asp Th	G AGC r Ser	TCC Ser 1085	Ser	GAG Glu	GGC Gly	AGC Ser	ACG Thr 109	Val	GAC Asp	TGC Cys	CCG Pro	GAC Asp 109	Pro	GAG Glu	3494
5	GAA AT Glu Il	C CTG e Leu 110	Arg	AAG Lys	ATC Ile	CCC Pro	GAG Glu 110	Leu	GCA Ala	CAT His	GAC Asp	CTG Leu 111	Asp	GAG Glu	CCC Pro	3542
10	GAT GA Asp As 11	C TGT p Cys 15	TTC Phe	AGA Arg	GAA Glu	GGC Gly 1120	Cys	ACT Thr	CGC Arg	CGC Arg	TGT Cys 1125	Pro	TGC Cys	TGC Cys	AAC Asn	3590
15	GTG AA Val As 1130	n Thr	Ser	Lys	Ser 1135	Pro	Trp	Ala	Thr	Gly 1140	Trp	Gln	Val	Arg	Lys 1145	3638
20	ACC TG Thr Cy	C TAC s Tyr	CGC Arg	ATC Ile 1150	Val	GAG Glu	CAC His	AGC Ser	TGG Trp 1155	Phe	GAG Glu	AGT Ser	TTC Phe	ATC Ile 1160	Ile	3686
	TTC ATC	G ATC	CTG Leu 1165	Leu	AGC Ser	AGT Ser	GGA Gly	GCG Ala 1170	Leu	GCC Ala	TTT Phe	GAG Glu	GAT Asp 1175	Asn	TAC Tyr	3734
25	CTG GAL Leu Gl	1 Glu 118	Lys	Pro	Arg	Val	Lys 1185	Ser	Val	Leu	Glu	Туг 1190	Thr	Asp	Arg	3782
30	GTG TTC Val Pho 11:	e Thr 95	Phe	Ile	Phe	Val 1200	Phe	Glu	Met	Leu	Leu 1205	Lys	Trp	Val	Ala	3830
35	TAT GGG Tyr Gly 1210	y Phe	Lys	Lys	Туг 1215	Phe	Thr	Asn	Ala	Trp 1220	Cys)	Trp	Leu	Asp	Phe 1225	3878
40	CTC AT	r GTG ∋ Val	AAC Asn	ATC Ile 1230	Ser	CTG	ACA Thr	AGC Ser	CTC Leu 1235	Ile	GCG Ala	AAG Lys	ATC Ile	CTT Leu 1240	Glu	3926
	TAT TCC	C GAC r Asp	GTG Val 1245	Ala	TCC Ser	ATC Ile	AAA Lys	GCC Ala 1250	Leu	CGG Arg	ACT Thr	CTC Leu	CGT Arg 1255	Ala	CTC Leu	39 74
45	CGA CCC	CTG Leu 126	Arg	GCT Ala	CTG Leu	TCT Ser	CGA Arg 1265	Phe	GAA Glu	GGC Gly	ATG Met	AGG Arg 1270	Val	GTG Val	GTG Val	4022
50	GAT GCC Asp Ala 12	a Leu	GTG Val	GGC Gly	GCC Ala	ATC Ile 1280	Pro	TCC Ser	ATC Ile	ATG Met	AAC Asn 1285	Val	CTC Leu	CTC Leu	GTC Val	4070
55	TGC CTC Cys Let 1290	C ATC	TTC Phe	TGG Trp	CTC Leu 1295	Ile	TTC Phe	AGC Ser	ATC Ile	ATG Met 1300	Gly	GTG Val	AAC Asn	CTC Leu	TTC Phe 1305	4118
60	GCC GGG Ala Gl	G AAA / Lys	TTT Phe	TCG Ser 1310	Lys	TGC Cys	GTC Val	GAC Asp	ACC Thr 1315	Arg	AAT Asn	AAC Asn	CCA Pro	TTT Phe 1320	Ser	4166
	AAC GTO	G AAT L Asn	TCG Ser 1325	Thr	ATG Met	GTG Val	AAT Asn	AAC Asn 1330	Lys	TCC Ser	GAG Glu	TGT Cys	CAC His 1335	Asn	CAA Gln	4214

	AAC Asn	AGC Ser	ACC Thr 1340	Gly	CAC His	TTC Phe	TTC Phe	TGG Trp 1345	Val	AAC Asn	GTC Val	AAA Lys	GTC Val 1350	Asn	TTC Phe	GAC Asp	4262
5	AAC Asn	GTC Val 1355	Ala	ATG Met	GGC Gly	TAC Tyr	CTC Leu 1360	Ala	CTT Leu	CTT Leu	CAG Gln	GTG Val 1365	Ala	ACC Thr	TTC Phe	AAA Lys	4310
10	GGC Gly 1370	Trp	ATG Met	GAC Asp	ATA Ile	ATG Met 1375	Tyr	GCA Ala	GCT Ala	GTT Val	GAT Asp 1380	Ser	GGA Gly	GAG Glu	ATC Ile	AAC Asn 1385	4358
15	AGT Ser	CAG Gln	CCT Pro	AAC Asn	TGG Trp 1390	Glu	AAC Asn	AAC Asn	TTG Leu	TAC Tyr 1395	Met	TAC Tyr	CTG Leu	TAC Tyr	TTC Phe 1400	Val	4406
20	GTT Val	TTC Phe	ATC Ile	ATT Ile 1405	Phe	GGT Gly	GGC Gly	TTC Phe	TTC Phe 1410	Thr	CTG Leu	AAT Asn	CTC Leu	TTT Phe 141	Val	GGG Gly	4454
20	GTC Val	ATA Ile	ATC Ile 1420	Asp	AAC Asn	TTC Phe	AAC Asn	CAA Gln 1425	Gln	AAA Lys	AAA Lys	AAG Lys	CTA Leu 1430	Gly	GGC Gly	CAG Gln	4502
25	GAC Asp	ATC Ile 1435	Phe	ATG Met	ACA Thr	GAA Glu	GAG Glu 1440	Gln	AAG Lys	AAG Lys	TAC Tyr	TAC Tyr 144	AAT Asn	GCC Ala	ATG Met	AAG Lys	4550
30	AAG Lys 1450	Leu	GGC Gly	TCC Ser	AAG Lys	AAA Lys 145	Pro	CAG Gln	AAG Lys	CCC Pro	ATC Ile 1460	Pro	CGG Arg	CCC Pro	CTG Leu	AAT Asn 1465	4598
35	AAG Lys	TAC Tyr	CAA Gln	GGC Gly	TTC Phe 147	Val	TTT Phe	GAC Asp	ATC Ile	GTG Val 147	Thr	AGG Arg	CAA Gln	GCC Ala	TTT Phe 148	Asp	4646
40	ATC Ile	ATC Ile	ATC Ile	ATG Met 148	Val	CTC Leu	ATC Ile	TGC Cys	CTC Leu 149	Asn	ATG Met	ATC Île	ACC Thr	ATG Met 149	Met	GTG Val	4694
40	GAG Glu	ACC Thr	GAC Asp 150	Glu	CAG Gln	GGC Gly	GAG Glu	GAG Glu 150	Lys	ACG Thr	AAG Lys	GTT Val	CTG Leu 151	Gly	AGA Arg	ATC Ile	4742
45	AAC Asn	CAG Gln 151	Phe	TTT Phe	GTG Val	GCC Ala	GTC Val 152	Phe	ACG Thr	GGC Gly	GAG Glu	TGT Cys 152	GTG Val 5	ATG Met	AAG Lys	ATG Met	4790
50	TTC Phe 153	Ala	CTG Leu	CGA Arg	CAG Gln	TAC Tyr 153	Tyr	TTC Phe	ACC Thr	AAC Asn	GGC Gly 154	Trp	AAC Asn	GTG Val	TTC Phe	GAC Asp 1545	4838
55	TTC Phe	ATA Ile	GTG Val	GTG Val	ATC Ile 155	Leu	TCC Ser	ATT Ile	GGG Gly	AGT Ser 155	Leu	CTG Leu	TTT Phe	TCT Ser	GCA Ala 156	Ile	4886
66	CTT Leu	AAG Lys	TCA Ser	CTG Leu 156	Glu	AAC Asn	TAC Tyr	TTC Phe	TCC Ser 157	Pro	ACG Thr	CTC Leu	TTC Phe	CGG Arg 157	Val	ATC Ile	4934
60	CGT Arg	CTG Leu	GCC Ala 158	Arg	ATC Ile	GGC Gly	CGC Arg	ATC Ile 158	Leu	AGG Arg	CTG Leu	ATC Ile	CGA Arg 159	Ala	GCC Ala	AAG Lys	4982

	GGG ATT Gly Ile 159	Arg					Ala					Leu				5030
5	TTC AAC Phe Asn 1610					Leu					Phe					5078
10	TTC GGC Phe Gly				Phe					Asp					Asp	5126
15	GAC ATG Asp Met			Phe					Asn					Leu		5174
20	CAG ATC		Thr					Asp					Pro			5222
	AAC ACG Asn Thr 167	Gly					Asp					Asn				5270
25	TCC CGG Ser Arg 1690					Ser					Ile					5318
30	ACC TAC				Ser					Val					Ala 👵	5366
35	GTG ATT Val Ile			Asn					Thr					Glu		5414
40	CTG AGC Leu Ser		Asp					Phe					Glu			5462
	GAC CCG Asp Pro	Glu					Ile					Leu				5510
45	GCG GAC Ala Asp 1770	ACG Thr	CTC Leu	TCC Ser	GGC Gly 1775	Pro	CTT Leu	AGA Arg	ATC Ile	CCC Pro 1780	Lys	CCC Pro	AAC Asn	CAG Gln	AAT Asn 1785	5558
50	ATA TTA				Asp					Pro					His	5606
55	TGT CTC			Leu					Lys					Glu		5654
60	GGG GAG		Asp					Asn					Phe			5702
•	ACC AAT Thr Ass 183	ı Leu					Tyr					Thr				5750

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	TGG AAG CAG GAA GAC CTC TCA GCC ACA GTC ATT CAA AAG GCC TAC CGG Trp Lys Gln Glu Asp Leu Ser Ala Thr Val Ile Gln Lys Ala Tyr Arg 1850 1855 1860 1865	5798
5	AGC TAC ATG CTG CAC CGC TCC TTG ACA CTC TCC AAC ACC CTG CAT GTG Ser Tyr Met Leu His Arg Ser Leu Thr Leu Ser Asn Thr Leu His Val 1870 1875 1880	5846
10	CCC AGG GCT GAG GAG GAT GGC GTG TCA CTT CCC GGG GAA GGC TAC ATT Pro Arg Ala Glu Glu Asp Gly Val Ser Leu Pro Gly Glu Gly Tyr Ile 1885 1890 1895	5894
15	ACA TTC ATG GCA AAC AGT GGA CTC CCG GAC AAA TCA GAA ACT GCC TCT Thr Phe Met Ala Asn Ser Gly Leu Pro Asp Lys Ser Glu Thr Ala Ser 1900 1905 1910	5942
20	GCT ACG TCT TTC CCG CCA TCC TAT GAC AGT GTC ACC AGG GGC CTG AGT Ala Thr Ser Phe Pro Pro Ser Tyr Asp Ser Val Thr Arg Gly Leu Ser 1915 1920 1925	5990
20	GAC CGG GCC AAC ATT AAC CCA TCT AGC TCA ATG CAA AAT GAA GAT GAG Asp Arg Ala Asn Ile Asn Pro Ser Ser Met Gln Asn Glu Asp Glu 1930 1940 1945	6038
25	GTC GCT GCT AAG GAA GGA AAC AGC CCT GGA CCT CAG TGAAGGCACT Val Ala Ala Lys Glu Gly Asn Ser Pro Gly Pro Gln 1950	6084
30	CAGGCATGCA CAGGGCAGGT TCCAATGTCT TTCTCTGCTG TACTAACTCC TTCCCTCTGG	6144
30	AGGTGGCACC AACCTCCAGC CTCCACCAAT GCATGTCACT GGTCATGGTG TCAGAACTGA	6204
	ATGGGGACAT CCTTGAGAAA GCCCCCACCC CAATAGGAAT CAAAAGCCAA GGATACTCCT	6264
35	CCATTCTGAC GTCCCTTCCG AGTTCCCAGA AGATGTCATT GCTCCCTTCT GTTTGTGACC	6324
	AGAGACGTGA TTCACCAACT TCTCGGAGCC AGAGACACAT AGCAAAGACT TTTCTGCTGG	6384
40	TGTCGGGCAG TCTTAGAGAA GTCACGTAGG GGTTGGTACT GAGAATTAGG GTTTGCATGA	6444
	CTGCATGCTC ACAGCTGCCG GACAATACCT GTGAGTCGGC CATTAAAATT AATATTTTTA	6504
	AAGTTAAAAA AAAAAAAAA	6524
45	(2) INFORMATION FOR SEQ ID NO:2:	
50	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 1957 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: protein	
55	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:	
	Met Glu Leu Pro Phe Ala Ser Val Gly Thr Thr Asn Phe Arg Arg Phe 1 5 10 15	
60	Thr Pro Glu Ser Leu Ala Glu Ile Glu Lys Gln Ile Ala Ala His Arg 20 25 30	
	Ala Ala Lys Lys Ala Arg Thr Lys His Arg Gly Gln Glu Asp Lys Gly 35 40 45	

	Glu	Lys 50	Pro	Arg	Pro	Gln	Leu 55	Asp	Leu	Lys	Asp	Cys 60	Asn	Gln	Leu	Pro
5	Lys 65	Phe	Tyr	Gly	Glu	Leu 70	Pro	Ala	Glu	Leu	Val 75	Gly	Glu	Pro	Leu	Glu 80
10	Asp	Leu	Asp	Pro	Phe 85	Tyr	Ser	Thr	His	Arg 90	Thr	Phe	Met	Val	Leu 95	Asn
	Lys	Ser	Arg	Thr 100	Ile	Ser	Arg	Phe	Ser 105	Ala	Thr	Trp	Ala	Leu 110	Trp	Leu
15	Phe	Ser	Pro 115	Phe	Asn	Leu	Ile	Arg 120	Arg	Thr	Ala	Ile	Lys 125	Val	Ser	Val
	His	Ser 130	Trp	Phe	Ser	Ile	Phe 135	Ile	Thr	Ile	Thr	Ile 140	Leu	Val	Asn	Cys
20	Val 145	Cys	Met	Thr	Arg	Thr 150	Asp	Leu	Pro	Glu	Lys 155	Val	Glu	Tyr	Val	Phe 160
25	Thr	Val	Ile	Tyr	Thr 165	Phe	Glu	Ala	Leu	Ile 170	Lys	Ile	Leu	Ala	Arg 175	Gly
23	Phe	Cys	Leu	Asn 180	Glu	Phe	Thr	Tyr	Leu 185	Arg	Asp	Pro	Trp	Asn 190	Trp	Leu
30	Asp	Phe	Ser 195	Val	Ile	Thr	Leu	Ala 200	Tyr	Val	Gly	Ala	Ala 205	Ile	Asp	Leu
	Arg	Gly 210	Ile	Ser	Gly	Leu	Arg 215	Thr	Phe	Arg	Val	Leu 220	Arg	Ala	Leu	Lys
35	Thr 225	Val	Ser	Val	Ile	Pro 230	Gly	Leu	Lys	Val	Ile 235	Val	Gly	Ala	Leu	Ile 240
40	His	Ser	Val	Arg	Lys 245	Leu	Ala	Asp	Val	Thr 250	Ile	Leu	Thr	Val	Phe 255	Cys
	Leu	Ser	Val	Phe 260	Ala	Leu	Val	Gly	Leu 265	Gln	Leu	Phe	Lys	Gly 270	Asn	Leu
45	Lys	Asn	Lys 275	Cys	Ile	Arg	Asn	Gly 280	Thr	Asp	Pro	His	Lys 285	Ala	Asp	Asn
	Leu	Ser 290	Ser	Glu	Met	Ala	Glu 295	Tyr	Val	Ser	Ile	Lys 300	Pro	Gly	Thr	Thr
50	Asp 305	Pro	Leu	Leų	Cys	Gly 310	Asn	Gly	Ser	Asp	Ala 315	Gly	His	Cys	Pro	Gly 320
55	Gly	Tyr	Val	Cys	Leu 325	Lys	Thr	Pro	Asp	Asn 330	Pro	Asp	Phe	Asn	Tyr 335	Thr
	Ser	Phe	Asp	Ser 340	Phe	Āla	Trp	Ala	Phe 345	Leu	Ser	Leu	Phe	Arg 350	Leu	Met
60	Thr	Gln	Asp 355	Ser	Trp	Glu	Arg	Leu 360	Tyr	Gln	Gln	Thr	Leu 365	Arg	Ala	Ser
	Gly	Lys 370	Met	Tyr	Met	Val	Phe 375	Phe	Val	Leu	Val	Ile 380	Phe	Leu	Gly	Ser

	Phe 385	Tyr	Leu	Val	Asn	Leu 390	Île	Leu	Ala	Val	Val 395	Thr	Met	Ala	Tyr	Glu 400
5	Glu	Gln	Ser	Gln	Ala 405	Thr	Ile	Ala	Glu	Ile 410	Glu	Ala	Lys	Glu	Lys 415	Lys
	Phe	Gln	Glu	Ala 420	Leu	Glu	Val	Leu	Gln 425	Lys	Glu	Gln	Glu	Val 430	Leu	Ala
10	Ala	Leu	Gly 435	Ile	Asp	Thr	Thr	Ser 440	Leu	Gln	Ser	His	Ser 445	Gly	Ser	Pro
15	Leu	Ala 450	Ser	Lys	Asn	Ala	Asn 455	Glu	Aṛg	Arg	Pro	Arg 460	Val	Lys	Ser	Arg
13	Val 465	Ser	Glu	Gly	Ser	Thr 470	Asp	Asp	Asn	Arg	Ser 475	Pro	Gln	Ser	Asp	Pro 480
20	Tyr	Asn	Gln	Arg	Arg 485	Met	Ser	Phe	Leu	Gly 490	Leu	Ser	Ser	Gly	Arg 495	Arg
	Arg	Ala	Ser	His 500	Gly	Ser	Val	Phe	His 505	Phe	Arg	Ala	Pro	Ser 510	Gln	Asp
25	Ile	Ser	Phe 515	Pro	Asp	Gly	Ile	Thr 520	Pro	Asp	Asp	Gly	Val 525	Phe	His	Gly
30	Asp	Gln 530	Glu	Ser	Arg	Arg	Gly 535	Ser	Ile	Leu	Leu	Gly 540	Arg	Gly	Ala	Gly
	Gln 545	Thr	Gly	Pro	Leu	Pro 550	Arg	Ser	Pro	Leu	Pro 555	Gln	Ser	Pro	Asn	Pro 560
35	Gly	Arg	Arg	His	Gly 565	Glu	Glu	Gly	Gln	Leu 570	Gly	Val	Pro	Thr	Gly 575	Glu
				580	Ala				585					590		
40			595		Ser			600					605			
45		610			Val		615					620				
	625				Leu	630					635					640
50					Trp 645					650					655	
				660	Leu				665					670		
55			675		Val			680					685			
60		690			Phe		695					700				
	Thr 705	Val	Phe	Phe	Thr	Met 710	Glu	Met	Ala	Phe	Lys 715	Ile	Ile	Ala	Phe	Asp 720

		Pro	Tyr	Tyr	Tyr	Phe 725	Gln	Lys	Lys	Trp	Asn 730	Ile	Phe	Asp	Cys	Val 735	Ile
	5 .	Val	Thr	Val	Ser 740	Leu	Leu	Glu	Leu	Ser 745	Ala	Ser	Lys	Lys	Gly 750	Ser	Leu
		Ser	Val	Leu 755	Arg	Thr	Leu	_	Leu 760	Leu	Arg	Val	Phe	Lys 765	Leu	Ala	Lys
	10	Ser	Trp 770	Pro	Thr	Leu	Asn	Thr 775	Leu	Ile	Lys	Ile	Ile 780	Gly	Asn	Ser	Val
	15	Gly 785	Ala	Leu	Gly	Asn	Leu 790	Thr	Phe	Ile	Leu	Ala 795	Ile	Ile	Val	Phe	Ile 800
		Phe	Ala	Leu	Val	Gly 805	Lys	Gln	Leu	Leu	Ser 810	Glu	Asp	Tyr	Gly	Cys 815	Arg
•	20	Lys	Asp	Gly	Val 820	Ser	Val	Trp	Asn	Gly 825	Glu	Lys	Leu	Arg	Trp 830	His	Met
		Cys	Asp	Phe 835	Phe	His	Ser	Phe	Leu 840	Val	Val	Phe	Arg	Ile 845	Leu	Cys	Gly
	25	Glu	Trp 850	Ile	Glu	Asn	Met	Trp 855	Val	Cys	Met	Glu	Val 860	Ser	Gln	Lys	Ser
	30	Ile 865	Cys	Leu	Ile	Leu	Phe 870	Leu	Thr	Val	Met	Val 875	Leu	Gly	Asn	Leu	Val 880
		Val	Leu	Asn	Leu	Phe 885	Ile	Ala	Leu	Leu	Leu 890	Asn	Ser	Phe	Ser	Ala 895	Asp
	35	Asn	Leu	Thr	Ala 900	Pro	Glu	Asp	Asp	Gly 905	Glu	Val	Asn	Asn	Leu 910	Gln	Leu
	40	Ala	Leu	Ala 915	Arg	Ile	Gln	Val	Leu 920	Gly	His	Arg	Ala	Ser 925	Arg	Ala	Ser
		Ala	Ser 930	Tyr	Ile	Ser	Ser	His 935	Cys	Arg	Phe	His	Trp 940	Pro	Lys	Val	Glu
	45	Thr 945	Gln	Leu	Gly	Met	Lys 950	Pro	Pro	Leu	Thr	Ser 955	Ser	Glu	Ala	Lys	Asn 960
		His	Ile	Ala	Thr	Asp 965	Ala	Val	Ser	Ala	Ala 970	Val	Gly	Asn	Leu	Thr 975	Lys
	50	Pro	Ala	Leu	Ser 980	Ser	Pro	Lys		Asn 985	His	Gly	Asp	Phe	Ile 990	Thr	Asp
	55	Pro	Asn	Val 995	Trp	Val	Ser	Val	Pro 1000		Ala	Glu	Gly	Glu 1005		Asp	Leu
		Asp	Glu 1010		Glu	Glu	Asp	Met 1019		Gln	Ala	Ser	Gln 1020		Ser	Trp	Gln
	60	Glu 1029		Asp	Pro	Lys	Gly 1030		Gln	Glu	Gln	Leu 1035		Gln	Val	Gln	Lys 1040
	-	Cys	Glu	Asn	His	Gln 1045	Ala	Ala	Arg	Ser	Pro 1050		Ser	Met	Met	Ser 1055	

	Glu	Asp	Leu	Ala 1060		Tyr	Leu	Gly	Glu 1065		Trp	Lys	Arg	Lys 1070	Asp	Ser
5	Pro	Gln	Val 1075	Pro	Ala	Glu	Gly	Val 1080		Asp	Thr	Ser	Ser 1085		Glu	Gly
	Ser	Thr 1090		Asp	Cys	Pro	Asp 1095		Glu	Glu	Ile	Leu 1100	Arg	Lys	Ile	Pro
LO	Glu 1105		Ala	His	Asp	Leu 1110		Glu	Pro	Asp	Asp 1115	Cys	Phe	Arg	Glu	Gly 1120
15	Cys	Thr	Arg	Arg	Cys 1125		Cys	Cys	Asn	Val 1130	Asn)	Thr	Ser	Lys	Ser 1135	Pro
13	Trp	Ala	Thr	Gly 1140		Gln	Val	Arg	Lys 1145		Cys	Tyr	Arg	Ile 1150	Val	Glu
	His	Ser		Phe	Glu	Ser	Phe			Phe	Met	Ile	Leu 1165	Leu	Ser	Ser
20	Gly	Ala 1170		Ala	Phe	Glu	Asp 1175			Leu	Glu	Glu 1180	Lys		Arg	Val
25	Lys 1185		Val	Leu	Glu	Tyr 1190		Asp	Arg	Val	Phe 1195		Phe	Ile	Phe	Val 1200
	Phe	Glu	Met	Leu	Leu 1205		Trp	Val	Ala	Tyr 1210		Phe	Lys	Lys	Tyr 1215	Phe
30	Thr	Asn	Ala	Trp 1220		Trp	Leu	Asp	Phe 1225		Ile	Val	Asn	Ile 1230	Ser	Leu
. -	Thr	Ser	Leu 1235	Ile 5	Ala	Lys	Ile	Leu 1240		Tyr	Ser	Asp	Val 1245	Ala 5	Ser	Ile
35	Lys	Ala 125		Arg	Thr	Leu	Arg 125		Leu	Arg	Pro	Leu 1260		Ala	Leu	Ser
40	Arg 1269		Glu	Gly	Met	Arg 127		Val	Val	Asp	Ala 127		Val	Gly	Ala	Ile 128
	Pro	Ser	Ile	Met	Asn 128		Leu	Leu	Val	Cys 129		Ile	Phe	Trp	Leu 1299	Ile
45	Phe	Ser	Ile	Met 1300		Val	Asn	Leu	Phe 1305	Ala 5	Gly	Lys	Phe	Ser 1310	Lys)	Cys
50	Val	Asp	Thr 131	Arg 5	Asn	Asn	Pro	Phe 132	Ser O	Asn	Val	Asn	Ser 132	Thr 5	Met	Val
30	Asn	Asn 133		Ser	Glu	Cys	His 133		Gln	Asn	Ser	Thr 134	Gly 0	His	Phe	Phe
55	Trp 134		Asn	Val	Lys	Val 135	Asn 0	Phe	Asp	Asn	Val 135		Met	Gly	Tyr	Leu 136
	Ala	Leu	Leu	Gln	Val 136		Thr	Phe	Lys	Gly 137		Met	Asp	Ile	Met 137	
60	Ala	Ala	Val	Asp 138		Gly	Glu	Ile	Asn 138		Gln	Pro	Asn	Trp 139	Glu O	Asn
	Asn	Leu	Tyr 139	Met 5	Туr	Leu	Tyr	Phe 140		Val	Phe	Ile	Ile 140	Phe 5	Gly	Gly

	Phe	Phe 141	Thr	. Leu	a Asn	Leu	Phe 141	Val	Gly	Val	Ile	11e) Asr	ı Phe	. Asn
5	Gln 142	Gln 5	Lys	Lys	: Lys	Leu 143	Gly 0	Gly	Gln	Asp	Ile 143	Phe	Met	Thr	Glu	Glu 1440
10	Gln	Lys	Lys	Туг	Tyr 144	Asn 5	Ala	Met	Lys	Lys 145	Leu 0	Gly	Ser	Lys	Lys 145	
,	Gln	Lys	Pro	Ile 146	Pro 0	Arg	Pro	Leu	Asn 146	Lys 5	Tyr	Gln	Gly	Phe 147		Phe
15	Asp	Ile	Val 147	Thr 5	Arg	Gln	Ala	Phe 148	Asp 0	Ile	Ile	Ile	Met 148		Leu	Ile
	Суѕ	Leu 149	Asn 0	Met	Ile	Thr	Met 149	Met 5	Val	Glu	Thr	Asp 150		Gln	Gly	Glu
20	Glu 150	Lys 5	Thr	Lys	Val	Leu 1510	Gly)	Arg	Ile	Asn	Gln 151	Phe 5	Phe	Val	Ala	Val 1520
25	Phe	Thr	Gly	Glu	Cys 1525	Val	Met	Lys	Met	Phe 1530	Ala O	Leu	Arg	Gln	Tyr 153	
	Phe	Thr	Asn	Gly 154	Trp 0	Asn	Val	Phe	Asp 154	Phe 5	Ile	Val	Val	Ile 1550		Ser
30	Ile	Gly	Ser 155	Leu 5	Leu	Phe	Ser	Ala 1560	Ile)	Leu	Lys	Ser	Leu 1569		Asn	Tyr
	Phe	Ser 1570	Pro	Thr	Leu	Phe	Arg 1575	Val	Ile	Arg	Leu	Ala 1580		Ile	Gly	Arg
35	Ile 1589	Leu	Arg	Leu	Ile	Arg 1590	Ala	Ala	Lys	Gly	Ile 1599	Arg	Thr	Leu	Leu	Phe 1600
40					Ser 1605					1610)				1615	5
				1620					1625	5				1630	•	
45			1635	•	Glu			1640	1				1645			
		1650)		Met		1655					1660				_
50	1002	1				1670					1675		•			1680
55					Pro . 1685					1690					1695	
	Pro	Ala	Val	Gly 1700	Ile	Ile	Phe	Phe	Thr 1705	Thr	Tyr	Ile		Ilē 1710		Phe
60			1715	ļ	Asn i			1720					1725			
	Val	Ala 1730	Thr	Glu	Glu :	Ser '	Thr 1735	Glu	Pro	Leu	Ser	Glu 1740	Asp	Asp	Phe	Asp

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	Met Ph 1745	e Tyr	Glu	Thr	Trp 1750		Lys	Phe	Asp	Pro 1755	Glu 5	Ala	Thr	Gln	Phe 1760
5	Ile Al	a Phe	Ser	Ala 1765		Ser	Asp	Phe	Ala 1770	Asp)	Thr	Leu	Ser	Gly 1775	Pro
10	Leu Ar	g Ile	Pro 1780		Pro	Asn	Gln	Asn 1785		Leu	Ile	Gln	Met 1790		Leu
10	Pro Le	u Val 179		Gly	Asp	Lys	Ile 1800		Cys	Leu	Asp	Ile 1805		Phe	Ala
15	Phe Th	r Lys 10	Asn	Val	Leu	Gly 1815		Ser	Gly	Glu	Leu 1820		Ser	Leu	Lys
	Thr As	n Met	Glu	Glu	Lys 1830		Met	Ala	Thr	Asn 1835		Ser	Lys	Ala	Ser 1840
20	Tyr Gl	u Pro	Ile	Ala 1849		Thr	Leu	Arg	Trp 1850		Gln	Glu	Asp	Leu 1855	
25	Ala Th	r Val	Ile 1860		Lys	Ala	Tyr	Arg 1865		Tyr	Met	Leu	His 1870		Ser
	Leu Th	r Leu 187		Asn	Thr	Leu	His 1880		Pro	Arg	Ala	Glu 1889		Asp	Gly
30	Val Se	r Leu 90	Pro	Gly	Glu	Gly 189		Ile	Thr	Phe	Met 1900		Asn	Ser	Gly
	Leu Pr 1905	o Asp	Lys	Ser	Glu 1910		Ala	Ser	Ala	Thr 1915		Phe	Pro	Pro	Ser 1920
35	Tyr As			1929	5				1930)				1935	5
40	Ser Se	er Ser	Met 1940		Asn	Glu	Asp	Glu 1949		Ala	Ala	Lys	Glu 1950		Asn
	Ser Pr	o Gly 195		Gln											
45	(2) IN	IFORMA	TION	FOR	SEQ	ID I	NO : 3	:							
50	((QUENCA) L' B) T' C) S' D) To	ENGTI YPE : TRANI	H: 25 nucl	573 leic ESS:	base acio sino	pai: d	rs					,	
5 5	į)	li) MO	LECU:	LE T	YPE:	CDN	A					•			
60	(:	ix) FE ((ATUR A) N. B) L	AME/	KEY: ION:	CDS 561	21	26							
	()	ki) SE	QUEN	CE D	ESCR	IPTI	ON:	SEQ	ID N	O:3:					
	CTGGG	AGAGA	AAGC	GTCT	CG C	CTAG	CGAC'	r cc	CAGA	GCTT	TAA	GCCG	GGA 2	AGGG	ACAAGC

	GTCAGGACAT CTCAGAATCC CGAACCTTCT AGGGAGGGAG GTTCTTACCT CCATGCTTCC	120
_	CGTAGGAACC TAATCCCAAT TATTTAGCTG TATTTATAAT ACAAAATATG AATGTTAAAT	180
5	GTACAAAATG CTTTCCCAGC ATGCCTGCAT CTCCTCCTAG AGTCCTGTTC CCAAGCCCTC	240
	TCTACTCTCA GTACTGTAGA AAAGAAATAA GCTTTACGTG AGAAACCCAG GCACTGGATC	300
10	TTATCCAGGT GCTCACCTCA GAGTCTTTAG TGGGTGTAGC GCTGTGGTAG AGCATTTGGT	360
	TATAGATACA AACCCAGGGC AGGGAGACTG CAGTGGCCAT TCTCTCCCAG GCCAGACGTG	420
1.5	CCCTGATCCT TCCCACAGAG ATGAGAAGGC TGGAACCAGA ACACTCAGGT TTTGGCTTCT	480
15	CTTGGGGGAG GAGAGGTAAT CTTGTTACTT TAATAACATC AGTGTGTCCC TCTCCTCTAC	540
20	TAGGAGGCCA GGACATCTTC ATG ACA GAA GAG CAG AAG AAG TAC TAC AAT Met Thr Glu Glu Gln Lys Lys Tyr Tyr Asn 1 5 10	590
25	GCC ATG AAG AAG CTG GGC TCC AAG AAA CCC CAG AAG CCC ATC CCA CGG Ala Met Lys Lys Leu Gly Ser Lys Lys Pro Gln Lys Pro Ile Pro Arg 15 20 25	638
	CCC CTG AAT AAG TAC CAA GGC TTC GTG TTT GAC ATC GTG ACC AGG CAA Pro Leu Asn Lys Tyr Gln Gly Phe Val Phe Asp Ile Val Thr Arg Gln 30 35 40	686
30	GCC TTT GAC ATC ATC ATG GTT CTC ATC TGC CTC AAC ATG ATC ACC Ala Phe Asp Ile Ile Met Val Leu Ile Cys Leu Asn Met Ile Thr 45 50 55	73 4
35	ATG ATG GTG GAG ACC GAC GAG CAG GGC GAG GAG AAG ACG AAG GTT CTG Met Met Val Glu Thr Asp Glu Gln Gly Glu Glu Lys Thr Lys Val Leu 60 65 70	782
40	GGC AGA ATC AAC CAG TTC TTT GTG GCC GTC TTC ACG GGC GAG TGT GTG Gly Arg Ile Asn Gln Phe Phe Val Ala Val Phe Thr Gly Glu Cys Val 75 80 85 90	830
45	ATG AAG ATG TTC GCC CTG CGA CAG TAC TAC TTC ACC AAC GGC TGG AAC Met Lys Met Phe Ala Leu Arg Gln Tyr Tyr Phe Thr Asn Gly Trp Asn 95 100 105	878
43	GTG TTC GAC TTC ATA GTG GTG ATC CTG TCC ATT GGG AGT CTG CTG TTT Val Phe Asp Phe Ile Val Val Ile Leu Ser Ile Gly Ser Leu Leu Phe 110 115 120	926
50	TCT GCA ATC CTT AAG TCA CTG GAA AAC TAC TTC TCC CCG ACG CTC TTC Ser Ala Ile Leu Lys Ser Leu Glu Asn Tyr Phe Ser Pro Thr Leu Phe 125 130 135	974
55	CGG GTC ATC CGT CTG GCC AGG ATC GGC CGC ATC CTC AGG CTG ATC CGA Arg Val Ile Arg Leu Ala Arg Ile Gly Arg Ile Leu Arg Leu Ile Arg 140 145 150	1022
60	GCA GCC AAG GGG ATT CGC ACG CTG CTC TTC GCC CTC ATG ATG TCC CTG Ala Ala Lys Gly Ile Arg Thr Leu Leu Ph. Ala Leu Met Met Ser Leu 155 160 165 170	1070
	CCC GCC CTC TTC AAC ATC GGC CTC CTC CTC TTC CTC GTC ATG TTC ATC Pro Ala Leu Phe Asn Ile Gly Leu Leu Phe Leu Val Met Phe Ile 175 180 185	1118

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e	TCC Ser								1166
5	ATC Ile								1214
10	CTG Leu 220								1262
15	ATC Ile								1310
20	AAC Asn								1358
25	TTC Phe								1406
	ATC Ile								1454
30	GAG Glu 300								1502
35	AAG Lys								1550
40	GAC Asp								1598
45	CAG Gln								1646
	ATC Ile							 	1694
50	GAA Glu 380								1742
55	ATG Met								1790
60	CTC Leu								1838
	TAC Tyr								1886

5	CTG CAT GTG CCC AGG GCT GAG GAG GAT GGC GTG TCA CTT CCC GGG GAA Leu His Val Pro Arg Ala Glu Glu Asp Gly Val Ser Leu Pro Gly Glu 445 450 455	L934
•	GGC TAC ATT ACA TTC ATG GCA AAC AGT GGA CTC CCG GAC AAA TCA GAA Gly Tyr Ile Thr Phe Met Ala Asn Ser Gly Leu Pro Asp Lys Ser Glu 460 470	L982
10	ACT GCC TCT GCT ACG TCT TTC CCG CCA TCC TAT GAC AGT GTC ACC AGG Thr Ala Ser Ala Thr Ser Phe Pro Pro Ser Tyr Asp Ser Val Thr Arg 475 480 485 490	2030
15	GGC CTG AGT GAC CGG GCC AAC ATT AAC CCA TCT AGC TCA ATG CAA AAT Gly Leu Ser Asp Arg Ala Asn Ile Asn Pro Ser Ser Met Gln Asn 495 500 505	2078
20	GAA GAT GAG GTC GCT AAG GAA GGA AAC AGC CCT GGA CCT CAG TGAAGGCACT	?
20	Glu Asp Glu Val Ala Ala Lys Glu Gly Asn Ser Pro Gly Pro Gln 510 515 520	
	CAGGCATGCA CAGGGCAGGT TCCAATGTCT TTCTCTGCTG TACTAACTCC TTCCCTCTGG	2193
25	AGGTGGCACC AACCTCCAGC CTCCACCAAT GCATGTCACT GGTCATGGTG TCAGAACTGA	2253
	ATGGGGACAT CCTTGAGAAA GCCCCCACCC CAATAGGAAT CAAAAGCCAA GGATACTCCT	2313
30	CCATTCTGAC GTCCCTTCCG AGTTCCCAGA AGATGTCATT GCTCCCTTCT GTTTGTGACC	2373
	AGAGACGTGA TTCACCAACT TCTCGGAGCC AGAGACACAT AGCAAAGACT TTTCTGCTGG	2433
	TGTCGGGCAG TCTTAGAGAA GTCACGTAGG GGTTGGTACT GAGAATTAGG GTTTGCATGA	2493
35	CTGCATGCTC ACAGCTGCCG GACAATACCT GTGAGTCGGC CATTAAAATT AATATTTTTA	2553
	AAGTTAAAAA AAAAAAAAA	2573
40	(2) INFORMATION FOR SEQ ID NO:4:	
45	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 521 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: protein	
50	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:	
	Met Thr Glu Glu Gln Lys Lys Tyr Tyr Asn Ala Met Lys Lys Leu Gly 1 5 10 15	
55	Ser Lys Lys Pro Gln Lys Pro Ile Pro Arg Pro Leu Asn Lys Tyr Gln 20 25 30	
60	Gly Phe Val Phe Asp Ile Val Thr Arg Gln Ala Phe Asp Ile Ile Ile 35 40 45	
- -	Met Val Leu Ile Cys Leu Asn Met Ile Thr Met Met Val Glu Thr Asp 50 55 60	

	Glu 65	Gln	Gly	Glu	Glu	Lys 70	Thr	Lys	Val	Leu	Gly 75	Arg	Ile	Asn	Gln	Phe 80
5	Phe	Val	Ala	Val	Phe 85	Thr	Gly	Glu	Cys	Val 90	Met	Lys	Met	Phe	Ala 95	Leu
	Arg	Gln	Tyr	Tyr 100	Phe	Thr	Asn	Gly	Trp 105	Asn	Val	Phe	Asp	Phe 110	Ile	Val
10	Val	Ile	Leu 115	Ser	Ile	Gly	Ser	Leu 120	Leu	Phe	Ser	Ala	Ile 125	Leu	Lys	Ser
15	Leu	Glu 130	Asn	Tyr	Phe	Ser	Pro 135	Thr	Leu	Phe	Arg	Val 140	Ile	Arg	Leu	Ala
15	Arg 145	Ile	Gly	Arg	Ile	Leu 150	Arg	Leu	Ile	Arg	Ala 155	Ala	Lys	Gly	Ile	Arg 160
20	Thr	Leu	Leu	Phe	Ala 165	Leu	Met	Met	Ser	Leu 170	Pro	Ala	Leu	Phe	Asn 175	Ile
	Gly	Leu	Leu	Leu 180	Phe	Leu	Val	Met	Phe 185	Ile	Tyr	Ser	Ile	Phe 190	Gly	Met
25	Ala	Ser	Phe 195	Ala	Asn	Val	Val	Asp 200	Glu	Ala	Gly	Ile	Asp 205	Asp	Met	Phe
30	Asn	Phe 210	Lys	Thr	Phe	Gly	Asn 215	Ser	Met	Leu	Cys	Leu 220	Phe	Gln	Ile	Thr
	Thr 225	Ser	Ala	Gly	Trp	Asp 230	Gly	Leu	Leu	Ser	Pro 235	Ile	Leu	Asn	Thr	Gly 240
35	Pro	Pro	Tyr	Cys	Asp 245	Pro	Asn	Leu	Pro	Asn 250	Ser	Asn	Gly	Ser	Arg 255	Gly
	Asn	Cys	Gly	Ser 260	Pro	Ala	Val	Gly	Ile 265	Ile	Phe	Phe	Thr	Thr 270	Tyr	Ile
40	Ile	Ile	Ser 275	Phe	Leu	Ile	Val	Val 280	Asn	Met	Tyr	Ile	Ala 285	Val	Ile	Leu
45	Glu	Asn 290	Phe	Asn	Val	Ala	Thr 295	Glu	Glu	Ser	Thr	Glu 300	Pro	Leu	Ser	Glu
	Asp 305	Asp	Phe	Asp	Met	Phe 310	Tyr	Glu	Thr	Trp	Glu 315	Lys	Phe	Asp	Pro	Glu 320
50	Ala	Thr	Gln	Phe	Ile 325	Ala	Phe	Ser	Ala	Leu 330	Ser	Asp	Phe	Ala	Asp 335	Thr
	Leu	Ser	Gly	Pro 340	Leu	Arg	Ile	Pro	Lys 345	Pro	Asn	Gln	Asn	Ile 350	Leu	Ile
55	Gln	Met	Asp 355	Leu	Pro	Leu	Val	Pro 360	Gly	Asp	Lys	Ile	His 365	Cys	Leu	Asp
60	Ile	Leu 370	Phe	Ala	Phe	Thr	Lys 375	Asn	Val	Leu	Gly	Glu 380	Ser	Gly	Glu	Leu
	Asp 385	Ser	Leu	Lys	Thr	Asn 390	Met	Glu	Glu	Lys	Phe 395	Met	Ala	Thr	Asn	Leu 400
	Ser	Lys	Ala	Ser	Tyr	Glu	Pro	Ile	Ala	Thr	Thr	Leu	Arg	Trp	Lys	Gln

					405					410					415		
	Glu	Ąsp	Leu	Ser 420	Ala	Thr	Val	Ile	Gln 425	Lys	Ala	Tyr	Arg	Ser 430	Tyr	Met	
5	Leu	His	Arg 435	Ser	Leu	Thr	Leu	Ser 440	Asn	Thr	Leu	His	Val 445	Pro	Arg	Ala	
10	Glu	Glu 450	Asp	Gly	Val	Ser	Leu 455	Pro	Gly	Glu	Gly	Tyr 460	Ile	Thr	Phe	Met	
	Ala 465	Asn	Ser	Gly	Leu	Pro 470	Asp	Lys	Ser	Glu	Thr 475	Ala	Ser	Ala	Thr	Ser 480	
15	Phe	Pro	Pro	Ser	Tyr 485	Asp	Ser	Val	Thr	Arg 490	Gly	Leu	Ser	Asp	Arg 495	Ala	
	Asn	Ile	Asn	Pro 500	Ser	Ser	Ser	Met	Gln 505	Asn	Glu	Asp	Glu	Val 510	Ala	Ala	
20	Lys	Glu	Gly 515	Asn	Ser	Pro	Gly	Pro 520	Gln		,		٠				
25	(2)	INFO	ORMAT	гіои	FOR	SEQ	ID 1	10:5	:								
30		(i)	() ()	A) LI B) Ti C) Si	ENGTI YPE : TRANI	i: 70 nuci DEDNI	CTER 052 h leic ESS: line	acio sino	pai:	rs							
		(ii)) MOI	LECUI	LE T	PE:	cDN2	Ą									
35		(ix)	-	A) N2	AME/I		CDS 204	660	02								
40		(xi) SE	QUEN	CE DI	ESCR:	IPTIC	ON:	SEQ :	ID NO	0:5:						
	TAG	CTTG	CTT (CTGC	TAAT	GC T	ACCC	CAGG	CT	rtag:	ACAG	AGA	ACAG	ATG (GCAG	atggag	60
45	TTT	CTTA'	TTG (CCAT	GCGC	AA A	CGCT	GAGC	C CA	CCTC	ATGA	TCC	CGGA	ccc (CATG	GTTTTC	120
	AGT.	AGAC	AAC (CTGG	GCTA	AG A	AGAG	ATCT	C CG	ACCT'	TATA	GAG	CAGC	AAA (GAGT	GTAAAT	180
50	TCT	TCCC	CAA (GAAG.	AATG	AG A	AG A'					TT GO he A					230
55							TTC Phe					Leu					278
6.0							CGG Arg										326
60					Asp		GGC Gly			Pro							374

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	AAA Lys	GAC Asp	TGT Cys 60	AAC Asn	CAG Gln	CTG Leu	CCC Pro	AAG Lys 65	TTC Phe	TAT Tyr	GGT Gly	GAG Glu	CTC Leu 70	CCA Pro	GCA Ala	GAA Glu	422
5	CTG Leu	GTC Val 75	GGG Gly	GAG Glu	CCC Pro	CTG Leu	GAG Glu 80	GAC Asp	CTA Leu	GAC Asp	CCT	TTC Phe 85	TAC Tyr	AGC Ser	ACA Thr	CAC His	470
10											ACC Thr 100						518
15	GCC Ala	ACT Thr	TGG Trp	GCC Ala	CTG Leu 110	TGG Trp	CTC Leu	TTC Phe	AGT Ser	CCC Pro 115	TTC Phe	AAC Asn	CTG Leu	ATC Ile	AGA Arg 120	AGA Arg	566
20											TTC Phe						614
											ACC Thr						662
25											TAC Tyr						710
30											AAT Asn 180						758
35											GTC Val						806
40	GTG Val	GGT Gly	GCA Ala	GCG Ala 205	ATA Ile	GAC Asp	CTC Leu	CGA Arg	GGA Gly 210	ATC Ile	TCA Ser	GGC Gly	CTG Leu	CGG Arg 215	ACA Thr	TTC Phe	854
											GTG Val						902
45											AGG Arg						950
50	ACT Thr 250	ATC Ile	CTC Leu	ACA Thr	GTC Val	TTC Phe 255	TGC Cys	CTG Leu	AGC Ser	GTC Val	TTC Phe 260	GCC Ala	TTG Leu	GTG Val	GGC Gly	CTG Leu 265	998
55	CAG Gln	CTC Leu	TTT Phe	AAG Lys -	GGG Gly 270	AAC Asn	CTT Leu	AAG Lys	AAC Asn	AAA Lys 275	TGC Cys	ATC Ile	AGG Arg	AAC Asn	GGA Gly 280	ACA Thr	1046
60											GAA Glu						1094
											CTG Leu						1142

	GAT Asp	GCT Ala 315	GGT Gly	CAC His	TGC Cys	CCT Pro	GGA Gly 320	GGC Gly	TAT Tyr	GTC Val	TGC Cys	CTG Leu 325	AAA Lys	ACT Thr	CCT Pro	GAC Asp		1190
5										GAT Asp								1238
10										GAC Asp 355				_				1286
15										ATG Met								1334
20										CTG Leu								1382
										AGC Ser							:	1430
25										GAA Glu							•	1478
30										GGG Gly 435							:	1526
35										TCC Ser							:	1574
40										GAG Glu								1622
										CAG Gln							:	1670
45										AGC Ser							:	1718
50										TTT Phe 515								1766
55										GAA Glu								1814
60										GGT Gly								1862
-										AGA Arg								1910

	CTC Leu 570	GGA Gly	GTG Val	CCC Pro	ACT Thr	GGT Gly 575	GAG Glu	CTT Leu	ACC Thr	GCT Ala	GGA Gly 580	GCG Ala	CCT Pro	GAA Glu	GGC Gly	CCG Pro 585	:	1958
5	GCA Ala	CTC Leu	GAC Asp	ACT Thr	ACA Thr 590	GGG Gly	CAG Gln	AAG Lys	AGC Ser	TTC Phe 595	CTG Leu	TCT Ser	GCG Ala	GGC Gly	TAC Tyr 600	TTG Leu	;	2006
10	AAC Asn	GAA Glu	CCT Pro	TTC Phe 605	CGA Arg	GCA Ala	CAG Gln	AGG Arg	GCC Ala 610	ATG Met	AGC Ser	GTT Val	GTC Val	AGT Ser 615	ATC Ile	ATG Met	:	2054
15	ACT Thr	TCT Ser	GTC Val 620	ATT Ile	GAG Glu	GAG Glu	CTT Leu	GAA Glu 625	GAG Glu	TCT Ser	AAG Lys	CTG Leu	AAG Lys 630	TGC Cys	CCA Pro	CCC Pro	:	2102
	TGC Cys	TTG Leu 635	ATC Ile	AGC Ser	TTC Phe	GCT Ala	CAG Gln 640	AAG Lys	TAT Tyr	CTG Leu	ATC Ile	TGG Trp 645	GAG Glu	TGC Cys	TGC Cys	CCC Pro	:	2150
20	AAG Lys 650	TGG Trp	AGG Arg	AAG Lys	TTC Phe	AAG Lys 655	ATG Met	GCG Ala	CTG Leu	TTC Phe	GAG Glu 660	CTG Leu	GTG Val	ACT Thr	GAC Asp	CCC Pro 665	;	2198
25	TTC Phe	GCA Ala	GAG Glu	CTT Leu	ACC Thr 670	ATC Ile	ACC Thr	CTC Leu	TGC Cys	ATC Ile 675	GTG Val	GTG Val	AAC Asn	ACC Thr	GTC Val 680	TTC Phe	:	2246
30	ATG Met	GCC Ala	ATG Met	GAG Glu 685	CAC His	TAC Tyr	CCC Pro	ATG Met	ACC Thr 690	GAT Asp	GCC Ala	TTC Phe	GAT Asp	GCC Ala 695	ATG Met	CTT Leu	:	2294
35						GTC Val											:	2342
40						TTC Phe											;	2390
						GTC Val 735											;	2438
45	GCA Ala	TCC Ser	AAG Lys	AAG Lys	GGC Gly 750	AGC Ser	CTG Leu	TCT Ser	GTG Val	CTC Leu 755	CGT Arg	TCC Ser	TTA Leu	CGC Arg	TTG Leu 760	GCA Ala	;	2486
50	CTC Leu	GAC Asp	ACT Thr	ACA Thr 765	GGG Gly	CAG Gln	AAG Lys	AGC Ser	TTC Phe 770	CTG Leu	TCT Ser	GCG Ala	GGC Gly	TAC Tyr 775	TTG Leu	AAC Asn	:	2534
55	GAA Glu	CCT Pro	TTC Phe 780	CGA Arg	GCA Ala	CAG Gln	AGG Arg	GCC Ala 785	Met	AGC Ser	GTT Val	GTC Val	AGT Ser 790	ATC Ile	ATG Met	ACT Thr	:	2582
60	TCT Ser	GTC Val 795	ATT Ile	GAG Glu	GAG Glu	CTT Leu	GAA Glu 800	GAG Glu	TCT Ser	AAG Lys	CTG Leu	AAG Lys 805	TGC Cys	CCA Pro	CCC Pro	TGC Cys		2630
60		Ile				CAG Gln 815												2678

 	TGG Trp	AGG Arg	AAG Lys	TTC Phe	AAG Lys 830	ATG Met	GCG Ala	CTG Leu	TTC Phe	GAG Glu 835	CTG Leu	GTG Val	ACT Thr	GAC Asp	CCC Pro 840	TTC Phe		2726
5	GCA Ala	GAG Glu	CTT Leu	ACC Thr 845	ATC Ile	ACC Thr	CTC Leu	TGC Cys	ATC Ile 850	GTG Val	GTG Val	AAC Asn	ACC Thr	GTC Val 855	TTC Phe	ATG Met	2	2774
10	GCC Ala	ATG Met	GAG Glu 860	CAC His	TAC Tyr	CCC Pro	ATG Met	ACC Thr 865	GAT Asp	GCC Ala	TTC Phe	GAT Asp	GCC Ala 870	ATG Met	CTT Leu	CAA Gln		2822
15	GCC Ala	GGC Gly 875	AAC Asn	ATT Ile	GTC Val	TTC Phe	ACC Thr 880	GTG Val	TTT Phe	TTC Phe	ACA Thr	ATG Met 885	GAG Glu	ATG Met	GCC Ala	TTC Phe	2	870
20	AAG Lys 890	ATC Ile	ATT Ile	GCC Ala	TTC Phe	GAC Asp 895	CCC Pro	TAC Tyr	TAT Tyr	TAC Tyr	TTC Phe 900	CAG Gln	AAG Lys	AAG Lys	TGG Trp	AAT Asn 905	2	918
	ATC Ile	TTC Phe	GAC Asp	TGT Cys	GTC Val 910	ATC Ile	GTC Val	ACC Thr	GTG Val	AGC Ser 915	CTT Leu	CTG Leu	GAG Glu	CTG Leu	AGT Ser 920	GCA Ala	2	966
25	TCC Ser	AAG Lys	AAG Lys	GGC Gly 925	AGC Ser	CTG Leu	TCT Ser	GTG Val	CTC Leu 930	CGT Arg	TCC Ser	TTA Leu	CGC Arg	TTG Leu 935	CTG Leu	CGG Arg	: 3	014
30	GTC Val	TTC Phe	AAG Lys 940	CTG Leu	GCC Ala	AAG Lys	TCC Ser	TGG Trp 945	CCC Pro	ACC Thr	CTG Leu	AAC Asn	ACC Thr 950	CTC Leu	ATC Ile	AAG Lys	3	062
35	ATC Ile	ATC Ile 955	GGG Gly	AAC Asn	TCA Ser	GTG Val	GGG Gly 960	GCC Ala	CTG Leu	GGC Gly	AAC Asn	CTG Leu 965	ACC Thr	TTT Phe	ATC Ile	CTG Leu	3	110
					TTC Phe												3	158
					TGC Cys 990											Glu	3	206
					CAC His					Phe					Val		3	254
50	TTC Phe	CGA Arg	ATC Ile 1020	Leu	TGC Cys	GGG Gly	GAG Glu	TGG Trp 1025	Ile	GAG Glu	AAC Asn	ATG Met	TGG Trp 1030	Val	TGC Cys	ATG Met	3	302
			Ser		AAA Lys			Cys					Leu				3	350
		Leu			CTA Leu		Val					Ile					3	398
					GCG Ala 1070	Asp					Pro					Glu	3	446

	GTG Val	AAC Asn	AAC Asn	TTG Leu 1085	Gln	TTA Leu	GCA Ala	CTG Leu	GCC Ala 1090	Arg	ATC Ile	CAG Gln	GTA Val	CTT Leu 1099	Gly	CAT His	:	3494
5	CGG Arg	GCC Ala	AGC Ser 1100	AGG Arg	GCC	ATC Ile	GCC Ala	AGT Ser 1105	Tyr	ATC Ile	AGC Ser	AGC Ser	CAC His	Cys	CGA Arg	TTC Phe	:	3542
10			Pro					Gln				AAG Lys 1129	Pro				:	3590
15		Ser					His					GCT Ala)					;	3638
20						Lys					Ser	CCC Pro				His	:	3686
					Thr					Trp		TCT Ser			Ile		:	3734
25				Ser					Leu			GAT Asp		Glu			:	3782
30			Ser					Glu				GGA Gly 1205	Gln				;	3830
35		Pro					Cys					GCA Ala)					:	3878
40						Ser					Pro	TAC Tyr				Ser	:	3926
					Asp					Pro		GAG Glu			Asp		:	3974
45	ACG Thr	AGC Ser	TCC Ser 1260	Ser	GAG Glu	GGC Gly	AGC Ser	ACG Thr 1265	Val	GAC Asp	TGC Cys	CCG Pro	GAC Asp 1270	Pro	GAG Glu	GAA Glu	4	4022
50	ATC Ile	CTG Leu 1275	Arg	AAG Lys	ATC Ile	CCC Pro	GAG Glu 1280	Leu	GCA Ala	GAT Asp	GAC Asp	CTG Leu 1289	qzA	GAG Glu	CCC Pro	GAT Asp	4	4070
55		Cys					Cys					CCC Pro					4	4118
60	AAT Asn	ACT Thr	AGC Ser	AAG Lys	TCT Ser 1310	Pro	TGG Trp	GCC Ala	ACA Thr	GGC Gly 1315	Trp	CAG Gln	GTG Val	CGC Arg	AAG Lys 1320	Thr	•	4166
					Val					Phe		AGT Ser			Ile		•	4214

4982

							•	75-							
	ATG ATC Met Ile	CTG CT Leu Le 1340	C AGC	AGT Ser	GGA Gly	GCG Ala 1345	Leu	GCC Ala	TTT Phe	GAG Glu	GAT Asp 1350	Asn	TAC Tyr	CTG Leu	4262
5	GAA GAG Glu Glu 1359	Lys Pr	C CGA	GTG Val	AAG Lys 1360	Ser	GTG Val	CTG Leu	GAG Glu	TAC Tyr 1365	Thr	GAC Asp	CGA Arg	GTG Val	4310
10	TTC ACC Phe Thr 1370	TTC AT	C TTC e Phe	GTC Val 1375	Phe	GAG Glu	ATG Met	CTG Leu	CTC Leu 1380	Lys	TGG Trp	GTA Val	GCC Ala	TAT Tyr 1385	4358
15	GGC TTC Gly Phe	AAA AA Lys Ly	G TAT s Tyr 1390	Phe	ACC Thr	AAT Asn	GCC Ala	TGG Trp 1395	Суѕ	TGG Trp	CTG Leu	GAC Asp	TTC Phe 1400	Leu	4406
20	ATT GTG Ile Val	AAC AT Asn Il 14	e Ser	CTG Leu	ACA Thr	AGC Ser	CTC Leu 1410	Ile	GCG Ala	AAG Lys	ATC Ile	CTT Leu 1415	Glu	TAT Tyr	4454
20	TCC GAC Ser Asp	GTG GC Val Al 1420	G TCC a Ser	ATC Ile	AAA Lys	GCC Ala 1425	Leu	CGG Arg	ACT Thr	CTC Leu	CGT Arg 1430	Ala	CTC Leu	CGA Arg	4502
25	CCG CTG Pro Leu 143	Arg Al	T CTG a Leu	TCT Ser	CGA Arg 1440	Phe	GAA Glu	GGC Gly	ATG Met	AGG Arg 1449	Val	GTG Val	GTG Val	GAT Asp	4550
30	GCC CTC Ala Leu 1450	GTG GG Val Gl	C GCC y Ala	ATC Ile 1455	Pro	TCC Ser	ATC Ile	ATG Met	AAC Asn 1460	Val	CTC Leu	CTC Leu	GTC Val	TGC Cys 1465	4598
35	CTC ATC Leu Ile	TTC TG Phe Tr	G CTC p Leu 1470	Ile	TTC Phe	AGC Ser	ATC Ile	ATG Met 1475	Gly	GTG Val	AAC Asn	CTC Leu	TTC Phe 148	Ala	4646
4.0	GGG AAA Gly Lys	Phe Se	G AAG r Lys 85	TGC Cys	GTC Val	GAC Asp	ACC Thr 149	Arg	AAT Asn	AAC Asn	ČCA Pro	TTT Phe 149	Ser	AAC Asn	4694
40	GTG AAT Val Asn	TCG AC Ser Th	G ATG	GTG Val	AAT Asn	AAC Asn 1509	Lys	TCC Ser	GAG Glu	TGT Cys	CAC His 151	Asn	CAA Gln	AAC Asn	4742
45	AGC ACC Ser Thr 151	Gly Hi	C TTC s Phe	TTC Phe	TGG Trp 152	Val	AAC Asn	GTC Val	AAA Lys	GTC Val 152	Asn	TTC Phe	GAC Asp	AAC Asn	4790
50	GTC GCT Val Ala 1530	ATG GO Met Gl	C TAC y Tyr	CTC Leu 153	Ala	CTT Leu	CTT Leu	CAG Gln	GTG Val 154	Ala	ACC Thr	TTC Phe	AAA Lys	GGC Gly 1545	4838
55	TGG ATG Trp Met	GAC AT	A ATG e Met 155	Tyr	GCA Ala	GCT Ala	GTT Val	GAT Asp 155	Ser	GGA Gly	GAG Glu	ATC Ile	AAC Asn 156	Ser	4886
60	CAG CCT Gln Pro	Asn Ti	GG GAG p Glu 565	AAC Asn	AAC Asn	TTG Leu	TAC Tyr 157	Met	TAC Tyr	CTG Leu	TAC Tyr	TTC Phe 157	Val	GTT Val	4934

TTC ATC ATT TTC GGT GGC TTC TTC ACG CT; AAT CTC TTT GTT GGG GTC Phe Ile Phe Gly Gly Phe Phe Thr Leu Asn Leu Phe Val Gly Val 1580 1585 1590

60

1580

	ATA Ile	ATC Ile 1595	Asp	AAC Asn	TTC Phe	AAC Asn	CAA Gln 1600	Gln	AAA Lys	AAA Lys	AAG Lys	CTA Leu 1605	Gly	GGC Gly	CAG Gln	GAC Asp	5030
5	ATC Ile 1610	Phe	ATG Met	ACA Thr	GAA Glu	GAG Glu 1615	Gln	AAG Lys	AAG Lys	TAC Tyr	TAC Tyr 1620	Asn	GCC Ala	ATG Met	AAG Lys	AAG Lys 1625	5078
10	CTG Leu	GGC Gly	TCC Ser	AAG Lys	AAA Lys 1630	Pro	CAG Gln	AAG Lys	CCC Pro	ATC Ile 1635	Pro	CGG Arg	CCC Pro	CTG Leu	AAT Asn 1640	Lys	5126
15	TAC Tyr	CAA Gln	GGC Gly	TTC Phe 1645	Val	TTT Phe	GAC Asp	ATC Ile	GTG Val 1650	Thr	AGG Arg	CAA Gln	GCC Ala	TTT Phe 1655	Asp	ATC Ile	5174
20	ATC Ile	Ile	Met 1660	Val	Leu	Ile	Cys	Leu 1665	Asn	Met	Ile	Thr	Met 1670	Met)	Val	Glu	5222
		Asp 1675	Glu	Gln	Gly	Glu	Glu 1680	Lys)	Thr	Lys	Val	Leu 1685	Gly	Arg	Ile	Asn	5270
25	CAG Gln 1690	Phe	Phe	Val	Ala	Val 1695	Phe	Thr	Gly	Glu	Cys 1700	Val	Met	Lys	Met	Phe 1705	5318
30	GCC Ala	Leu	Arg	Gln	Tyr 1710	Tyr)	Phe	Thr	Asn	Gly 1715	Trp	Asn	Val	Phe	Asp 1720	Phe)	5366
35		Val	Val	Ile 1725	Leu	Ser	Ile	Gly	Ser 1730	Leu)	Leu	Phe	Ser	Ala 1739	Ile	Leu	5414
40		Ser	Leu 1740	Glu)	Asn	Tyr	Phe	Ser 1749	Pro	Thr	Leu	Phe	Arg 1750	Val	Ile	Arg	5462
		Ala 1759	Arg	Ile	Gly	Arg	Ile 1760	Leu)	Arg	Leu	Ile	Arg 1765	Ala	Ala	Lys	Gly	5510
45	Ile 1770	Arg	Thr	Leu	Leu	Phe 177	•	Leu	Met	Met	Ser 1780	Leu)	Pro	Ala	Leu	Phe 1785	5558
50	Asn	Ile	Gly	Leu	Leu 1790	Leu)	TTC Phe	Leu	Val	Met 1799	Phe 5	Ile	Tyr	Ser	Ile 1800	Phe)-	5606
55	Gly	Met	Ala	Ser 180	Phe 5	Ala	AAC Asn	Val	Val 181	Asp O	Glu	Ala	Gly	Ile 1819	Asp	Asp	5654
60	ATG Met	TTC Phe	AAC Asn 182	Phe	AAG Lys	ACC Thr	TTT Phe	GGC Gly 182	Asn	AGC Ser	ATG Met	CTG Leu	TGC Cys 1830	Leu	TTC Phe	CAG Gln	5702
	ATC Ile	ACC Thr 183	Thr	TCG Ser	GCC Ala	GGC Gly	TGG Trp 184	Ąsp	GGC Gly	CTC Leu	CTC Leu	AGC Ser 184	Pro	ATC Ile	CTC Leu	AAC Asn	5750

	ACG GGG Thr Gly 1850	CCT CC	C TAC	TGC GAC Cys Asp 1855	CCC Pro	AAC Asn	CTG Leu	CCC Pro 1	Asn S	AGC AAG Ser Ası	GGC Gly	TCC Ser 1865		5798
5	CGG GGG Arg Gly	AAC TO Asn Cy	GC GGG 's Gly 1870	AGC CCG Ser Pro	GCG Ala	GTG Val	GGC Gly 1879	Ile :	ATC T Ile P	TTC TTC	ACC Thr	Thr		5846
10	Tyr Ile	Ile Il 18	e Ser 85	TTC CTC Phe Leu	Ile	Val 1890	Val	Asn N	Met T	yr Ile 189	Ala 5	Val		5894
15	ATT CTG Ile Leu	GAG AA Glu As 1900	C TTC n Phe	AAC GTA Asn Val	GCC Ala 1905	Thr	GAG Glu	GAG A	Ser T	CG GAC hr Glu 910	CCC Pro	CTG Leu		5942
20	AGC GAG Ser Glu 1915	Asp As	C TTC p Phe	GAC ATG Asp Met 192	Phe	TAT Tyr	GAG Glu	Thr T	rgg g Frp G 1925	AG AAG lu Lys	TTC Phe	GAC Asp		5990
	CCG GAG Pro Glu 1930	GCC AC Ala Th	C CAG r Gln	TTC ATT Phe Ile 1935	GCC Ala	TTT Phe	TCT Ser	GCC C Ala L 1940	CTC T Leu S	CA GAC er Asp	TTC Phe	GCG Ala 1945		6038
25	GAC ACG Asp Thr	CTC TC Leù Se	C GGC r Gly 1950	Pro Leu	AGA Arg	Ile	CCC Pro 1955	Lys F	CCC A	AC CAG sn Gln	AAT Asn 196	Ile	ber , q. 6	6086
30	TTA ATC Leu Ile	CAG AT Gln Me 19	t Asp	CTG CCG Leu Pro	TTG Leu	GTC Val 1970	Pro	GGG G Gly A	SAT A	AG ATC ys Ile 197	His	TGT Cys		6134
35	CTG GAC Leu Asp	ATC CT Ile Le 1980	T TTT u Phe	GCC TTC Ala Phe	ACA Thr 1985	Lys .	AAC Asn	GTC T Val L	eu G	GA GAA ly Glu 990	TCC Ser	GGG Gly		6182
40	GAG TTG Glu Leu 1995	Asp Se	C CTG r Leu	AAG ACC Lys Thr 2000	Asn	ATG (GAA Glu	Glu L	AG T' ys Pl	TT ATG he Met	GCG Ala	ACC Thr		6230
••	AAT CTC Asn Leu 2010	TCC AA Ser Ly	s Ala	TCC TAT Ser Tyr 2015	GAA Glu	CCA . Pro	Ile	GCC A Ala T 2020	CC AC	CC CTC hr Leu	CGG Arg	TGG Trp 2025		6278
45	AAG CAG Lys Gln	GAA GA Glu As	C CTC p Leu 2030	Ser Ala	ACA Thr	Val :	ATT Ile 2035	CAA A Gln L	AG G ys A	CC TAC la Tyr	CGG Arg 2040	Ser		6326
50	TAC ATG Tyr Met	CTG CA Leu Hi 20	s Arg	TCC TTG Ser Leu	ACA Thr	CTC ' Leu : 2050	TCC . Ser .	AAC A Asn T	CC C	TG CAT eu His 205	Val	CCC Pro		6374
55	AGG GCT Arg Ala	GAG GAG Glu Gl 2060	G GAT u Asp	GGC GTG Gly Val	TCA Ser 2065	Leu :	CCC (Pro (GGG G Gly G	lu G	GC TAC ly Tyr 070	AGT Ser	ACA Thr		6422
60	TTC ATG Phe Met 2075	Ala As	C AGT	GGA CTC Gly Leu 2080	Pro	GAC A	AAA ' Lys :	Ser G	AA AC lu Ti 085	CT GCC nr Ala	TCT Ser	GCT Ala		6470
	ACG TCT Thr Ser 2090	TTC CC	o Pro	TCC TAT Ser Tyr 2095	GAC Asp	AGT (Ser '	Val '	ACC A Thr A 2100	GG GG rg G]	GC CTG ly Leu	AGT Ser	GAC Asp 2105		6518

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						Pro					Gln					GTC Val		6566
5					Gly					Pro			AGGC2	ACT (CAGG	CATGO	CA	6619
	CAG	GCAC	GT 1	CCA	ATGTO	T T	rctc:	rgcto	G TAC	CTAAC	CTCC	TTC	CTC	rgg .	AGGT	GGCAC	C:C	6679
10	AAC	TCC#	AGC (CTCCA	ACCAF	AT GO	CATG	rcaci	r gg:	CATO	GTG	TCAC	JAAC'	rga i	ATGG	GGACA	T	6739
	CCTT	rgag <i>i</i>	LAA C	ccc	CAC	cc ,cz	ATA	GAAT	r CA	AAAG	CAA	GGAT	CACTO	CT (CCAT'	rctga	'C	6799
15	GTC	CTTC	CCG A	GTTC	CCAC	A AC	GATGI	CAT	r GC	rccc	TCT	GTT	rgtg <i>i</i>	ACC I	AGAG	ACGTG	A	6859
	TTC	ACCA	CT T	CTC	GAGC	C AC	GAGAC	CACAT	r ago	CAAAC	SACT	TTTC	TGC	rgg '	TGTC	GGCA	.G	6919
	TCTT	ragac	SAA C	STCAC	GTAC	G G	STTGO	TACI	r GAG	GAATT	ragg	GTTT	GCAT	rga (CTGC	ATGCT	'C	6979
20	ACAC	CTGC	CG C	JACA.	TACC	T G	rgagi	rcgg	CA	TAA	TTA	AATA	TTTT	TA Z	AAGT'	AAAA	A	7039
	AAA	\AAA.	AAA A	LAA														7052
25																		
	(2)						ID N											
30		,	(1) :	(A)	LEN TYP	NGTH:	RACTE : 213 amino GY: 3	32 ar	nino id	acio	is							
		i)	li) N	OLEC	CULE	TYPE	E: pi	rotei	in									
35							=			Q ID	NO:6	5:						
35	Met 1	(>	ci) S	SEQUE	ENCE	DESC	CRIPT	rion:	: SE(-		5: Asn	Phe	Arg	Arg 15	Phe		
35	1	() Glu	ci) S Leu	SEQUE Pro	ENCE Phe 5	DESC Ala	CRIPT	rion: Val	: SE(Thr 10	Thr				15			
40	1 Thr	Glu Pro	ci) S Leu Glu	Pro Ser 20	Phe 5 Leu	DESC Ala Ala	Ser Glu	rion: Val Ile	Gly Glu 25	Thr 10 Lys	Thr Gln	Asn	Ala	Ala 30	15 His	Arg		
	1 Thr Ala	Glu Pro Ala	ci) S Leu Glu Lys 35	Pro Ser 20 Lys	Phe 5 Leu Ala	DESC Ala Ala Arg	Ser Glu Thr	Val Ile Lys 40	Gly Glu 25 His	Thr 10 Lys Arg	Thr Gln Gly	Asn Ile	Ala Glu 45	Ala 30 Asp	15 His Lys	Arg Gly		
40	1 Thr Ala Glu	Glu Pro Ala Lys 50	Leu Glu Lys 35	Pro Ser 20 Lys	Phe 5 Leu Ala	DESC Ala Ala Arg Gln	Ser Glu Thr Leu 55	Val Ile Lys 40 Asp	Gly Glu 25 His	Thr 10 Lys Arg	Thr Gln Gly Asp	Asn Ile Gln Cys	Ala Glu 45 Asn	Ala 30 Asp	15 His Lys Leu	Arg Gly Pro		
40	Thr Ala Glu Lys 65	Glu Pro Ala Lys 50 Phe	ci) S Leu Glu Lys 35 Pro	Pro Ser 20 Lys Arg	Phe 5 Leu Ala Pro	DESC Ala Ala Arg Gln Leu 70	Ser Glu Thr Leu 55	Val Ile Lys 40 Asp	Gly Glu 25 His Leu Glu	Thr 10 Lys Arg Lys	Thr Gln Gly Asp Val 75	Asn Ile Gln Cys 60	Ala Glu 45 Asn Glu	Ala 30 Asp Gln Pro	15 His Lys Leu Leu	Arg Gly Pro Glu 80		
40	Thr Ala Glu Lys 65 Asp	Glu Pro Ala Lys 50 Phe	Leu Glu Lys 35 Pro Tyr	SEQUE Pro Ser 20 Lys Arg Gly	Phe 5 Leu Ala Pro Glu	DESC Ala Ala Arg Gln Leu 70	Ser Glu Thr Leu 55 Pro	Val Ile Lys 40 Asp Ala	Glu 25 His Leu Glu	Thr 10 Lys Arg Lys Leu Arg 90	Thr Gln Gly Asp Val 75 Thr	Asn Ile Gln Cys 60 Gly	Ala Glu 45 Asn Glu Met	Ala 30 Asp Gln Pro	15 His Lys Leu Leu	Arg Gly Pro Glu 80 Asn		
40 45 50	Thr Ala Glu Lys 65 Asp	Glu Pro Ala Lys 50 Phe Leu Ser	Leu Clu Lys 35 Pro Tyr Asp	SEQUE Pro Ser 20 Lys Arg Gly Pro	Phe 5 Leu Ala Pro Glu Phe 85	DESC Ala Ala Arg Gln Leu 70 Tyr	Ser Glu Thr Leu 55 Pro Ser Arg	Val Ile Lys 40 Asp Ala Thr	Glu 25 His Leu Glu His	Thr 10 Lys Arg Lys Leu Arg 90	Thr Gln Gly Asp Val 75 Thr	Asn Ile Gln Cys 60 Gly Phe	Ala Glu 45 Asn Glu Met	Ala 30 Asp Gln Pro Val Leu 110	15 His Lys Leu Leu 95	Arg Gly Pro Glu 80 Asn		
40 45	Thr Ala Glu Lys 65 Asp Lys	Glu Pro Ala Lys 50 Phe Leu Ser	Leu Glu Lys 35 Pro Tyr Asp Arg	SEQUE Pro Ser 20 Lys Arg Gly Pro Thr 100 Phe	Phe 5 Leu Ala Pro Glu Phe 85 Ile Asn	DESC Ala Ala Arg Gln Leu 70 Tyr Ser Leu	Ser Glu Thr Leu 55 Pro Ser Arg	Val Ile Lys 40 Asp Ala Thr Phe Arg 120	Gly Glu 25 His Leu Glu His Ser 105 Arg	Thr 10 Lys Arg Lys Leu Arg 90 Ala	Thr Gln Gly Asp Val 75 Thr Thr	Asn Ile Gln Cys 60 Gly Phe	Ala Glu 45 Asn Glu Met Ala Lys 125	Ala 30 Asp Gln Pro Val Leu 110 Val	15 His Lys Leu Leu 95 Trp	Arg Gly Pro Glu 80 Asn Leu Val		

	145					150					155					160
5	Thr	Val	Ile	Tyr	Thr 165	Phe	Glu	Ala	Leu	Ile 170	Lys	Ile	Leu	Ala	Arg 175	Gly
J	Phe	Cys	Leu	Asn 180	Glu	Phe	Thr	Tyr	Leu 185	Arg	Asp	Pro	Trp	Asn 190	Trp	Leu
10	Asp	Phe	Ser 195	Val	Ile	Thr	Leu	Ala 200	Tyr	Val	Gly	Ala	Ala 205	Ile	Asp	Leu
	Arg	Gly 210	Ile	Ser	Gly	Leu	Arg 215	Thr	Phe	Arg	Val	Leu 220	Arg	Ala	Leu	Lys
15	Thr 225	Val	Ser	Val	Ile	Pro 230	Gly	Leu	Lys	Val	Ile 235	Val	Gly	Ala	Leu	Ile 240
20	His	Ser	Val	Arg	Lys 245	Leu	Ala	Asp	Val	Thr 250	Ile	Leu	Thr	Val	Phe 255	Cys
	Leu	Ser	Val	Phe 260	Ala	Leu	Val	Gly	Leu 265	Gln	Leu	Phe	Lys	Gly 270	Asn	Leu
25	Lys	Asn	Lys 275	Cys	Ile	Arg	Asn	Gly 280	Thr	Asp	Pro	His	Lys 285	Ala	Asp	Asn
	Leu	Ser 290	Ser	Glu	Met	Ala	Glu 295	Tyr	Ile	Phe	Ile	Lys 300	Pro	Gly	Thr	Thr
30	Asp 305	Pró	Leu	Leu	Cys	Gly 310	Asn	Gly	Ser	Asp	Ala 315	Gly	His	Cys	Pro	Gly 320
35	Gly	Tyr	Val	Cys	Leu 325	Lys	Thr	Pro	Asp	Asn 330	Pro	Asp	Phe	Asn	Tyr 335	Thr
	Ser	Phe	Asp	Ser 340	Phe	Ala	Trp	Ala	Phe 345	Leu	Ser	Leu	Phe	Arg 350	Leu	Met
40	Thr	Gln	Asp 355	Ser	Trp	Glu	Arg	Leu 360	Tyr	Gln	Gln	Thr	Leu 365	Arg	Ala	Ser
45	Gly	Lys 370	Met	Tyr	Met	Val	Phe 375	Phe	Val	Leu	Val	Ile 380	Phe	Leu	Gly	Ser
13	Phe 385	Tyr	Leu	Val	Asn	Leu 390	Ile	Leu	Ala	Val	Val 395	Thr	Met	Ala	Tyr	Glu 400
50	Glu	Gln	Ser	Gln	Ala 405	Thr	Ile	Ala	Glu	Ile 410	Glu	Ala	Lys	Glu	Lys 415	Lys
	Phe	Gln	Glu	Ala 420	Leu	Glu	Val	Leu	Gln 425	Lys	Glu	Gln	Glu	Val 430	Leu	Ala
55	Ala	Leu	Gly 435	Ile	Asp	Thr	Thr	Ser 440	Leu	Gln	Ser	His	Ser 445	Gly	Ser	Pro
60	Leu	Ala 450	Ser	Lys	Asn	Ala	Asn 455	Glu	Arg	Arg	Pro	Arg 460	Val	Lys	Ser	Arg
- ·	Val 465	Ser	Glu	Gly	Ser	Thr 470	Asp	Asp	Asn	Arg	Ser 475	Pro	Gln	Ser	Asp	Pro 480

	Tyr	Asn	Gln	Arg	Arg 485	Met	Ser	Phe	Leu	Gly 490	Leu	Ser	Ser	Gly	Arg 495	Arg
5	Arg	Ala	Ser	His 500	Gly	Ser	Val	Phe	His 505	Phe	Arg	Ala	Pro	Ser 510	Gln	Asp
	Ile	Ser	Phe 515	Pro	Asp	Gly	Ile	Thr 520	Pro	Asp	Asp	Gly	Val 525	Phe	His	Gly
10	Asp	Gln 530	Glu	Ser	Arg	Arg	Gly 535	Ser	Ile	Leu	Leu	Gly 540	Arg	Gly	Ala.	Gly
15	Gln 545	Thr	Gly	Pro	Leu	Pro 550	Arg	Ser	Pro	Leu	Pro 555	Gln	Ser	Pro	Asn	Pro 560
15	Gly	Arg	Arg	His	Gly 565	Glu	Glu	Gly	Gln	Leu 570	Gly	Val	Pro	Thr	Gly 575	Glu
20	Leu	Thr	Ala	Gly 580	Ala	Pro	Glu	Gly	Pro 585	Ala	Leu	Asp	Thr	Thr 590	Gly	Gln
	Lys	Ser	Phe 595	Leu	Ser	Ala	Gly	Tyr 600	Leu	Asn	Glu	Pro	Phe 605	Arg	Ala	Gln
25	Arg	Ala 610	Met	Ser	Val	Val	Ser 615	Ile	Met	Thr	Ser	Val 620	Ile	Glu	Glu	Leu
30	Glu 625	Glu	Ser	Lys	Leu	Lys 630	Cys	Pro	Pro	Cys	Leu 635	Ile	Ser	Phe	Ala	Gln 640
30	Lys	Tyr	Leu	Ile	Trp 645	Glu	Cys	Cys	Pro	Lys 650	Trp	Arg	Lys	Phe	Lys 655	Met
35	Ala	Leu	Phe	Glu 660	Leu	Val	Thr	Asp	Pro 665	Phe	Ala	Glu	Leu	Thr 670	Ile	Thr
	Leu	Cys	Ile 675	Val	Val	Asn	Thr	Val 680	Phe	Met	Ala	Met	Glu 685	His	Tyr	Pro
40	Met	Thr 690	Asp	Ala	Phe	Asp	Ala 695	Met	Leu	Gln	Ala	Gly 700	Asn	Ile	Val	Phe
45	Thr 705	Val	Phe	Phe	Thr	Met 710	Glu	Met	Ala	Phe	Lys 715	Ile	Ile	Ala	Phe	Asp 720
	Pro	Tyr	Tyr	Tyr	Phe 725	Gln	Lys	Lys	Trp	Asn 730	Ile	Phe	Asp	Cys	Val 735	Ile
50	Val	Thr	Val	Ser 740	Leu	Leu	Glu	Leu	Ser 745	Ala	Ser	Lys	Lys	Gly 750	Ser	Leu
	Ser	Val	Leu 755	Arg	Ser	Leu	Arg	Leu 760	Ala	Leu	Asp	Thr	Thr 765	Gly	Gln	Lys
55	Ser	Phe 770	Leu	Ser	Ala	Gly	Tyr 775	Leu	Asn	Glu	Pro	Phe 780	Arg	Ala	Gln	Arg
60	Ala 785	Met	Ser	Val	Val	Ser 790	He	Met	Thr	Ser	Val 795	Ile	Glu	Glu	Leu	Glu 800
	Glu	Ser	Lys	Leu	Lys 805	Cys	Pro	Pro	Cys	Leu 810	Ile	Ser	Phe	Ala	Gln 815	Lys

	Tyr	Leu	Ile	Trp 820	Glu	Cys	Cys	Pro	Lys 825	Trp	Arg	Lys	Phe	Lys 830	Met	Ala
5	Leu	Phe	Glu 835	Leu	Val	Thr	Asp	Pro 840	Phe	Ala	Glu	Leu	Thr 845	Ile	Thr	Leu
	Cys	Ile 850	Val	Val	Asn	Thr	Val 855	Phe	Met	Ala	Met	Glu 860	His	Tyr	Pro	Met
10	Thr 865	Asp	Ala	Phe	Asp	Ala 870	Met	Leu	Gln	Ala	Gly 875	Asn	Ile	Val	Phe	Thr 880
15	Val	Phe	Phe	Thr	Met 885	Glu	Met	Ala	Phe	Lys 890	Ile	Ile	Ala	Phe	Asp 895	Pro
	Tyr	Tyr	Tyr	Phe 900	Gln	Lys	Lys	Trp	Asn 905	Ile	Phe	Asp	Cys	Val 910	Ile	Val
20	Thr	Val	Ser 915	Leu	Leu	Glu	Leu	Ser 920	Ala	Ser	Lys	Lys	Gly 925	Ser	Leu	Ser
25	Val	Leu 930	Arg	Ser	Leu	Arg	Leu 935	Leu	Arg	Val	Phe	Lys 940	Leu	Ala	Lys	Ser
	Trp 945	Pro	Thr	Leu	Asn	Thr 950	Leu	Ile	Lys	Ile	Ile 955	Gly	Asn	Ser	Val	Gly 960
30	Ala	Leu	Gly	Asn	Leu 965	Thr	Phe	Ile	Leu	Ala 970	Ile	Ile	Val	Phe	Ile 975	Phe
	Ala	Leu	Val	Gly 980	Lys	Gln	Leu	Leu	Ser 985	Glu	Asp	Tyr	Gly	Суs 990	Arg	Lys
35	Asp	Gly	Val 995	Ser	Val	Trp	Asn	Gly 1000		Lys	Leu	Arg	Trp 1005		Met	Cys
40	Asp	Phe 1010		His	Ser	Phe	Leu 1019	Val	Val	Phe	Arg	Ile 1020		Cys	Gly	Glu
	Trp 1025		Glu	Asn	Met	Trp 1030		Cys	Met	Glu	Val 1035		Gln	Lys	Ser	Ile 1040
45	Cys	Leu	Ile	Leu	Phe 1045		Thr	Val	Met	Val 1050		Gly	Asn	Leu	Val 1055	
	Leu	Asn	Leu	Phe 1060		Ala	Leu	Leu	Leu 1065		Ser	Phe	Ser	Ala 1070	•	Asn
50	Leu	Thr	Ala 1075		Glu	Asp	Asp	Gly 1080		Val	Asn	Asn	Leu 1085		Leu	Ala
55	Leu	Ala 1090		Ile	Gln	Val	Leu 1099	Gly	His	Arg	Ala	Ser 1100		Ala	Ile	Ala
	Ser 1105		Ile	Ser	Ser	His 1110		Arg	Phe	Arg	Trp 1115		Lys	Val	Glu	Thr 1120
60	Gln	Leu	Gly	Met	Lys 1129		Pro	Leu	Thr	Ser 1130		Glu	Ala	Lys	Asn 1135	
	Ile	Ala	Thr	Asp 1140		Val	Ser	Ala	Ala 1145		Gly	Asn	Leu	Thr 1150	_	Pro

	Ala	Leu	Ser 1155	Ser	Pro	Lys	Glu	Asn 1160		Gly	Asp	Phe	Ile 1165	Thr	Asp	Pro
5	Asn	Val 1170		Val	Ser	Val	Pro 1175		Ala	Glu	Gly	Glu 1180	Ser	Asp	Leu	Asp
	Glu 1185		Glu	Glu	Asp	Met 1190	Glu	Gln	Ala	Ser	Gln 1195	Ser	Ser	Trp	Gln	Glu 1200
10	Glu	Asp	Pro	Lys	Gly 1205		Gln	Glu	Gln	Leu 1210		Gln	Val	Gln	Lys 1215	Cys
	Glu	Asn	His	Gln 1220		Ala	Arg	Ser	Pro 1225		Ser	Met	Met	Ser 1230	Ser	Glu
15	qzA	Leu	Ala 1235	Pro	Туг	Leu	Gly	Glu 1240	Ser		Lys	Arg	Lys 1245	Asp		Pro
2.0	Gln	Val 1250		Ala	Glu	Gly	Val 1255		Asp	Thr	Ser	Ser 1260		Glu	Gly	Ser
20	Thr 1265		Asp	Cys	Pro	Asp 1270		Glu	Glu	Ile	Leu 1275	Arg	Lys	Ile	Pro	Glu 1280
25	Leu	Ala	Asp	Asp	Leu 1285		Glu	Pro	Asp	Asp 1290		Phe	Thr	Glu	Gly 1295	Cys ;
	Thr	Arg	Arg	Cys 1300		Cys	Cys	Asn	Val 1309		Thr	Ser	Lys	Ser 131	Pro	Trp
30	Ala	Thr	Gly 1315	Trp	Gln	Val	Arg	Lys 1320		Cys	Tyr	Arg	Ile 1329	Val	Glu	His
25	Ser	Trp 1330		Glu	Ser	Phe	Ile 133		Phe	Met	Ile	Leu 1340		Ser	Ser	Gly
35	Ala 134		Ala	Phe	Glu	Asp 1350		Tyr	Leu	Glu	Glu 1359		Pro	Arg	Val	Lys 1360
40	Ser	Val	Leu	Glu	Tyr 136		Asp	Arg	Val	Phe 137		Phe	Ile	Phe	Val 137	Phe 5
	Glu	Met	Leu	Leu 1380		Trp	Val	Ala	Tyr 138		Phe	Lys	Lys	Tyr 139	Phe 0	Thr
45	Asn	Ala	Trp 139	Cys 5	Trp	Leu	Asp	Phe 140		Ile	Val	Asn	Ile 140		Leu	Thr
50	Ser	Leu 141		Ala	Lys	Ile	Leu 141		Tyr	Ser	Asp	Val 142	Ala O	Ser	Ile	Lys
50	Ala 142		Arg	Thr	Leu	Arg 143		Leu	Arg	Pro	Leu 143		Ala	Leu	Ser	Arg 1440
55	Phe	Glu	Gly	Met	Arg 144		Val	Val	Asp	Ala 145		Val	Gly	Ala	Ile 145	Pro 5
	Ser	Ile	Met	Asn 146		Leu	Leu	Val	Cys 146		Ile	Phe	Trp	Leu 147	Ile O	Phe
60	Ser	Ile	Met 147	Gly 5	Val	Asn	Leu	Phe 148		Gly	Lys	Phe	Ser 148	Lys 5	Cys	Val
	Asp	Thr 149		Asn	Asn	Pro	Phe	Ser 5	Asn	Val	Asn	Ser 150	Thr 0	Met	Val	Asn

	Asn 150		Ser	Glu	Cys	His 1510		Gln	Asn	Ser	Thr 151		His	Phe	Phe	Trp 1520
5	Val	Asn	Val	Lys	Val 1525		Phe	Asp	Asn	Val 1530		Met	Gly	Tyr	Leu 1539	
10	Leu	Leu	Gln	Val 1540		Thr	Phe	Lys	Gly 1549	-	Met	Asp	Ile	Met 1550	-	Ala
	Ala	Val	Asp 1555		Gly	Glu	Ile	Asn 1560		Gln	Pro	Asn	Trp 1569		Asn	Asn
15	Leu	Tyr 1570		Tyr	Leu	Tyr	Phe 1575	Val	Val	Phe	Ile	Ile 1580		Gly	Gly	Phe
	Phe 1585		Leu	Asn	Leu	Phe 1590		Gly	Val	Ile	Ile 1599		Asn	Phe	Asn	Gln 1600
20	Gln	Lys	Lys	Lys	Leu 1605		Gly	Gln	Asp	Ile 1610		Met	Thr	Glu	Glu 1615	
25	Lys	Lys	Tyr	Tyr 1620		Ala	Met	Lys	Lys 1629		Gly	Ser	Lys	Lys 1630		Gln
	Lys	Pro	Ile 1635		Arg	Pro	Leu	Asn 1640		Tyr	Gln	Gly	Phe 1645		Phe	Asp
30	Ile	Val 1650		Arg	Gln	Ala	Phe 1655	Asp	Ile	Ile	Ile	Met 1660		Leu	Ile	Cys
	Leu 1669		Met	Ile	Thr	Met 1670		Val	Glu	Thr	Asp 1675		Gl'n	Gly	Glu	Glu 1680
35	Lys	Thr	Lys	Val	Leu 1685		Arg	Ile	Asn	Gln 1690		Phe	Val	Ala	Val 1695	
40	Thr	Gly	Glu	Cys 1700		Met	Lys	Met	Phe 1705		Leu	Arg	Gln	Tyr 1710	-	Phe
	Thr	Asn	Gly 1715	_	Asn	Val	Phe	Asp 1720		Ile	Val	Val	Ile 1725		Ser	Ile
45	Gly	Ser 1730		Leu	Phe	Ser	Ala 1735	Ile	Leu	Lys	Ser	Leu 1740		Asn	Tyr	Phe
	Ser 1749		Thr	Leu	Phe	Arg 1750		Ile	Arg	Leu	Ala 1755		Ile	Gly	Arg	Ile 1760
50	Leu	Arg	Leu	Ile	Arg. 1765		Ala	Lys	Gly	Ile 1770		Thr	Leu	Leu	Phe 1775	
55	Leu	Met	Met	Ser 1780		Pro	Ala	Leu	Phe 1785		Ile	Gly	Leu	Leu 1790		Phe
			1799	5				Ile 1800)				1805	5		
60	Val	Val 1810		Glu	Ala	Gly	Ile 1815	Asp	Asp	Met	Phe	Asn 1820		Lys	Thr	Phe
	Gly 1825		Ser	Met	Leu	Cys 1830		Phe	Gln	Ile	Thr 1835		Ser	Ala	Gly	Trp 1840

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	Asp	Gly	Leu	Leu	Ser 1845		Ile	Leu	Asn	Thr 1850	Gly)	Pro	Pro	Tyr	Cys 1855	Asp
5	Pro	Asn	Leu	Pro 1860	Asn)	Ser	Asn	Gly	Ser 1865		Gly	Asn	Cys	Gly 1870	Ser)	Pro
10	Ala	Val	Gly 1875		Ile	Phe	Phe	Thr 1880		Tyr	Ile	Ile	Ile 1885		Phe	Leu
10	Ile	Val 1890		Asn	Met	Tyr	Ile 1895		Val	Ile	Leu	Glu 1900		Phe	Asn	Val
15	Ala 1905		Glu	Glu	Ser	Thr 1910		Pro	Leu	Ser	Glu 1915		Asp	Phe	Asp	Met 1920
	Phe	Tyr	Glu	Thr	Trp 1925		Lys	Phe	Asp	Pro 1930		Ala	Thr	Gln	Phe 1935	
20	Ala	Phe	Ser	Ala 1940	Leu)	Ser	Asp	Phe	Ala 1949		Thr	Leu	Ser	Gly 1950		Leu
25	Arg	Ile	Pro 1955		Pro	Asn	Gln	Asn 1960		Leu	Ile	Gln.	Met 1965		Leu	Pro
	Leu	Val 1970		Gly	Asp	Lys	Ile 1975		Cys	Leu	Asp	Ile 1980		Phe	Ala	Phe
30	1989	5			Leu	1990)				1995	5				2000
					Lys 2005	5				2010)				2015	5
35				2020					2025	5				2030)	
40			203	5	Lys			2040)				2049	5		
		205	0		Thr		2059	5				2060	0			
45	2069	5	•		Glu	2070	כ				2079	5				2080
					Glu 2085	5				2090)				2095	5
50				210					210	5				2110)	
55			211	5	Asn	Glu	Asp	Glu 212		Ala	Ala	Lys	Glu 2125		Asn	Ser
	Pro	Gly 213	Pro 0	Gln												

- 60 (2) INFORMATION FOR SEQ ID NO:7:
- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 6527 base pairs
 (B) TYPE: nucleic acid

							SS: line	_	gle								
5	•	(ii)	MOL	ECUL	E TY	PE:	cDNA										
10		(ix)		.) NA	ME/K	EY:	CDS 204.	.607	77								
		(xi)	SEÇ	UENC	E DE	SCRI	PTIC	N: S	SEQ I	ON C	:7:						
	TAGO	TTGC	TT C	TGCT	'AATC	C TA	'CCC	AGGC	CTI	TAGA	CAG	AGAA	CAGA	TG G	CAGA	TGGAG	60
15	TTTC	TTAT	TG C	CATO	CGCA	A AC	GCTG	SAGCO	CAC	CTCA	TGA	TCCC	GGAC	cc c	ATGO	TTTTC	120
	AGTA	GAC	AAC C	TGGG	CTAP	G AA	GAGA	TCTC	CGA	CCTI	ATA	GAGC	AGCA	AA G	AGTO	TAAAT	180
20	TCTT	cccc	CAA G	AAGA	ATGA	AG AA				C CC eu Pr					_		230
25			AAT Asn														278
30			ATT Ile														326
35			CAG Gln														374
,,			TGT Cys 60														422
40			GGG Gly														470
45			TTC Phe		Val		Asn	Lys	Ser		Thr	Ile			Phe		518
50	GCC Ala	ACT Thr	TGG Trp	GCC Ala	CTG Leu 110	TGG Trp	CTC Leu	TTC Phe	AGT Ser	CCC Pro 115	TTC Phe	AAC Asn	CTG Leu	ATC Ile	AGA Arg 120	AGA Arg	566
	ACA Thr	GCC Ala	ATC Ile	AAA Lys 125	GTG Val	TCT Ser	GTC Val	CAT His	TCC Ser 130	TGG Trp	TTC Phe	TCC Ser	ATA Ile	TTC Phe 135	ATC Ile	ACC Thr	614
55	ATC Ile	Thr	ATT Ile 140	TTG Leu	GTC Val	AAC Asn	TGC Cys	GTG Val 145	TGC Cys	ATG Met	ACC Thr	CGA Arg	ACT Thr 150	GAT Asp	CTT Leu	CCA Pro	662
60	GAG Glu	AAA Lys 155	GTC Val	GAG Glu	TAC Tyr	GTC Val	TTC Phe 160	Thr	GTC Val	ATT Ile	TAC Tyr	ACC Thr 165	TTC Phe	GAG Glu	GCT Ala	CTG Leu	710

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	ATT Ile 170	AAG Lys	ATA Ile	CTG Leu	GCA Ala	AGA Arg 175	GGG Gly	TTT Phe	TGT Cys	CTA Leu	AAT Asn 180	GAG Glu	TTC Phe	ACT Thr	TAT Tyr	CTT Leu 185	758
5	CGA Arg	GAT Asp	CCG Pro	TGG Trp	AAC Asn 190	TGG Trp	CTG Leu	GAC Asp	TTC Phe	AGT Ser 195	GTC Val	ATT Ile	ACC Thr	TTG Leu	GCG Ala 200	TAT Tyr	806
10	GTG Val	GGT Gly	GCA Ala	GCG Ala 205	ATA Ile	GAC Asp	CTC Leu	CGA Arg	GGA Gly 210	ATC Ile	TCA Ser	GGC Gly	CTG Leu	CGG Arg 215	ACA Thr	TTC Phe	854
15	CGA Arg	GTT Val	CTC Leu 220	AGA Arg	GCC Ala	CTG Leu	AAA Lys	ACT Thr 225	GTT Val	TCT Ser	GTG Val	ATC Ile	CCA Pro 230	GGA Gly	CTG Leu	AAG Lys	902
20	GTC Val	ATC Ile 235	GTG Val	GGA Gly	GCC Ala	CTG Leu	ATC Ile 240	CAC His	TCA Ser	GTG Val	AGG Arg	AAG Lys 245	CTG Leu	GCC Ala	GAC Asp	GTG Val	950
20	ACT Thr 250	ATC Ile	CTC Leu	ACA Thr	GTC Val	TTC Phe 255	TGC Cys	CTG Leu	AGC Ser	GTC Val	TTC Phe 260	GCC Ala	TTG Leu	GTG Val	GGC Gly	CTG Leu 265	998
25	CAG Gln	CTC Leu	TTT Phe	AAG Lys	GGG Gly 270	AAC Asn	CTT Leu	AAG Lys	AAC Asn	AAA Lys 275	TGC Cys	ATC Ile	AGG Arg	AAC Asn	GGA Gly 280	ACA Thr	1046
30	GAT Asp	CCC Pro	CAC His	AAG Lys 285	GCT Ala	GAC Asp	AAC Asn	CTC	TCA Ser 290	TCT Ser	GAA Glu	ATG Met	GCA Ala	GAA Glu 295	TAC Tyr	ATC Ile	1094
35	TTC Phe	ATC Ile	AAG Lys 300	CCT Pro	GGT Gly	ACT Thr	ACG Thr	GAT Asp 305	CCC Pro	TTA Leu	CTG Leu	TGC Cys	GGC Gly 310	AAT Asn	GGG Gly	TCT Ser	1142
40	GAT Asp	GCT Ala 315	GGT Gly	CAC His	TGC Cys	CCT Pro	GGA Gly 320	GGC Gly	TAT Tyr	GTC Val	TGC Cys	CTG Leu 325	AAA Lys	ACT Thr	CCT Pro	GAC Asp	1190
	Asn 330	Pro	Asp	Phe	Asn	TAC Tyr 335	Thr	Ser	Phe	Asp	Ser 340	Phe	Ala	Tṛp	Ala	Phe 345	1238
45	CTC Leu	TCA Ser	CTG Leu	TTC Phe	CGC Arg 350	CTC Leu	ATG Met	ACG Thr	CAG Gln	GAC Asp 355	TCC Ser	TGG Trp	GAG Glu	CGC Arg	CTG Leu 360	Tyr	1286
50	Gln	Gln	Thr	Leu 365	Arg	GCT Ala	Ser	Gly	Lys 370	Met	Tyr	Met	Val	Phe 375	Phe	Val	1334
55	CTG Leu	GTT Val	ATT Ile 380	Phe	CTT Leu	GGA Gly	TCG Ser	TTC Phe 385	Tyr	CTG Leu	GTC Val	AAT Asn	TTG Leu 390	Ile	TTG Leu	GCC Ala	1382
60	GTG Val	GTC Val 395	Thr	ATG Met	GCG Ala	TAT Tyr	GAA Glu 400	GAG Glu	CAG Gln	AGC Ser	CAG Gln	GCA Ala 405	Thr	ATT Ile	GCA Ala	GAA Glu	1430
33	ATC Ile 410	Glu	. GCC . Ala	AAG Lys	GAA Glu	AAA Lys 415	Lys	TTC Phe	CAG Gln	GAA Glu	GCC Ala 420	Leu	GAG Glu	GTG Val	CTG Leu	CAG Gln 425	1478

						CTG Leu											1526
5						TCA Ser											1574
10						TCA Ser											1622
15						GAC Asp											1670
20						AGA Arg 495											1718
20						CAA Gln											1766
25						CAC His											1814
30	TTG Leu	CTG Leu	GGC Gly 540	AGG Arg	GGT Gly	GCT Ala	GGG Gly	CAG Gln 545	ACA Thr	GGT Gly	CCA Pro	CTC Leu	CCC Pro 550	AGG Arg	AGC Ser	CCA Pro	1862
35	CTG Leu	CCT Pro 555	CAG Gln	TCC Ser	CCC Pro	AAC Asn	CCT Pro 560	GGC Gly	CGT Arg	AGA Arg	CAT His	GGA Gly 565	GAA Glu	GAG Glu	GGA Gly	CAG Gln	1910
40						GGT Gly 575											1958
40						GGG Gly											2006
45						GCA Ala											2054
50						GAG Glu											2102
55						GCT Ala											2150
63						AAG Lys 655											2198
60						ATC Ile											2246

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	ATG Met	GCC Ala	ATG Met	GAG Glu 685	CAC His	TAC Tyr	CCC Pro	ATG Met	ACC Thr 690	GAT Asp	GCC Ala	TTC Phe	GAT Asp	GCC Ala 695	ATG Met	CTT Leu	2294
5						GTC Val											2342
10						TTC Phe											2390
15						GTC Val 735											2438
20						AGC Ser											2486
						GCC Ala											2534
25						TCA Ser											2582
30						TTC Phe											2630
35						TGC Cys 815											2678
40						CAC His											2726
	Val	Phe	Arg	Ile 845	Leu	TGC Cys	Gly	Glu	Trp 850	Ile	Glu	Asn	Met	Trp 855	Val	Cys	2774
45						AAA Lys											2822
50 .						CTA Leu											2870
55						GCG Ala 895											2918
60						CAG Gln											2966
						GCC Ala											3014

											ATG Met						3062
5											GAT Asp 965						3110
10											AGC Ser						3158
- 15											GTC Val				Ile		3206
20				Ser					Leu		GAA Glu			Glu			3254
			Ser					Glu			AAG Lys		Gln				3302
25		Pro					Cys				CAG Gln 1045	Ala					3350
30	Ala					Ser					CCA Pro					٠	3398
35					Asp					Pro	GCC Ala				Asp	,	3446
40				Ser					Val		TGC Cys			Pro			3494
			Arg					Leu			GAC Asp		Asp				3542
45		Cys					Cys				TGT Cys 1129	Pro					3590
50	Asn					Pro					TGG Trp						3638
55					Val					Phe	GAG Glu				Ile		3686
60				Leu					Leu		TTT Phe			Asn			3734
			Lys					Ser			GAG Glu		Thr				3782

	GTG Val	TTC Phe 1195	Thr	TTC Phe	ATC Ile	TTC Phe	GTC Val 1200	Phe	GAG Glu	ATG Met	CTG Leu	CTC Leu 1205	Lys	TGG Trp	GTA Val	GCC Ala	3830
5	TAT Tyr 1210	Gly	TTC Phe	AAA Lys	AAG Lys	TAT Tyr 1215	Phe	ACC Thr	AAT Asn	GCC Ala	TGG Trp 1220	Cys	TGG Trp	CTG Leu	GAC Asp	TTC Phe 1225	3878
10	CTC Leu	ATT Ile	GTG Val	AAC Asn	ATC Ile 1230	Ser	CTG Leu	ACA Thr	AGC Ser	CTC Leu 1235	Ile	GCG Ala	AAG Lys	ATC Ile	CTT Leu 1240	Glu	3926
15					Ala					Leu					GCC Ala		3974
20				Arg					Phe					Val	GTG Val		4022
20	GAT Asp	GCC Ala 1275	Leu	GTG Val	GGC Gly	GCC Ala	ATC Ile 1280	Pro	TCC Ser	ATC Ile	ATG Met	AAC Asn 1285	Val	CTC Leu	CTC Leu	GTC Val	4070
25		Leu					Ile					Gly			CTC Leu		4118
30						Lys					Arg				TTT Phe 1320	Ser	4166
35	AAC Asn	GTG Val	AAT Asn	TCG Ser 1325	Thr	ATG Met	GTG Val	AAT Asn	AAC Asn 1330	Lys	TCC Ser	GAG Glu	TGT Cys	CAC His 133	AAT Asn 5	CAA Gln	4214
40	AAC Asn	AGC Ser	ACC Thr 1340	Gly	CAC His	TTC Phe	TTC Phe	TGG Trp 1349	Val	AAC Asn	GTC Val	AAA Lys	GTC Val 1350	Asn	TTC Phe	GAC Asp	4262
40			Ala					Ala					Ala		TTC Phe		4310
45	GGC Gly 1370	Trp	ATG Met	GAC Asp	ATA Ile	ATG Met 137	Tyr	GCA Ala	GCT Ala	GTT Val	GAT Asp 1380	Ser	GGA Gly	GAG Glu	ATC Ile	AAC Asn 1385	4358
50	AGT Ser	CAG Gln	CCT Pro	AAC Asn	TGG Trp 139	Glu	AAC Asn	AAC Asn	TTG Leu	TAC Tyr 1395	Met	TAC Tyr	CTG Leu	TAC Tyr	TTC Phe 1400	Val	4406
55	GTT Val	TTC Phe	ATC Ile	ATT Ile 140	Phe	GGT Gly	GGC Gly	TTC Phe	TTC Phe 141	Thr	CTG Leu	AAT Asn	CTC Leu	TTT Phe 141	GTT Val 5	GGG Gly	4454
60				Asp					Gln					Gly	GGC Gly		4502
60			Phe					Gln					Asn		ATG Met		4550

	AAG CTG Lys Leu 1450	GGC TC Gly Se	C AAG r Lys	AAA Lys 1455	Pro	CAG Gln	AAG Lys	CCC Pro	ATC Ile 1460	Pro	CGG Arg	CCC Pro	CTG Leu	AAT Asn 1465	4598
5	AAG TAC Lys Tyr	CAA GG Gln Gl	C TTC y Phe 1470	Val	TTT Phe	GAC Asp	ATC Ile	GTG Val 1475	Thr	AGG Arg	CAA Gln	GCC Ala	TTT Phe 1480	Asp	4646
10	ATC ATC Ile Ile	ATC AT Ile Me 14	t Val	CTC . Leu	ATC Ile	TGC Cys	CTC Leu 1490	Asn	ATG Met	ATC Ile	ACC Thr	ATG Met 1495	Met	GTG Val	4694
15	GAG ACC Glu Thr	GAC GA Asp Gl 1500	G CAG u Gln	GGC (Glu	GAG Glu 1505	Lys	ACG Thr	AAG Lys	GTT Val	CTG Leu 1510	Gly	AGA Arg	ATC Ile	4742 ·
20.	AAC CAG Asn Gln 151	Phe Ph	T GTG e Val	Ala	GTC Val 1520	Phe	ACG Thr	GGC Gly	GAG Glu	TGT Cys 1525	Val	ATG Met	AAG Lys	ATG Met	4790
20 [.]	TTC GCC Phe Ala 1530	CTG CG Leu Ar	A CAG g Gln	TAC Tyr 1535	Tyr	TTC Phe	ACC Thr	AAC Asn	GGC Gly 1540	Trp	AAC Asn	GTG Val	TTC Phe	GAC Asp 1545	4838
25	TTC ATA Phe Ile	GTG GT Val Va	G ATC 1 Ile 1550	Leu	TCC Ser	ATT Ile	GGG Gly	AGT Ser 1555	Leu	CTG Leu	TTT Phe	TCT Ser	GCA Ala 1560	Ile	4886
30	CTT AAG Leu Lys	Ser Le	G GAA u Glu 65	AAC Asn	TAC Tyr	TTC Phe	TCC Ser 1570	Pro	ACG Thr	CTC Leu	TTC Phe	CGG Arg 1579	Val	ATC Ile	4934
35	CGT CTG Arg Leu	GCC AG Ala Ar 1580	G ATC g Ile	GGC Gly	CGC Arg	ATC Ile 1585	Leu	AGG Arg	CTG Leu	ATC Ile	CGA Arg 1590	Ala	GCC Ala	AAG Lys	4982
40	GGG ATT Gly Ile 159	Arg Th				Ala					Leu				5030
40	TTC AAC Phe Asn 1610	ATC GG	CTC y Leu	CTC Leu 1615	Leu	TTC Phe	CTC Leu	GTC Val	ATG Met 1620	Phe	ATC Ile	TAC Tyr	TCC Ser	ATC Ile 1625	5078
45	TTC GGC Phe Gly	ATG GO Met Al	C AGC a Ser 163	Phe	GCT Ala	AAC Asn	GTC Val	GTG Val 1635	Asp	GAG Glu	GCC Ala	GGC Gly	ATC Ile 164	Asp	5126
50	GAC ATG Asp Met	Phe As	C TTC in Phe 545	AAG Lys	ACC Thr	TTT Phe	GGC Gly 165	Asn	AGC Ser	ATG Met	CTG Leu	TGC Cys 165	Leu	TTC Phe	5174
55	CAG ATO	ACC AC Thr Th	CC TCG ir Ser	GCC Ala	GGC Gly	TGG Trp 166	Asp	GGC Gly	CTC Leu	CTC Leu	AGC Ser 1670	Pro	ATC Ile	CTC Leu	5222
60	AAC ACG Asn Thr 167	Gly P	CT CCC	TAC Tyr	TGC Cys 168	Asp	CCC Pro	AAC Asn	CTG Leu	CCC Pro 168	Àsn	AGC Ser	AAC _Asn	GGC Gly	5270
	TCC CGC Ser Arc 1690	GGG A	AC TGC an Cys	GGG Gly 1699	Ser	CCG Pro	GCG Ala	GTG Val	GGC Gly 170	Ile	ATC Ile	TTC Phe	TTC Phe	ACC Thr 1705	5318

																		-266
	ACC TA	AC A yr I	TC .	Ile	ATC Ile 1710	Ser	TTC Phe	CTC Leu	ATC Ile	GTG Val 1715	Val	AAC Asn	Met	TAC Tyr	Ile 1720	Ala	-	5366
5	GTG AT	TT C le L	eu	GAG Glu 1725	Asn	TTC Phe	AAC Asn	GTA Val	GCC Ala 1730	Thr	GAG Glu	GAG Glu	AGC Ser	ACG Thr 1735	Glu	CCC Pro	Š	5414
10	CTG AC	er G	AG lu .740	Asp	GAC Asp	TTC Phe	GAC Asp	ATG Met 1745	Phe	TAT Tyr	GAG Glu	ACC Thr	TGG Trp 1750	Glu	AAG Lys	TTC Phe	<u>:</u>	5462
15	GAC CO Asp Pi	CG G ro G 755	AG Slu	GCC Ala	ACC Thr	CAG Gln	TTC Phe 1760	Ile	GCC Ala	TTT Phe	TCT Ser	GCC Ala 1765	Leu	TCA Ser	GAC Asp	TTC Phe		5510
20	GCG GA Ala As 1770	AC A sp T	CG hr	CTC Leu	TCC Ser	GGC Gly 1775	Pro	CTT Leu	AGA Arg	ATC Ile	CCC Pro 1780	Lys	CCC Pro	AAC Asn	CAG Gln	AAT Asn 1785	ţ	5558
20	ATA T	TA A eu I	TC le	CAG Gln	ATG Met 1790	Asp	CTG Leu	CCG Pro	TTG Leu	GTC Val 1795	Pro	GGG Gly	GAT Asp	AAG Lys	ATC Ile 1800	His		5606
25	TGT C	TG G eu A	Asp	ATC Ile 1805	Leu	TTT Phe	GCC Ala	TTC Phe	ACA Thr 1810	Lys	AAC Asn	GTC Val	TTG Leu	GGA Gly 181	Glu	TCC Ser	!	5654
30	GGG G	lu I	rTG Leu L820	Asp	TCC Ser	CTG Leu	AAG Lys	ACC Thr 1825	Asn	ATG Met	GAA Glu	GAG Glu	AAG Lys 1830	Phe	ATG Met	GCG Ala	:	5702
35	ACC A. Thr A.	AT C sn I 835	CTC Leu	TCC Ser	AAA Lys	GCA Ala	TCC Ser 1840	Tyr	GAA Glu	CCA Pro	ATA Ile	GCC Ala 1849	Thr	ACC Thr	CTC Leu	CGG Arg	1	5750
40	TGG A Trp L 1850	AG (CAG	GAA Glu	GAC Asp	CTC Leu 1859	Ser	GCC Ala	ACA Thr	GTC Val	ATT Ile 1860	Gln	AAG Lys	GCC Ala	TAC Tyr	CGG Arg 1865		5798
40	AGC T Ser T	AC A	ATG Met	CTG Leu	CAC His 187	Arg	TCC Ser	TTG Leu	ACA Thr	CTC Leu 1879	Ser	AAC Asn	ACC Thr	CTG Leu	CAT His 1880	Val		5846
45	CCC A Pro A	.GG (GCT Ala	GAG Glu 1885	Glu	GAT Asp	GGC Gly	GTG Val	TCA Ser 189	Leu	CCC Pro	GGG Gly	GAA Glu	GGC Gly 189	Tyr	AGT Ser		5894
50	ACA T	he l	ATG Met 1900	Ala	AAC Asn	AGT Ser	GGA Gly	CTC Leu 190	Pro	GAC Asp	AAA Lys	TCA Ser	GAA Glu 191	Thr	GCC Ala	TCT Ser		5942
55	GCT A Ala T	CG '	Ser	TTC Phe	CCG Pro	CCA Pro	TCC Ser 192	Tyr	GAC Asp	AGT Ser	GTC Val	ACC Thr 192	Arg	GGC Gly	CTG Leu	AGT Ser		5990
	GAC C Asp A 1930	GG (GCC Ala	AAC Asn	ATT Ile	AAC Asn 193	Pro	TCT Ser	AGC Ser	TCA Ser	ATG Met 194	Gln	AAT Asn	GAA Glu	GAT Asp	GAG Glu 1945		6038
6 0	GTC C	SCT (GCT Ala	AAG Lys	GAA Glu 195	Gly	AAC Asn	AGC Ser	CCT Pro	GGA Gly 195	Pro	CAG Gln	TGA	AGGC	ACT			6084

	CAG	GCAT	GCA	CAGG	GCAG	GT T	'CCAA	TGTC	т тт	CTCT	GCTG	TAC	TAAC	TCC	TTCC	CTCTG	3	6144
	AGG	TGGC	ACC .	AACC'	TCCA	SC C	TCCA	CCAA	T GC	ATGT	CACT	GGT	CATG	GTG	TCAG	AACTG	A	6204
5	ATG	GGGA	CAT	CCTT	GAGA/	AA G	cccc	CACC	C CA	ATAG	GAAT	CAA	AAGC	CAA	GGAT	ACTCC	r	6264
	CCA	TTCT	GAC (GTCC	CTTC	CG A	GTTC	CCAG	A AG	ATGT	CATT	GCT	CCCT	TCT	GTTT	GTGAC	2	6324
10	AGA	GACG	TGA '	TTCA	CCAAC	ст т	CTCG	GAGC	C AG	AGAC.	ACAT	AGC	AAAG	ACT	TTTC	TGCTG	3	6384
10	TGT	CGGG	CAG '	TCTT	AGAG <i>I</i>	AA G	TCAC	GTAG	G GG	TTGG	TACT	GAG	AATT.	AGG	GTTT	GCATG	A	6444
	CTG	CATG	CTC	ACAG	CTGCC	G G	ACAA	TACC	r gt	GAGT	CGGC	CAT	TAAA	АТТ	AATA	TTTTT	A	6504
15	AAG'	TTAA	AAA A	AAAA	AAAA	AA A	AA											6527
	(2)	INF	ORMA'	TION	FOR	SEQ	ID :	NO:8	:									
20			(i) :	(A)	ENCE) LEN) TYP) TOP	IGTH PE:	: 19 amin	57 ar o ac:	mino id		ds							
25		(:	ii) 1	MOLE	CULE	TYP	E: p	rote	in									
		(:	xi) :	SEQUI	ENCE	DES	CRIP'	TION	: SE	Q ID	NO:8	B:					•	
30	Met 1	Glu	Leu	Pro	Phe 5	Ala	Ser	Val	Gly	Thr 10	Thr	Asn	Phe	Arg	Arg 15	Phe		
	Thr	Pro	Glu	Ser 20	Leu	Ala	Glu	Ile	Glu 25	Lys	Gln	Ile	Ala	Ala 30		Arg		
35	Ala	Ala	Lys 35	Lys	Ala	Arg	Thr	Lys 40	His	Arg	Gly	Gln	Glu 45	Asp	Lys	Gly		
40	Glu	Lys 50	Pro	Arg	Pro	Gln	Leu 55	Asp	Leu	Lys	Asp	Cys 60	Asn	Gln	Leu	Pro		
•	Lys 65	Phe	Tyr	Gly	Glu	Leu 70	Pro	Ala	Glu	Leu	Val 75	Gly	Glu	Pro	Leu	Glu 80		
45	Asp	Leu	Asp	Pro	Phe 85	Tyr	Ser	Thr	His	Arg 90	Thr	Phe	Met	Val	Leu 95	Asn		
	Lys	Ser	Arg	Thr 100	Ile	Ser	Arg	Phe	Ser 105	Ala	Thr	Trp	Ala	Leu 110	Trp	Leu		
50	Phe	Ser	Pro 115	Phe	Asn	Leu	Ile	Arg 120	Arg	Thr	Ala	Ile	Lys 125	Val	Ser	Val		
55	His	Ser 130	Trp	Phe	Ser	Ile	Phe 135	Ile	Thr	Ile	Thr	Ile 140	Leu	Val	Asn	Cys		
	Val 145	Cys	Met	Thr	Arg	Thr 150	Asp	Leu	Pro	Glu	Lys 155	Val	Glu	Tyr	Val	Phe 160		
60	Thr	Val	Ile	Tyr	Thr 165	Phe	Glu	Ala	Leu	Ile 1	Lys	Ile	Leu	Ala	Arg 175	Gly		
	Phe	Cys	Leu	Asn 180	Glu	Phe	Thr	Tyr	Leu 185	Arg	Asp	Pro	Trp	Asn 190		Leu		

	Asp	Phe	Ser 195	Val	Ile	Thr	Leu	Ala 200	Tyr	Val	Gly	Ala	Ala 205	Ile	Asp	Leu
5	Arg	Gly 210	Ile	Ser	Gly	Leu	Arg 215	Thr	Phe	Arg	Val	Leu 220	Arg	Ala	Leu	Lys
	Thr 225	Val	Ser	Val	Ile	Pro 230	Gly	Leu	Lys	Val	Ile 235	Val	Gly	Ala	Leu	Ile 240
10	His	Ser	Val	Arg	Lys 245	Leu	Ala	Asp	Val	Thr 250	Ile	Leu	Thr	Val	Phe 255	Cys
15	Leu	Ser	Val	Phe 260	Ala	Leu	Val	Gly	Leu 265	Gln	Leu	Phe	Lys	Gly 270	Asn	Leu
1,7	Lys	Asn	Lys 275	Cys	Ile	Arg	Asn	Gly 280	Thr	Asp	Pro	His	Lys 285	Ala	Asp	Asn
20	Leu	Ser 290	Ser	Glu	Met	Ala	Glu 295	Tyr	Ile	Phe	Ile	Lys 300	Pro	Gly	Thr	Thr
	Asp 305	Pro	Leu	Leu	Cys	Gly 310	Asn	Gly	Ser	Asp	Ala 315	Gly	His	Cys	Pro	Gly 320
25	Gly	Tyr	Val	Cys	Leu 325	Lys	Thr	Pro	Asp	Asn 330	Pro	Asp	Phe	Asn	Tyr 335	Thr
30	Ser	Phe	Asp	Ser 340	Phe	Ala	Trp	Ala	Phe 345	Leu	Ser	Leu	Phe	Arg 350	Leu	Met
	Thr	Gln	Asp 355	Ser	Trp	Glu	Arg	Leu 360	Tyr	Gln	Gln	Thr	Leu 365	Arg	Ala	Ser
35		Lys 370	Met	Tyr	Met	Val	Phe 375	Phe	Val	Leu	Val	Ile 380	Phe	Leu	Gly	Ser
40	Phe 385	Tyr	Leu	Val	Asn	Leu 390	Ile	Leu	Ala	Val	Val 395	Thr	Met	Ala	Tyr	Glu 400
40	Glu	Gln	Ser	Gln	Ala 405	Thr	Ile	Ala	Glu	Ile 410	Glu	Ala	Lys	Glu	Lys 415	Lys
45	Phe	Gln						Leu		Lys		Gln	Glu	Val 430	Leu	Ala
	Ala	Leu	Gly 435	Ile	Asp	Thr	Thr	Ser 440	Leu	Gln	Ser	His	Ser 445	Gly	Ser	Pro
50	Leu	Ala 450	Ser	Lys	Asn	Ala	Asn 455	Glu	Arg	Arg	Pro	Arg 460	Val	Lys	Ser	Arg
55	Val 465	Ser	Glu	Gly	Ser	Thr 470	Asp	Asp	Asn	Arg	Ser 475	Pro	Gln	Ser	Asp	Pro 480
,,	Tyr	Asn	Gln	Arg	Arg 485	Met	Ser	Phe	Leu	Gly 490	Leu	Ser	Ser	Gly	Arg 495	Arg
60	Arg	Ala	Ser	His 500		Ser	Val	Phe	'His 505	Phe	Arg	Ala	Pro	Ser 510	Gln	Asp
	Ile	Ser	Phe 515	Pro	Asp	Gly	Ile	Thr 520	Pro	Asp	Asp	Gly	Val 525	Phe	His	Gly

	Asp	Gln 530	Glu	Ser	Arg	Arg	Gly 535	Ser	Ile	Leu	Leu	Gly 540	Arg	Gly	Ala	Gly
5	Gln 545	Thr	Gly	Pro	Leu	Pro 550	Arg	Ser	Pro	Leu	Pro 555	Gln	Ser	Pro	Asn	Pro 560
	Gly	Arg	Arg	His	Gly 565	Glu	Glu	Gly	Gln	Leu 570	Gly	Val	Pro	Thr	Gly 575	Glu
10	Leu	Thr	Ala	Gly 580	Ala	Pro	Glu	Gly	Pro 585	Ala	Leu	Asp	Thr	Thr 590	Gly	Gln
15	Lys	Ser	Phe 595	Leu	Ser	Ala	Gly	T yr 600	Leu	Asn	Glu	Pro	Phe 605	Arg	Ala	Gln
	Arg	Ala 610	Met	Ser	Val	Val	Ser 615	Ile	Met	Thr	Ser	Val 620	Ile	Glu	Glu	Leu
20	Glu 625	Glu	Ser	Lys	Leu	Lys 630	Cys	Pro	Pro	Cys	Leu 635	Ile	Ser	Phe	Ala	Gln 640
	Lys	Tyr	Leu	Ile	Trp 645	Glu	Cys	Cys	Pro	Lys 650.		Arg	Lys	Phe	Lys 655	Met
25	Ala	Leu	Phe	Glu 660	Leu	Val	Thr	Asp	Pro 665	Phe	Ala	Glu	Leu	Thr 670	Ile	Thr
30	Leu	Cys	Ile 675	Val	Val	Asn	Thr	Val 680	Phe	Met	Ala	Met	Glu 685	His	Tyr	Pro
	Met	Thr 690	Asp	Ala	Phe	Asp	Ala 695	Met	Leu	Gln	Ala	Gly 700	Asn	Ile	Val	Phe
35	Thr 705	Val	Phe	Phe	Thr	Met 710	Glu	Met	Ala	Phe	Lys 715	Ile	Ile	Ala	Phe	Asp 720
	Pro	Tyr	Tyr	Tyr	Phe 725	Gln	Lys	Lys	Trp	Asn 730	Ile	Phe	Asp	Cys	Val 735	Ile
40	Val	Thr	Val	Ser 740	Leu	Leu	Glu	Leu	Ser 7 4 5	Ala	Ser	Lys	Lys	Gly 750	Ser	Leu
45	Ser	Val	Leu 755	Arg	Ser	Leu	Arg	Leu 760	Leu	Arg	Val	Phe	Lys 765	Leu	Ala	Lys
	Ser	Trp 770	Pro	Thr	Leu	Asn	Thr 775	Leu	Ile	Lys	Ile	Ile 780	Gly	Asn	Ser	Val
50	Gly 785	Ala	Leu	Gly	Asn	Leu 790	Thr	Pne	Ile	Leu	Ala 795	Ile	Ile	Val	Phe	Ile 800
	Phe	Ala	Leu	Val	Gly 805	Lys	Gln	Leu	Leu	Ser 810	Glu	Asp	Tyr	Gly	Cys 815	Arg
55	Lys	Asp	Gly	Val 820	Ser	Val	Trp	Asn	Gly 825	Glu	Lys	Leu	Arg	Trp 830	His	Met
60			835	•	His			840					845			
		850			Asn		855					860				
	Ile	Cys	Leu	Ile	Leu	Phe	Leu	Thr	Val	Met	Val	Leu	Gly	Asn	Leu	Val

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	865					870					875					880
5	Val L	eu	Asn	Leu	Phe 885	Ile	Ala	Leu	Leu	Leu 890	Asn	Ser	Phe	Ser	Ala 895	Asp
5	Asn L	eu	Thr	Ala 900	Pro	Glu	Asp	Asp	Gly 905	Glu	Val	Asn	Asn	Leu 910	Gln	Leu
10	Ala L	.eu	Ala 915	Arg	Ile	Gln	Val	Leu 920	Gly	His	Arg	Ala	Ser 925	Arg	Ala	Ile
	Ala S	Ser 30	Tyr	Ile	Ser	Ser	His 935	Cys	Arg	Phe	Arg	Trp 940	Pro	Lys	Val	Glu
15	Thr G	Sln	Leu	Gly	Met	Lys 950	Pro	Pro	Leu	Thr	Ser 955	Ser	Glu	Ala	Lys	Asn 960
20	His I	le	Ala	Thr	Asp 965	Ala	Val	Ser	Ala	Ala 970	Val	Gly	Asn	Leu	Thr 975	Lys
	Pro A	la	Leu	Ser 980	Ser	Pro	Lys	Glu	Asn 985	His	Gly	Asp	Phe	Ile 990	Thr	Asp
25	Pro A	sn	Val 995	Trp	Val	Ser	Val	Pro 1000		Ala	Glu	Gly	Glu 1005		Asp	Leu
30	Asp G	lu .010		Glu	Glu	Asp	Met 1015		Gln	Ala	Ser	Gln 1020		Ser	Trp	Gln
	Glu G 1025	Slu	Asp	Pro	Lys	Gly 1030		Gln	Glu	Gln	Leu 1035		Gln	Val	Gln	Lys 1040
35	Cys G	lu	Asn	His	Gln 1045		Ala	Arg	Ser	Pro 1050		Ser	Met	Met	Ser 1055	
	Glu A	qa	Leu	Ala 1060		Tyr	Leu	Gly	Glu 1065		Trp	Lys	Arg	Lys 1070	-	Ser
40	Pro G	ln	Val 1075		Ala	Glu	Gly	Val 1080		Asp	Thr	Ser	Ser 1085		Glu	Gly
45	Ser 1	hr .090		qzA	Cys	Pro	Asp 1095		Glu	Glu	Ile	Leu 1100	_	Lys	Ile	Pro
	Glu L 1105	eu	Ala	Asp	Asp	Leu 1110		Glu	Pro	Asp	Asp 1115		Phe	Thr	Glu	Gly 1120
50	Cys T	hr	Arg	Arg	Cys 1125		Cys	Cys	Asn	Val 1130		Thr	Ser	Lys	Ser 1135	
	Trp A	la	Thr	Gly 1140	-	Gln	Val	Arg	Lys 1145		Cys	Tyr	Arg	Ile 1150		Glu
55	His S		1155	5				1160)			-	1165	ò		
60		170)				1175	5				1180)			
	Lys S 1185	Ser	Val	Leu	Glu	Tyr 1190		Asp	Arg	Val	Phe 1195		Phe	Ile	Phe	Val 1200

	Phe	Glu	Met	Leu	Leu 1205	Гуs	Trp	Val	Ala	Tyr 121	Gly	Phe	Lys	Lys	Tyr 1215	
5	Thr	Asn _.	Ala	Trp 1220	_	Trp	Leu	Asp	Phe 1225		Ile	Val	Asn	Ile 1230		Leu
	Thr	Ser	Leu 1235		Ala	Lys	Ile	Leu 1240		Tyr	Ser	Asp	Val 1245		Ser	Ile
10	Lys	Ala 1250		Arg	Thr	Leu	Arg 1255		Leu	Arg	Pro	Leu 1260	_	Ala	Leu	Ser
15	Arg 1265		Glu	Gly	Met	Arg 1270		Val	Val	Asp	Ala 1275		Val	Gly	Ala	Ile 1280
	Pro	Ser	Ile	Met	Asn 1285	Val	Leu	Leu	Val	Cys 1290		Ile	Phe	Trp	Leu 1295	
20	Phe	Ser	Ile	Met 1300		Val	Asn	Leu	Phe 1305		Gly	Lys	Phe	Ser 1310		Cys
	Val	Asp	Thr 1315		Asn	Asn	Pro	Phe 1320		Asn	Val	Asn	Ser 1325		Met	Val
25	Asn	Asn 1330	_	Ser	Glu	Cys	His 1335		Gln	Asn	Ser	Thr 1340	_	His	Phe	Phe ·
30	Trp 1349		Asn	Val	Lys	Val 1350		Phe	Asp	Asn	Val 1355		Met	Gly	Tyr	Leu 1360
	Ala	Leu	Leu	Gln	Val 1365	Ala	Thr	Phe	Lys	Gly 1370		Met	Asp	Ile	Met 1379	
35	Ala	Ala	Val	Asp 1380		Gly	Glu	Ile	Asn 1385		Gln	Pro	Asn	Trp 1390		Asn
	Asn	Leu	Tyr 1399		Tyr	Leu	Tyr	Phe 1400		Val	Phe	Ile	Ile 1409		Gly	Gly
40		1410)			Leu	1419	5	-			1420) ·			
45	1429	5				1430)				1439	5				Glu 1440
					1449					145	0				1459	5
50		-		1460	כ	Arg			1469	5	_		-	1470)	
	_		147	5		Gln		148	0				148	5		
55	_	149	0			Thr	149	5				150	0			
60	150	5				Leu 151	0				151	5				1520
	Phe	Thr	Gly	Glu	Cys 152	Val 5	Met	Lys	Met	Phe 153		Leu	Arg	Gln	Tyr 153	

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	Phe	Thr	Asn	Gly 1540	Trp	Asn	Val	Phe	Asp 1545	Phe	Ile	Val	Val	Ile 1550	Leu	Ser
5	Ile	Gly	Ser 1555	Leu	Leu	Phe	Ser	Ala 1560		Leu	Lys	Ser	Leu 1565	Glu	Asn	Tyr
	Phe	Ser 1570		Thr	Leu	Phe	Arg 1575		Ile	Arg	Leu	Ala 1580	Arg)	Ile	Gly	Arg
10	Ile 1585		Arg	Leu	Ile	Arg 1590		Ala	Lys	Gly	Ile 1595	Arg	Thr	Leu	Leu	Phe 1600
15	Ala,	Leu	Met	Met	Ser 1605		Pro	Ala	Leu	Phe 1610		Ile	Gly	Leu	Leu 1615	Leu
13	Phe	Leu	Val	Met 1620		Ile	Tyr	Ser	Ile 1625		Gly	Met	Ala	Ser 1630	Phe	Ala
20	Asn	Val	Val 1635	Asp 5	Glu	Ala	Gly	Ile 1640		Asp	Met	Phe	Asn 1645	Phe	Lys	Thr
	Phe	Gly 1650		Ser	Met	Leu	Cys 1659		Phe	Gln	Ile	Thr 1660		Ser	Ala	Gly
25	Trp 1665		Gly	Leu	Leu	Ser 1670		Ile	Leu	Asn	Thr 1675		Pro	Pro	Tyr	Cys 1680
30				Leu	1689	5				1690)				1695	5
				Gly 1700)				1709	5				1710)	
35			171					1720)				1725	5		
		173	0	Glu			173	5				1740	0			
40	174	5		Glu		1750	0				175	5				1760
45				Ser	176	5				1770	0				1779	5
				Pro 178	0				1789	5				1790)	
50			179					180	0				180	5		
	Phe	Thr 181		Asn	Val	Leu	Gly 181		Ser	Gly	Glu	Leu 182	Asp 0	Ser	Leu	Lys
55	Thr 182		Met	Glu	Glu	Lys 183		Met	Ala	Thr	Asn 183		Ser	Lys	Ala	Ser 1840
60				Ile	184	5				185	0				185	5
	Ala	Thr	Val	Ile 186		Lys	Ala	Tyr	Arg 186	Sè <i>s</i> 5	Tyr	Met	Leu	His 187	Arg 0	Ser

	Leu	Thr	Leu 187		Asn	Thr	Leu	His 188		Pro	Arg	Ala	Glu 1885		Asp	Gly		
5	Val	Ser 189	Leu O	Pro	Gly	Glu	Gly 1899		Ser	Thr	Phe	Met 1900		Asn	Ser	Gly	•	
	Leu 190	Pro 5	Asp	Lys	Ser	Glu 1910		Ala	Ser	Ala	Thr 1915		Phe	Pro	Pro	Ser 1920		-
10	Tyr	Asp	Ser	Val	Thr 1925		Gly	Leu	Ser	Asp 1930		Ala	Asn	Ile	Asn 1935			
15	Ser	Ser	Ser	Met 1940		Asn	Glu	Asp	Glu 1945		Ala	Ala	Lys	Glu 1950		Asn		
	Ser	Pro	Gly 1955		Gln													
20	(2)	INFO	ORMAT	NOI	FOR	SEQ	ID N	10 : 9 :	:									
25		(i)	(F.	QUENC A) LE B) TY C) ST C) TO	ENGTH PE: PRANI	I: 21 nucl DEDNE	. bas .eic ESS:	se pa acio sino	airs 1									
30		(ii)	MOL	LECUI	E TY	PE:	CDNA	A										
		(xi)	SEC	QUENC	E DE	ESCRI	PTIC	ON: S	SEQ 1	D NC	9:9:							
3 5	CAGO	CTTCC	GCT C	CAGA	GTAI	C T												21
	(2)	INFO	ORMAI	CION	FOR	SEQ	ID N	10:10):									
40		(i)	(E	OUENC A) LE B) TY C) ST O) TO	ENGTH PE: PRANI	H: 22 nucl	bas eic SS:	se pa ació sino	irs 1									
45		(ii)	MOI	LECUI	E TY	PE:	CDNA	À.										
		(xi)	SEÇ	QUENC	E DE	ESCRI	PTIC	on: s	SEQ I	D NO	:10:							
50	TTCT	rcgco	CGT 1	CCAC	CACGO	A GA	7											22
	(2)	INFO	ORMAT	поп	FOR	SEQ	ID N	10:11	.:									
55		(i)	(E	QUENC A) LE B) TY	ENGTH	H: 4 amir	amir no ac	no ac										
. 0		(ii)	MOI	LECUI	LE TY	PE:	pept	ide										

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		(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:
		Phe Arg Leu Met 1
5	(2)	INFORMATION FOR SEQ ID NO:12:
10		(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 9 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
		(ii) MOLECULE TYPE: peptide
15		
		(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:
20		Thr Gln Asp Phe Trp Glu Asn Leu Tyr 1 5
	(2)	INFORMATION FOR SEQ ID NO:13:
25		(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 9 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
30		(ii) MOLECULE TYPE: peptide
		(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:
35		Thr Gln Asp Tyr Trp Glu Asn Leu Tyr 1 5
	(2)	INFORMATION FOR SEQ ID NO:14:
40		(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 9 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear
45		(ii) MOLECULE TYPE: peptide
50		(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:
50		Thr Gln Asp Cys Trp Glu Arg Leu Tyr 1 5
55	(2)	INFORMATION FOR SEQ ID NO:15:
ر ر		(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 9 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear
60		(ii) MOLECULE TYPE: peptide

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:
           Thr Gln Asp Ser Trp Glu Arg Leu Tyr
  5
      (2) INFORMATION FOR SEQ ID NO:16:
           (i) SEQUENCE CHARACTERISTICS:
                (A) LENGTH: 9 amino acids
10
                (B) TYPE: amino acid
                (D) TOPOLOGY: linear
         (ii) MOLECULE TYPE: peptide
15
         (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:
          Thr Gln Asp Phe Trp Glu Arg Leu Tyr
20
     (2) INFORMATION FOR SEQ ID NO:17:
          (i) SEQUENCE CHARACTERISTICS:
25
               (A) LENGTH: 7 amino acids
               (B) TYPE: amino acid
               (D) TOPOLOGY: linear
         (ii) MOLECULE TYPE: peptide
30
         (xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:
35
          Thr Gln Asp Ser Trp Glu Arg
     (2) INFORMATION FOR SEQ ID NO:18:
40
          (i) SEQUENCE CHARACTERISTICS:
               (A) LENGTH: 15 amino acids
               (B) TYPE: amino acid
               (D) TOPOLOGY: linear
45
         (ii) MOLECULE TYPE: peptide
         (xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:
50
          Gly Ser Thr Asp Asp Asn Arg Ser Pro Gln Ser Asp Pro Tyr Asn
          1
                                               10
     (2) INFORMATION FOR SEQ ID NO:19:
55
          (i) SEQUENCE CHARACTERISTICS:
              (A) LENGTH: 10 amino acids
               (B) TYPE: amino acid
               (D) TOPOLOGY: linear
60
         (ii) MOLECULE TYPE: peptide
```

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		(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:	
_		Ser Pro Lys Glu Asn His Gly Asp Phe Ile 1 5 10	
5	(2)	INFORMATION FOR SEQ ID NO:20:	
10		(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 9 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear	
15		(ii) MOLECULE TYPE: peptide	
		(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:	
20		Pro Asn His Asn Gly Ser Arg Gly Asn 1 5	
	(2)	INFORMATION FOR SEQ ID NO:21:	
25		(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 15 amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear	
30		(ii) MOLECULE TYPE: peptide	
		(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:	
35		Arg Leu Leu Arg Val Phe Lys Leu Ala Lys Ser Trp Pro Thr Leu 1 5 10 15	
	(2)	INFORMATION FOR SEQ ID NO:22:	
40		 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 21 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
45		(ii) MOLECULE TYPE: cDNA	
50		(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:	
	GCT'	TGCTGCG GGTCTTCAAG C	21
55	(2)	INFORMATION FOR SEQ ID NO:23:	
		 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 14 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear 	
60		(ii) MOLECULE TYPE: peptide	

	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:	
_	Leu Arg Ala Leu Pro Leu Arg Ala Leu Ser Arg Phe Glu Gly 1 5 10	
5	(2) INFORMATION FOR SEQ ID NO:24:	
10	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 21 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
15	(ii) MOLECULE TYPE: cDNA	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:	
20	ATCGAGACAG AGCCCGCAGC G	21
	(2) INFORMATION FOR SEQ ID NO:25:	
25	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 44 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
30	(ii) MOLECULE TYPE: cDNA	
35	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:	
	ACGGGTGCCG CAAGGACGGC GTCTCCGTGT GGAACGGCGA GAAG	44
	(2) INFORMATION FOR SEQ ID NO:26:	
40	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 45 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
45	(ii) MOLECULE TYPE: cDNA	
50	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:	
	GGCTATCCTT CCTCTTCCAG CTCTCACCCA GGTATGGAGC CAGGT	45
55	(2) INFORMATION FOR SEQ ID NO:27:	
	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 21 base pairs(B) TYPE: nucleic acid	
60	(C) STRANDEDNESS: single (D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: cDNA	

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	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:	
	TCCCGTACGC TGCAGCTCTT T	21
5	(2) INFORMATION FOR SEQ ID NO:28:	
10	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 15 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
15	(ii) MOLECULE TYPE: cDNA	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:	
20	CCCGGGGAAG GCTAC	15
	(2) INFORMATION FOR SEQ ID NO:29:	
25	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 15 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
30	(ii) MOLECULE TYPE: cDNA	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:	
35	GTCGACACCA GAAAT	15
	(2) INFORMATION FOR SEQ ID NO:30:	
40	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 30 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
45	(ii) MOLECULE TYPE: CDNA	
50	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:	30
	GGATCCTCTA GAGTCGACCT GCAGAAGGAA	30

(2) INFORMATION FOR SEQ ID NO:	(2)	INFORMATION	FOR	SEQ	ID	NO:31
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- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 24 base pairs
- 5 (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: cDNA

10

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:
- 15 TGACGCAGGA CTCCTGGGAG CGCC

24

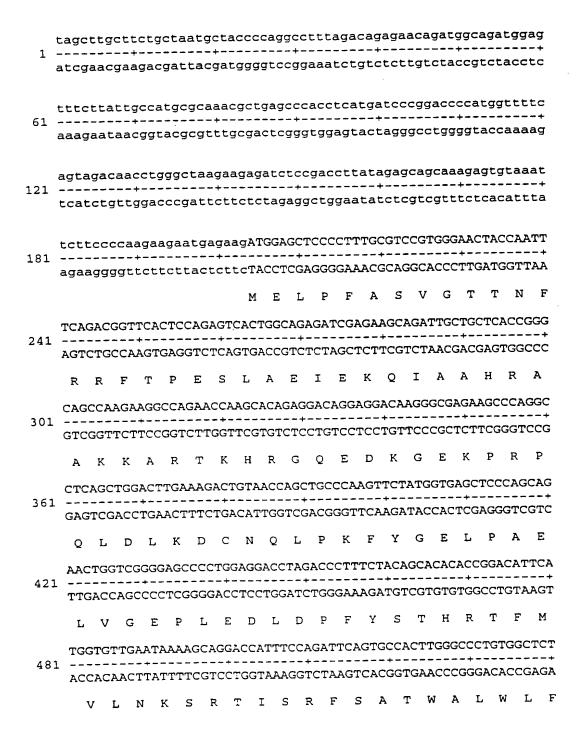
CLAIMS

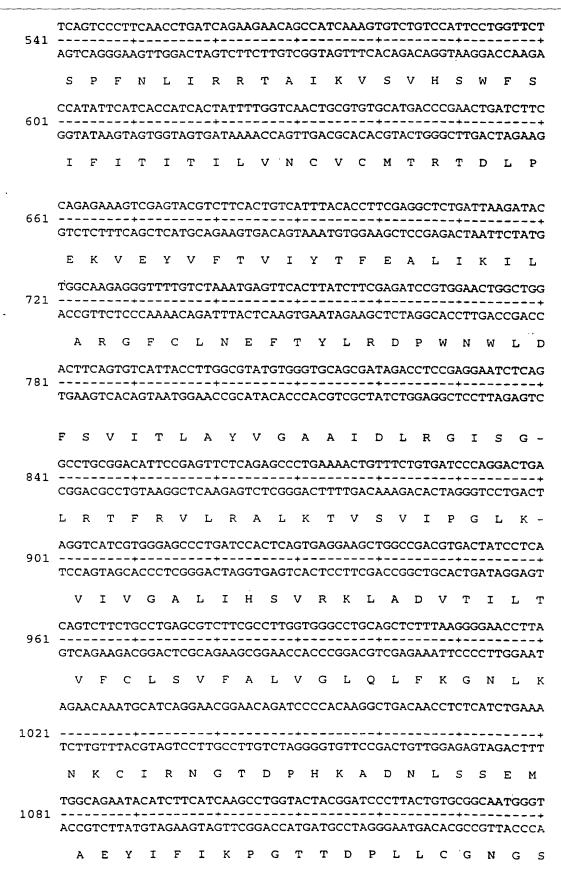
- 1. A mammalian sensory neuron sodium channel protein, wherein the sodium channel is insensitive to tetrodotoxin.
- 2. The sodium channel protein of claim 1 wherein said protein is derived from dorsal root ganglia.
- 3. The sodium channel protein of claim 2 wherein the sodium channel protein is a rat protein.
- 4. The sodium channel protein of claim 2 wherein the sodium channel protein is a human protein.
- 5. The sodium channel protein of claim 3 wherein said protein comprises the amino acid sequence shown in SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6 or SEQ ID NO: 8.
- 6. The sodium channel protein of claim 5 wherein said protein comprises the amino acid sequence of SEQ ID NO:2.
- 7. The sodium channel protein of claim 3 wherein said protein comprises the amino acid sequence encoded by the insert deposited in NCIMB deposit number 40744.
- 8. A nucleic acid sequence encoding the sodium channel protein of claims 1-7 or a complementary strand thereof.
- 9. The nucleic acid sequence of claim 8 wherein said nucleic acid sequence comprises the coding portion of the nucleic acid sequence shown in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5 or SEQ ID NO:7.
- 10. The nucleic acid sequence of claim 9 wherein said nucleic acid sequence comprises the coding portion of the nucleic acid sequence shown in SEQ ID NO:1.
 - 11. The nucleic acid that hybridizes to strand of claim 8 or claim 10.
- 12. A nucleic acid sequence encoding rat dorsal root ganglias sodium channel protein which comprises the sequence of the coding portion of the insert deposited in NCIMB deposit number 40744 or a complementary strand thereof.
 - 13. A vector comprising a nucleic acid sequence of claims 8-12.
- 14. A host cell transformed or transfected with a nucleic acid sequence of claims 8-12.

- anglion sodium channel, which channel is insensitive to tetrodotoxin, comprising contacting a test compound with said channel and detecting the activity of said channel.
 - 16. An antibody specific for the sodium channel protein of claim 1.
- 17. A nucleic acid sequence encoding the sodium channel protein of claims 1-7.
- 18. An expression vector comprising a nucleic acid sequence as defined in claim 12.
 - 19. A host cell comprising an expression vector as defined in claim 18.
- 20. A method of making a sodium channel protein as defined in any one of claims 1 to 7 which comprises culture of a host cell as defined in claim 19 under conditions suitable for expression of the sodium channel protein and optionally purifying the expressed sodium channel protein.

Figure la

Nucleic acid and amino acid sequence of TTXi DRG sodium channel





1111	CT	GATO	GCTO	GTO	CAC	rgc	CCT	GGA	.GGC	TAT	GTC	TGC	CTG	AAA	ACT	CCT	GAC	\AC(CCGC	ATT
1141							+				+			_+						TAA
	D	A	G	Н	С	P	G	G	Y	V	С	L	K	Т	P	D	N	P	D	F
1201	TT	ACT	ACA	CCA	GC1	TTT	GAT	TCC'	TTT	GCG'	TGG	GCA:	rtco	CTC	CAC	TGT	rrcc	GCC	TCA	TGA
		TGA	TGT	GGT	'CGA	AA	CTA	AGG	AAA	CGC	ACC	CGT	AAGO	GAG.	GTC	ACA	LAGO	CGG	AGT	ACT
	N	Y	T	s	F	D	s	F	A	W	A	F	L	s	L	F	R	L	М	т
	CGC	AGG	ACT	CCT	GGG	AGC	CGCC	TGI	raco	CAGO	CAGA	ACAC	TCC	:GGG	CTT	CTG	GGA	AAA	TGT.	ACA
1261				+			+-				+ -			+			-+-		ACA	+
	Q	D	s	W	E	R						L							Y	
	TGG	TCT	TTT	TCG	TGC	TGG	TTA	TTT	TCC	TTC	GAI	CGT	TCT	'ACC	TGG	TCA	Aጥጥ	ТСА	ייט טיי	TCC
1321				+			-+-		- -	+				+			-+-		AGA	+
	v	F																	L	
1201	CCG	TGG	TCA	CCA'	TGG	CGT	'ATG	AAG	AGC	CAGA	.GCC	AGG	CAA	CAA	TTG	CAG	AAA	TCG	AAG	CA
1381	GGC	ACC.	AGT	+ GGT	ACC	GCA	TAC	TTC	TCC	+ FTCT	CGG	TCC	 GTT	+ GTT.	 AAC	 GTC	-+- TTT.	AGC'	TTC	+ 3GT
	v	v	T	M	A	Y	E	E	Q	s	Q	Α	T	I	Α	E	I	E	A	ĸ
1 4 4 1	AGG.	AAA	AAA	AGT'	TCC.	AGG	AAG	ccc	TTG	AGG	TGC	TGC	AGA.	AGG.	AAC.	AGG.	AGG'	rgc'	rgg	CAG
1441	TCC	TTT'	r r t'	TCA	AGG	TCC	TTC	GGG	AAC	TCC	ACG	ACG	TCT	TCC'	TTG'	rcc'	TCC	ACG	ACC	TC
	E	K	К	F	Q	E	A	L	E	v	L	Q	K	E	Q	E	v	L	Α	A
1501	CCC	TGG	GGA	TTG2	ACA	CGA	CCT	CGC	TCC	AGT	CCC	ACA	GTG	GAT	CAC	CCT'	TAG	CCT	CCAA	AAA
	GGG.																	GGA	GGTI	TT
	L	G	I	D	Т	T	s	L	Q	s	Н	S	G	s	P	L	A	s	ĸ	N
1561	ACG	CCA	ATG!	AGA(GAA(GAC	CCA	GGG	TGA	AAT +	CAA	GGG'	TGT	CAG	AGG	GCT	CCAC	GG	\TGA	CA
	TGC	GGT:	rac:	rcto	CTT	CTG	GGT	CCC	ACT	TTA	GTT.	CCC	ACA	GTC:	rcc	CGAC	GTC	CCT	raci	GT
	A	N	E	R	R	P	R	V	K	S	R	V	S	E	G	S	Т	D	D	N
1621				+			-+-			+				+			-+			-+.
	TGT																			
																			S	
1681				+			-+-			+				+			-+			-+
	GTC																			
	ی	ĸ	ĸ	ĸ	Α	S	H	G	S	V	F	Н	F	R	Δ	P	9	Ω	ח	T

1741	TCT	CAT	TTC	CTG	ACG	GGA	TCA	CCC	CTC	ATO	ATO	GGG	TCI	TTC	CACC	GAC	ACC	AGG	AAA	GCC
2.42										-	TAC	ccc	AGA	AAG	TGC	CTC	TGC	TCC	TTT	CGG
	s	F	P	D	G	I	т	P	D	D	G	v	F	Н	G	D	Q	E	s	R
1801	GTC	GAG	GTT	CCA	TAT	TGC	TGG	GCA	'GGG	GTG	CTG	GGC	AGA	.CAG	GTC	CAC	TCC	CCA	.GGA	.GCC
1001	CAG	CTC	CAA	GGT														GGT	CCT	+ CGG
	R	G	s	I	L	L	G	R	G	A	G	Q	T	G	P	L	P	R	s	P
1061	CAC	TGC	CTC	AGT	ccc	CCA	LACC	CTC	GCC	GTA	GAC	ATC	GAG	AAG	AGG	GAC	AGC	TCC	GAG	TGC
1861	GTG	ACG	GAG	TCA	.GGG	GGI	TGG	GAC	CGG	CAT	CTG	TAC	CTC	+ TTC	TCC	CTC	TCG	AGC	CTC	+ ACG
	L	P	Q	s	P	N	P	G	R	R	Н	G	E	E	G	Q	L	G	v	P
1921	CCA	CTG	GTG	AGC	TTA	.CCG	CTG	GAG	CGC	CTG	AAG	GCC	:CGG	CAC	TCG	ACA	.CTA	CAG	GGC	AGA
1721	GGT	GAC	CAC	TCG	AAT	GGC	GAC	CTC	GCG	GAC	TTC	CGG	GCC	+ GTG	AGC	TGT	GAT	GTC	 CCG	+ TCT
	т	G	E	L	T	Α	G	Α	P	E	G	P	A	L	D	T	т	G	Q	к
1981	AGA	GCT	TCC	TGT +	CTG	CGG	GCT	ACT	TGA	ACG	AAC	CTT	TCC	GAG	CAC	AGA	GGG	CCA	TGA	GCG
	TCT	CGA	AGG.	ACA	GAC	GCC	CGA	TGA	ACT	TGC	TTG	GAA	AGG	CTC	GTG	TCT	ccc	GGT	ACT	CGC
	S	F	L	S	A	G	Y	L	N	E	P	F	R	A	Q	R	A	M	s	v
2041				+			-+-		 -	+				+			-+-			
	AAC.	AGT	CAT.	AGT.	ACT	GAA	GAC	AGT.	AAC	TCC	TCG.	AAC	TTC'	TCA	GAT'	TCG.	ACT	TCA	CGG	GTG
	V	_	Ι	M	T		V	_	E	Е	L	E	Ε	S	K	L	K	С	P	P
2101	CCT			+			-+-			+				+			-+-			+
	GGA					AGC	GAG'	TCT'	TCA'	TAG	ACT.	AGA	ccc	rca(CGA	CGG	GGT'	rca(CCT	CCT
		L	_	s 	F	A 	Q	K		L	I	W	E	С	С	P	K	W	R	K
2161	AGT			+			-+-			+				+ .			-+-			+
	TCA																			
													F							
2221	TCTC			+ -			-+-			+-				 -			-+			-+
													H							
	TCG																			
2281	AGC			+			-+-			+-							-+		. _	
	D	Α	М	L	Q	A	G	N	I	v	F	т	v	F	F	т	М	E	м	A

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5/17

2341	CCTTCAAGATCATTGCCTTCGACCCCTACTATTACTTCCAGAAGAAGTGGAATATCTTCC
2741	GGAAGTTCTAGTAACGGAAGCTGGGGATGATAATGAAGGTCTTCTTCACCTTATAGAAGC
	F K I I A F D P Y Y Y F Q K K W N I F D
2401	ACTGTGTCATCGTCACCGTGAGCCTTCTGGAGCTGAGTGCATCCAAGAAGGGCAGCCTGT
2401	TGACACAGTAGCAGTGGCACTCGGAAGACCTCGACTCACGTAGGTTCTTCCCGTCGGACA
	C V I V T V S L L E L S A S K K G S L S
	CTGTGCTCCGTTCCTTACGCTTGCTGCGGGTCTTCAAGCTGGCCAAGTCCTGGCCCACCC
2461	GACACGAGGCAAGGAATGCGAACGACGCCCAGAAGTTCGACCGGTTCAGGACCGGGTGGG
	V L R S L R L L R V F K L A K S W P T L
	TGAACACCCTCATCAAGATCATCGGGAACTCAGTGGGGGCCCTGGGCAACCTGACCTTTA
2521	
	N T L I K I I G N S V G A L G N L T F I
	TCCTGGCCATCATCGTCTTCATCTTCGCCCTGGTCGGAAAGCAGCTTCTCTCAGAGGACT
2581	AGGACCGGTAGTAGCAGAAGTAGAAGCGGGACCAGCCTTTCGTCGAAGAGAGTCTCCTGA
	LAIIVFIFALVGKQLLSEDY
	ACGGGTGCCGCAAGGACGGCGTCTCCGTGTGGAACGGCGAGAAGCTCCGCTGGCACATGT
2641	TGCCCACGGCGTTCCTGCCGCAGAGGCACACCTTGCCGCTCTTCGAGGCGACCGTGTACA
	G C R K D G V S V W N G E K L R W H M C
2701	GTGACTTCTTCCATTCCTTGCTGGTCGTCTTCCGAATCCTCTGCGGGGAGTGGATCGAGA
2701	CACTGAAGAAGGTAAGGAAGGACCAGCAGAAGGCTTAGGAGACGCCCCTCACCTAGCTCT
	D F F H S F L V V F R I L C G E W I E N
2761	ACATGTGGGTCTGCATGGAGGTCAGCCAGAAATCCATCTGCCTCATCCTCTTTGACTG
2.01	TGTACACCCAGACGTACCTCCAGTCGGTCTTTAGGTAGACGGAGTAGGAGAAGAACTGAC
	M W V C M E V S Q K S I C L I L F L T V
2821	TGATGGTGCTGGGCAACCTAGTGGTGCTCAACCTTTTCATCGCTTTACTGCTGAACTCCT
	ACTACCACGACCCGTTGGATCACCACGAGTTGGAAAAGTAGCGAAATGACGACTTGAGGA
	M V L G N L V V L N L F I A L L N S F
2881	TCAGCGCGGACAACCTCACGGCTCCAGAGGATGACGGGGAGGTGAACAACTTGCAGTTAG
	AGTCGCGCCTGTTGGAGTGCCGAGGTCTCCTACTGCCCCTCCACTTGTTGAACGTCAATC
	S A D N L T A P E D D G E V N N I. O I. A

29/1	CA	_TGC	SCCA	AGGZ	ATC	CAG	GTA(CTT	GGC	CAT	CGG	3CC2	AGC	AGG	GCC.	ATC	GCC.	AGT	TAC.	ATC.
2741											GCC									
	L	A	R	I	Q	V	L	G	H	R	A	S	R	A	I	A	s	Y	I	s
3001	GC2	AGCC	ACI	rgco	GAT	rtc	CGC'	rgg(ccc	AAG	GTGC	SAG	ACC	CAG	CTG	GGC.	ATG.	AAG	ccc	CCA
											CACC									GGT
	S	Н	С	R	F	R	W	P	K	V	E	T	Q	L	G	M	K	P	P	L
3061	TCA	CCA	GCT	CAG	AGG	CCA	AGA	ACC	ACA	TTG	CCA	CTG	ATG	CTG	TCA	GTG	CTC	CAC	STGO	GGA
	AGT	GGT	CGA	GTC	TCC	GGT	TCI	'TGG	TGI	'AAC	GGT	GAC	TAC	GAC	AGT	CAC	GAC	GTC	ACC	CCI
	T	S	S	E	A	K	N	Н	I	A	Т	D.	A	v	s	A	A	v	G	N
3121				+			-+-			+	AGG			+			-+-			
	TGG	ACT(GTT'	TCG	GTC	GAG	AGT	CAT	CGG	GGT	TCC'	TCT	TAG	TGC	CCC	TGA	AGT	'AGT	'GAC	TAG
											E							_		P
3181	CCA.			+			-+-			+				+			-+-			
	GGT															AGC	TGC	TCG	AGC	TCC
	N	V	W	٧	S	V	P	I	A	E	G	Ε	S	D	L	D	E	Ļ	E	E
3241	AAG.	ATA:	rgg/	AGC	AGG	CTT	CGC.	AGA	GCT	CCT	GGC	AGG	AAG	AGG	ACC	CCA	AGG	GAC	AGC	AGG
	TTC	TATA	ACC!	rcg	TCC	GAA	GCG	TCT	CGA	GGA	CCG!	rcc	rrc	rcc'	TGG	GGT	TCC	CTG	TCG	TCC
	D	М	E	Q	A	S	Q	S	s	W	Q	Ε	E	D	P	K	G	Q	Q	E
3301	AGC						-+-			+				+ -			-+-			+
	TCG												rcc	GTC	GT(CTT	CGG	GTC	GGA	GGT
											н	_	A						_	
3361	TGA		+				-+-			+-							-+-			+
	ACT																			
	CTC										G									
3421	GAG		+				-+-			+-							- +			+
			.000	,,,,,	JUC .		- 1 - 2		inc.	1610	3C 1 C	GAC	SGAC	MC I		-GT	-GT	JCC ₄	ACC'	rga
											S									
3481			+				-+-			+-	-		+				- +		 -	+
	CGG	CCI	GGC	TCT	rcci	rtti	AGG/	ACTO	CT	rct <i>i</i>	\GG@	GCI	CGA	CCC	TCI	raci	rgg <i>i</i>	ACC:	rgc:	rcg

3541				+			-+-			+	· -			+			-+-		ATA	+
	D	D	С	F	т	E	G	С	т	R	R		P	С		N	v		T	s
3601	GCA	AGT	CTC	CTT	GGG	CCA	CAG												TGG	
	CGT	TCA	GAG	GAA	.ccc	GGT	GTC												ACC	
	K	s	P	W	A	Т	G	W	Q	V	R	K	T	С	Y	R	I	v	E	Н
3661	ACA	GCT	GGT	TTG	AGA	GTT	TCA	TCA	TCI	TCA	TGA	TCC	TGC	TCA	GCA	GTG	GAG	CGC	TGG	CCT
																			ACC	+ GGA
	s	W	F	E	S	F	I	I	F	М	I	L	L	s	s	G	A	L	A	F
3721	TTG	AGG.	ATA.										AGT		TGC	TGG	AGT	ACA	CTG	ACC
	AAC	TCC'	TAT'	TGA	TGG	ACC	TTC	TCT	TTG	GGG	CTC	ACT	TCA	GGC	ACG	ACC	TCA	TGT	GAC	TGG
	E	D	N	Y	L	E	Ε	K	P	R	V	K	s	V	L	Ε	Y	Т	D	R
3781	GAG			+			-+-			+				+			-+-			+
	CTC.	F															GGA	TAC	CGA	AGT
			Т	F	I 	F 	v 	F 	E 	М	L	L	K	W	٧	A	Y	G	F	K
3841	AAA			+			-+-			+				+			-+-			+
	TTT					TAC	GGA	CCA	CGA	CCG	ACC'	TGA	AGG.	AGT.	AAC.	ACT'	TGT.	AGA	GGG	ACT
	К	Y	F	Т	N	Α	W	С	W	L	D	F	L	I	V	N	I	S	L	Т
3901	CAA	GCC:	rca:	rag +	CGA.	AGA	TCC' -+-	TTG. - - -	AGT.	ATT +	CCG.	ACG	TGG		CCA			ccc'	TTC	GGA
	GTT	CGG?	AGTA	ATC	GCT'	TCT.	AGG.	AAC	TCA'	TAA	GGC'	TGC.	ACC	GCA	GGT.	AGT'	TTC	GGG.	AAG	CT
	s	L	I	Α	K	I	L	Ε	Y	s	D.	V	A	S	I	K	A	L	R	T
3961	CTC'	rcc	TGC	CC'	rcc	GAC	CGC'	TGC	GGG	CTC'	TGT	CTC	GAT	rcg.	AAG	GCA'	TGA(GGG'	ragi	rgg
	GAG	AGG	CACC	GG2	AGG	CTG	GCG.	ACG	CCC	GAG.	ACA	GAG	CTA	AGC'	TTC	CGT	ACT	CCC	ATC	vcc.
	L	R	A	L	R	P	L	R	A	L	s	R	F	Ε	G	М	Ŕ	V	V	V
4021	TGG	ATG	CCI	rcg:	rgg	GCG	CCA'	rcc	CCT	CCA'	TCA'	rga.	ACG	rcc'	rcc	rcg:	CTO	GCC:	rcai	CT
	ACC'	raco	GGG <i>I</i>	AGC	ACC	CGC	GGT	AGG	GGA(GGT.	AGT	ACT	rgc	, AGG2	AGG	AGC	AGA	CGG2	AGTA	\GA
	D	A	L	V	G	A	I	P	s	I	М	N	v	L	L	V	С	L	I	F
4081	TCT	GGC1	CAT	CT	rca(GCA'	TCA'	rgg(GCG'	rga:	e CC:	rct:	TCG	CCG	GGA/	AAT	rtto	CGA2	AGTO	CG
	AGA	CCG	\GT <i>I</i>	AGA	AGT	CGT.	AGT	ACC	CGC	ACT'	i GGZ	AGA	AGC	GGC	CCT	rta.	AAA	GCT?	CAC	GC
	W	L	I	F	s	I	М	G	v	N	L	F	Α	G	ĸ	F	s	К	С	v

4141	TCC	ACA	CCA	GA.	ATA	ACC	CAT	TTI	rcc/	AACC	STGA	LTA	CGA	CGA	TGG	TGA	ATA	ACA	AGI	CCG
3737	AGC	TGT	'GGT	CTI	TAT	TGC	GTA	AAA	\GG!	rtgo	CACT	TAA	GCI	GCI	'ACC	ACT	TAT	TGT	TCA	GGC
	D	т	R	N	N	P	F	s	N	v	N	s	Т	М	v	N	N	ĸ	s	E
	AGI	GTC	ACA	ATC	AAA	ACA	.GCA	,CCC	GC	CACT	rtci	TCI	'GGG	TCA	ACG	TCA	AAG	TCA	ACT	TCG
4201	TCA	 CAG	TGI	TAC	TTT	TGI	CGI	GGC	CGC	GTG <i>I</i>	LAGA	AGA		+	TGC	AGT	-+- TTC	AGT	TGA	+ AGC
	С	н	N	Q	N	s	т	G	н	F	F	W	v	N	v	к	v	N	F	D
																	•		-	_
4261				+			-+-							+			-+-		GGA	+
	TGT	TGC	AGC	GAT	ACC	CGA	TGG	AGC	GTO	GAAC	AAG	TCC	ACC	GTT	'GGA	AGT	TTC	CGA	CCT	ACC
	N	V	A	М	G	Y	L	A	L	L	Q	v	A	T	F	K	G	W	М	D
4321		TAA			CAG	CTC	TTG	ITA	CCC	GAC	SAGA	TCA	ACA	GTC	AGC	CTA	ACT	GGG	AGA	ACA
4721					GTC	GAC	AAC	TAA	GGC	CTC	TCT	AGT	TGT	CAG	TCG	GAT	-+- TGA	ccc	TCT	+ TGT
	I	M	Y	A	Α	v	D	s	G	Ε	I	N	s	Q	P	N	W	E	N	N
	ACT	TGT	ACA	TGT	'ACC	TGT	'ACT	TCG	TCC	TTT	TCA	TCA	TTT	TCG	GTG	GCT	TCT	TCA	CGC	TGA
4381	TGA	 ACA	TGT	+ ACA	TGG	ACA	-+- TGA	AGC	AGC	AAA	AGT	AGT	 AAA	+~- AGC	CAC	 CGA	-+- AGA	 AGT	 GCG	+ ACT
	L	Y	М	Y	L	Y	F	v	v	F	I	I	F	G	G	F	F	T	L	N
	ATC	TCT	TTG	TTG	GGG	TCA	TAA	TCG	ACA	ACT	TCA	ACC	AAC	AGA.	AAA	AAA.	AGC'	ጥልር፡	GAG	300
4441				+			-+-			+				+			-+-		CTC	+
	L	F	v	G	v	I	ī	D	N	F	N	0	0	ĸ	ĸ	ĸ				
	_	_	•	_			_	_				_	_				L	G 	G TGG(Q
4501				+			-+-			+				+			-+-			+
																ACT"	rct'	rcg.	ACC	CGA
	D	Ι	F	М	_			_			_	_	-			K	K	L	G	S
4561	CCA			+			-+-			+				+			-+-			+
	GGT																			
	K	K	P	Q	K	P	I	P	R	P	L	N	K	Y	Q	G	F	V	F	D
4621	ACA	TCG'	TGA	CCA +	GGC	AAG	CCT -+-	TTG 	ACA	TCA	TCA	TCA'	TGGʻ	TTC	rca:	CTC	GCC	rca.	ACAT	rga
	TGT.																			
	I	V	Т	R	Q	A	F	D	I	I	I	M	v	L	I	С	L	N	M	I
4681	TCA	CCA'	TGA	TGG	TGG	AGA	CCG.	ACG	AGC	AGG	GCG.	AGG	AGA	AGAC	CGA/	AGG:	rtc:	rgg	GCAC	AA
4001	AGT																			
	T	м	М	v	E	т	D	E	Q	G	E	E	к	т	к	v	L	G	R	I

474	. 1	TC	AAC	CAG	TTC	TTT	GTG	GCC	GT	CTT	CAC	GG	GC	SAG	TG	TGI	rga:	TGA	AG!	TGI	TTC	ccc	TGC
-,-	-							+				+											TGC + ACG
		N	Q	F	F	V	A	. v	F		r	G	E	С	•	V	M	ĸ	М	F	A	L	R
480	1	GAC	AG	rac'	TAC	TTC	ACC	AAC	GGC	TG	GAA	.CG	rg1	TC	GA	CTI	'CA	rag	TGG	TGA	TCC	TGT	CCA
		CTG	TC	ATG	ATG	AAG	TGG	TTG	CCG	ACC	CTT	GC	ACA	AG	CT	GAA	GT?	ATC.	ACC	ACI	'AGG	aca	+ GGT
		Q		7 5	Y 1	F '	T I	N	G	W	N	V	F	' 1	D	F	I	v	V	I	L	s	ı
40.0		TTG	GGA	GTO	TG	TG:	rtt'	rcto	GCA	ATC	CT"	TAA	GT	CAG	CTO	GA	AAA	יכייט	ىتى ت	ጥርጥ	ccc		200
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		AGA.																					
		F						A										L	I	R	A	A	ĸ
4981		AGGG	GA'	TTC	GCA +	CGC	TGC	TCT	TCC	SCC	CTC	ATO	GAI	GT	'CC	CTC	CC	CGC	CCI	'CTI	CAA	CAT	CG
	J	יכככ	CT	AAG	CGT	GCG	ACG	AGA	AGC	GGG	GAG	TA	CTA	CA	.GG	GAC	GG	GCG	GGA	+ GAA	GTT	GTA	-+ GC
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5041		CCT										+											
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5101		CGT																					
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5341	TCA'	rcgʻ	IGG:	rca.	ACA'	TGT	ACA	TCG	CAG	TGA	TTC'	TGG.	AGA.	ACT'	TCA	ACG	TAG	CCA	CCG	AGG
	AGT	AGC	ACC	AGT"	TGT	ACA	TGT.	AGC	GTC.	ACT.	AAG.	ACC	TCT	TGA.	AGT	TGC.	ÀТС	ggt	GGC'	rcc
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5401	AGA	GCA	CGGZ	AGC	ccc	TGA	GCG.				TCG.	ACA'	TGT'	TCT	ATG.	AGA	CCT	GGG.	AGA	AGT
2401	TCT	CGT	GCC	rcg	GGG/	ACT	CGC		TGC'		AGC'	TGT.	ACA	AGA'	TAC	TCT	GGA	CCC	TCT:	rca
	s	T	E	P	L	s	Ε	D	D	F	D	M	F	Y	E	Т	W	E	K	F
- 4 - 4	TCG	ACC	CGG	AGG(CAC	CCC.	AGT'	TCA'	rtg(CT'	TTT(CTG	ccc'	rct	CAG	ACT'	rcg	CGG	ACA	CGC
5461	AGC'	rgg	GCC1	rcc	GGT	GGG'	TCA.	AGT	AAC	GGA.	AAA	GAC	GGG	+	GTC'	TGA	AGC	GCC'	rgt	+ GCG
	D	P	E	A	T	Q	F	I	A	F	s	A	L	s	D	F	A	D	T	L
5521	TCT	CCG	GCC	CTC	TA(GAA	TCC	CCA	AAC	CA	ACC	AGA	ATA:	rat:	'AA'	rcc	AGA'	TGG/	ACCI	rgc
JJ21	AGA	GCC	CGGC	GAG	AAT	CTT.	AGG	GGT'	rtg	GT'	rgg:	rct'	[AT	ATA	ATT	AGG'	rct.	ACC'	rgg/	\CG
	s	G	P	L	R	I	P	ĸ	P	N	Q	N	I	L	I	Q	M	D	L	P
5581	CGT	rgg'	rcc	CCG	GGZ	ATA	AGA'	TCC	ACT	GTC'	TGG/	ACA:	rcc:	rtt:	rtg	CCT	rca(CAA	AGA	CG
3301	GCA	ACC	AGG	GC	CCC.	rat	TCT.	AGG'	rga(CAG	ACC:	rgt/	AGGZ	LAA.	AAC	GGA/	AGT	GTT.	CTI	GC
	L	v	P	G	D	ĸ	I	Н	С	L	D	I	L	F	A	F	T	K	N	v
5641	TCT	rgg	GAG!	AAT	CCG	GGG.	AGT	TGG	ACT(CCC'	rga.	AGA	CCA	ATAT	rggi	AAG/	\GA	AGT:	rati	rgg
	AGA	ACC	CTC	rta(GGC	CCC	TCA	ACC'	rga	GG	ACT!	rcto	GT7	TAT	ACC:	rrc:	rct'	rca.	ATA	CC
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5701	CGA	CCA	ATC:	rct(CCA	AAG	CAT	CCT	ATG	AAC	CAA	rag	CAC	CAC	CC.	rcco	GT	GA.	AGCA	.GG
3,01	GCT	GGT'	raga	AGA	GGT'	TTC	GTA	GGA'	TAC'	TTG	GTT?	ATC	GTC	GTC	GG2	AGG	CA	CT:	rcgi	CC
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5761	AAG																			
	TTC																			
	D	L	S	A	T	v	I	Q	ĸ	A	Y	R	s	Y	M	L	Н	R	s	L
5821	TGA																			-
	ACT	GTG.	AGA(GGT	TGT	GGG	ACG	TAC.	ACG	GGT	ccc	GAC'	rcc:	rcc:	PAC	CGC	ACA	GTG/	AAGC	GC
	T	L	S	N	T	L	Н	V	P	R	A	E	Ε	Đ	G	V	S	L	P	G
5881	GGG																			
	CCC									•										
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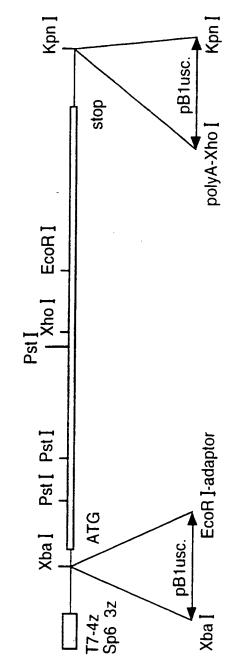
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5041	CTG	CTA																				
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6061	GCC	CTG	GAC	TC:	AGT	GAa	ggc -+-	act	cag	gca +	tg	cac	cag	ggc	agg	gtt 	cca	atç	rtc	ttt	ct	c t - +
	CGG	GAC	CTG	GAG	rca(CTt	ccg	tga	gtc	cgt	acc	gtç	gtc	CCS	jtco	caaç	ggt	tac	aga	aaa	gag	ga
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6121	gct																					gt -+
	cga																					
6181	cac						-+-			+				+				-+-				-+
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	ctt																					
6301		tgct					-+-	-		+		. – -		+				-+-				-+
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6361	aca	tago	caaa	agad	ctti	ttc	tgc	tgg	tgt	cgg	gca	ıgt	ct	tag	aga	agt	ca	cgt	agg	3 99	ttç	1 9
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Fig.1b.

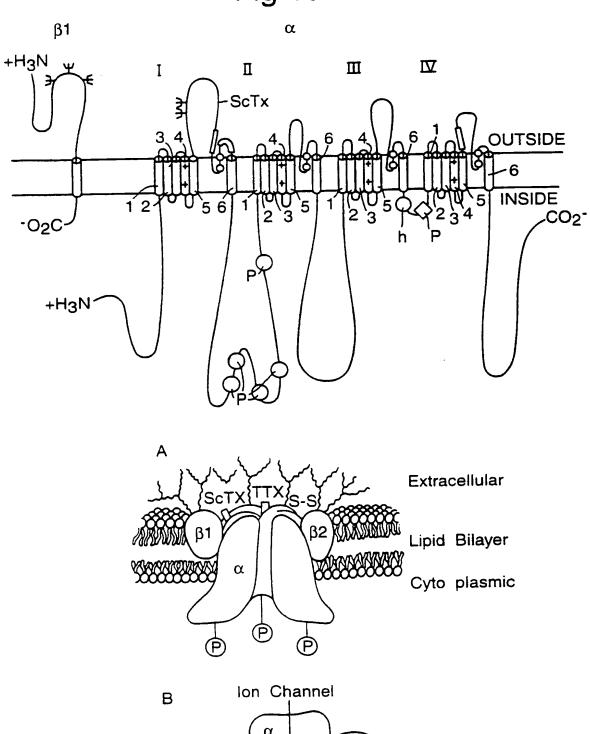
SNS-B voltage gated sodium channel PNC IB XOI-construct



Constructs were generated in pGem 3z and pGem 4z with bluescript polylinkers

Linearisation site is KPNI

Fig.1c.



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Sequence of PCR primers for isolation of human clone probes

- a) Highly conserved regions of all sodium channels
 - 1) Position 2475-2510 S4 Domain II

 Degenerate primers (20-24mers) encoding amino acid residues

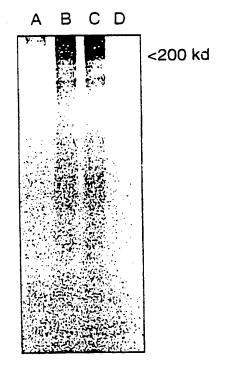
 RLLRVFKLAKSWPTL or non degenerate primers within this

 region e.g. 5' gcttgctgcgggtcttcaagc 3'
 - 2) Position 3961 4010 S4 Domain III

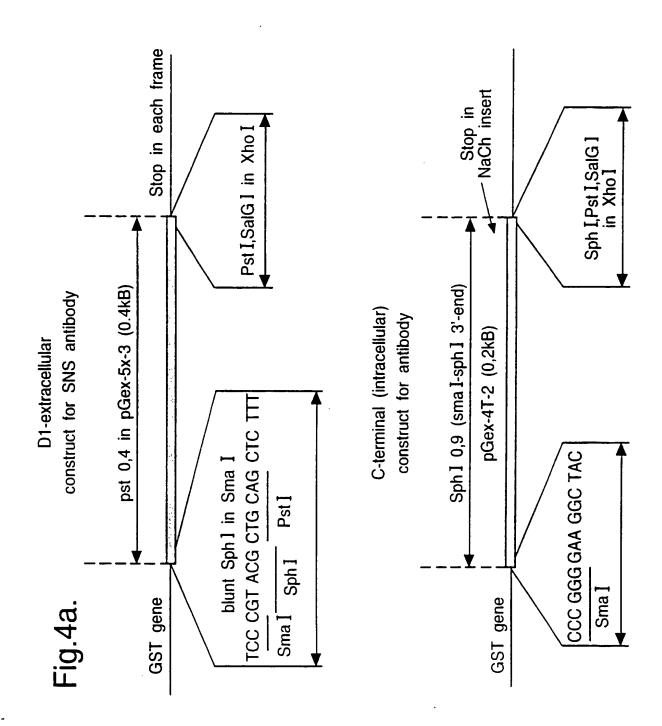
 Degenerate primers encoding the complementary strand encoding residues LRALPLRALSRFEG or non degenerate primers within this region e.g. 5' atcgagacagagccgcagcg 3'
- b) Unique sequence primers for SNS-homologues e.g. residues with the region 2641-2680
- e.g. 5' acgggtgccgcaaggacggcgtctccgtgtggaacggcgagaag 3' and complementary sequence within the region 3375 and 3420 e.g. 5' ggctatccttctctctccagctctcacccaggtatggagccaggt 3'

Fig.3.

In vitro synthesis of S-35 methionine labelled SNS-B voltage gated sodium channel in a coupled transcription/translation system

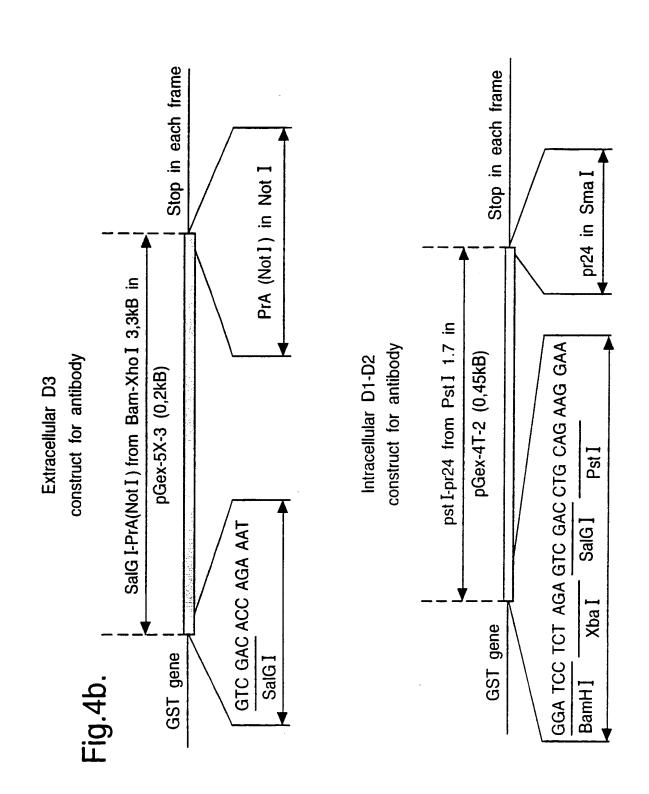


Autoradiograph of a 7.5% SDS polyacrylamide gel, showing the migration of labelled proteins compared to the sizes of known molecular weight markers (Amersham rainbow markers). Lane A control. Lane B SNS-B, Lane C SNS-B, Lane D control. The predicted 200kDa band representing the SNS-B sodium channel is arrowed.

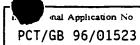


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	C12N3/ 10	•			
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Inter anal Application No

Contract	DOCUMENTS CONSIDERED TO BE BELLIANT	PCT/GB 96/01523
ategory *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	NEUROSCIENCE LETTERS, vol. 185, 1995, pages 70-73, XP002017245 J.B. ARBUCKLE AND R.J. DOCHERTY: "Expression of tetrodotoxin resistant sodium channels in capsaicin-sensitive	1,8,11, 13-20
	<pre>dorsal root ganglion neurons of adult rats" *see the whole article*</pre>	
X	BRAIN RESEARCH, vol. 639, 1994, pages 125-134, XP002017246 S. JEFTINIJA: "The role of tetrodotoxin-resistant sodium channels of small primary afferent fibers" *see the whole article*	1,8,11, 13-20
x	JOURNAL OF MEMBRANE BIOLOGY, vol. 116, 1990, pages 117-128, XP002017247 A. SCHWARTZ ET AL.: "Structural and developmental differences between three types of Na channels in dorsal root ganglion cells of newborn rats" *see the whole article*	1,8,11, 13-20
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